

[School of Health Professions](https://pearl.plymouth.ac.uk/hp-research) [Faculty of Health](https://pearl.plymouth.ac.uk/foh-research)

2024-08-21

Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study

Patricia Casas-Agustench

Jade M. Hayter

Odelia S. B. Ng

Lauren V. Hallewell

Nathaniel J. Clark

et al. See next page for additional authors

[Let us know how access to this document benefits you](https://forms.office.com/e/bejMzMGapB)

General rights

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Please cite only the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author. Take down policy

If you believe that this document breaches copyright please [contact the library](https://pearl.plymouth.ac.uk/about.html) providing details, and we will remove access to the work immediately and investigate your claim.

Follow this and additional works at: [https://pearl.plymouth.ac.uk/hp-research](https://pearl.plymouth.ac.uk/hp-research?utm_source=pearl.plymouth.ac.uk%2Fhp-research%2F529&utm_medium=PDF&utm_campaign=PDFCoverPages)

Recommended Citation

Casas-Agustench, P., Hayter, J., Ng, O., Hallewell, L., Clark, N., & Bescos, R. (2024) 'Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study', Foods, 13(16). Available at: <https://doi.org/10.3390/foods13162615>

This Article is brought to you for free and open access by the Faculty of Health at PEARL. It has been accepted for inclusion in School of Health Professions by an authorized administrator of PEARL. For more information, please contact openresearch@plymouth.ac.uk.

Authors

Patricia Casas-Agustench, Jade M. Hayter, Odelia S. B. Ng, Lauren V. Hallewell, Nathaniel J. Clark, and Raul Bescos

PEARL

Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study

Casas-Agustench, Patricia; Hayter, Jade M.; Ng, Odelia S. B.; Hallewell, Lauren V.; Clark, Nathaniel J.; Bescos, Raul

Published in: Foods

DOI: [10.3390/foods13162615](https://doi.org/10.3390/foods13162615)

Publication date: 2024

Document version: Publisher's PDF, also known as Version of record

Link: [Link to publication in PEARL](https://researchportal.plymouth.ac.uk/en/publications/12e8481e-c077-4df1-9ebe-ca8ded4b091f)

Citation for published version (APA):

Casas-Agustench, P., Hayter, J. M., Ng, O. S. B., Hallewell, L. V., Clark, N. J., & Bescos, R. (2024). Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study. Foods, 13(16), Article 2615. <https://doi.org/10.3390/foods13162615>

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Wherever possible please cite the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content

should be sought from the publisher or author.

Article

Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study

Patricia Casas-A[gus](https://orcid.org/0000-0002-3939-4743)tench * [,](https://orcid.org/0000-0003-4424-1087) Jade M. Hayter, Odelia S. B. Ng, Lauren V. Hallewell, Nathaniel J. Clark and Raul Bescos

> School of Health Professions, Faculty of Health, University of Plymouth, Plymouth PL4 6AB, UK; hayterjade94@gmail.com (J.M.H.); odeliang.ob2001@gmail.com (O.S.B.N.); nathaniel.clark@plymouth.ac.uk (N.J.C.); raul.bescos@plymouth.ac.uk (R.B.)

***** Correspondence: patricia.casas@plymouth.ac.uk; Tel.: +44-1752-588892

Abstract: Edible algae are a natural source of nutrients, including iodine, and can also contain nitrogen in the form of nitrate ($NO₃⁻$) and nitrite ($NO₂⁻$) as they can fix nitrogen from seawater. This study aimed to analyse the NO_3^- , NO_2^- , and iodine concentrations in eighteen macroalgae and five microalgae species commercially available in the United Kingdom. NO_3^- and NO_2^- concentrations were measured using high-performance liquid chromatography (HPLC), and iodine was determined using inductively coupled plasma mass spectrometry (ICP-MS). $NO₃⁻$ and iodine concentrations in macroalgae (NO₃⁻: 4050.13 ± 1925.01 mg/kg; iodine: 1925.01 ± 1455.80 mg/kg) were significantly higher than in microalgae species (NO₃⁻: 55.73 \pm 93.69 mg/kg; iodine: 17.61 \pm 34.87 mg/kg; $p < 0.001$ for both). In the macroalgae group, nori had the highest NO_3^- (17,191.33 \pm 980.89 mg/kg) and NO_2^- (3.64 \pm 2.38 mg/kg) content, as well as the highest iodine content. Among microalgae, *Dunaliella salina* had the highest concentration of $NO₃⁻$ (223.00 \pm 21.93 mg/kg) and iodine (79.97 \pm 0.76 mg/kg), while *Spirulina* had the highest concentration of NO₂⁻ (7.02 \pm 0.13 mg/kg). These results indicate that commercially available edible algae, particularly macroalgae species, could be a relevant dietary source of NO_3^- and iodine.

Keywords: edible algae; nitrate; nitrite; iodine

1. Introduction

Edible algae are photosynthetic aquatic organisms widely used in cuisines around the world for their nutritional value, which includes vitamins, minerals, antioxidants, and essential fatty acids [1]. These algae can be consumed as food or dietary supplements. They can be classified into macroalgae and microalgae. Macroalgae, commonly known as seaweeds, are multicellular algae and include examples such as kelp, nori, and dulse [2]. Microalgae are microscopic, unicellular, or multicellular algae, with *Spirulina* and *Chlorella* being notable examples [3].

This study focused on the nitrate (NO_3^-) content in edible algae as this anion is increasingly being considered a conditionally essential nutrient [4–6]. Dietary NO_3^- can enhance nitric oxide bioavailability, which is a key signaling molecule involved in many physiological processes, including cardiovascular regulation, neuronal signaling, and immune responses [4]. Additionally, dietary NO_3^- from green leafy vegetables or beetroot juice has a prebiotic effect on the oral microbiome, increasing the abundance of bacteria that can reduce the risk of periodontal disease [7–10]. This new evidence is shifting the traditional view on dietary $\overline{\rm NO_3}^-$ as a potential carcinogen [11]. Although an Acceptable Daily Intake (ADI) of 3.7 mg/kg/day of $NO₃⁻$ is still retained [12], the consumption of $NO₃$ ⁻-rich foods such as rocket, spinach, lettuce or beetroot can exceed the ADI levels [13]. However, recent evidence suggests a protective effect of plant-based $\mathrm{NO_3}^-$ against cancer and cardiovascular disease [14].

Citation: Casas-Agustench, P.; Hayter, J.M.; Ng, O.S.B.; Hallewell, L.V.; Clark, N.J.; Bescos, R. Nitrate, Nitrite, and Iodine Concentrations in Commercial Edible Algae: An Observational Study. *Foods* **2024**, *13*, 2615. [https://doi.org/10.3390/](https://doi.org/10.3390/foods13162615) [foods13162615](https://doi.org/10.3390/foods13162615)

Academic Editor: Yuhuan Liu

Received: 12 July 2024 Revised: 13 August 2024 Accepted: 15 August 2024 Published: 21 August 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

Current knowledge about the NO_3^- content in commercial edible algae is limited. A recent study by Martin-Leon et al. (2021) [15] reported substantial variation among different commercial algae species in Spain, with nori and kombu showing the highest concentrations of NO_3 ⁻ (± 3000 mg/kg). However, the analysis method used in this study lacked the sensitivity to measure $\rm NO_3^-$ at levels below 500 mg/kg. To address this limitation, the current study employed high-performance liquid chromatography (HPLC) with high sensitivity (up to 0.1 pmol) for the measurement of $NO₃⁻$ and nitrite ($NO₂⁻$) levels [16].

When NO_3^- is taken up by edible algae, it can be reduced to NO_2^- , which is then transported to the chloroplasts for reduction to ammonium [17]. Nitrogen incorporation is often rate-limited by the control of ammonium assimilation [17]. This is relevant because while NO_3^- is safe even at high doses, NO_2^- can cause serious harm at considerably lower levels [18]. Consequently, an ADI level of 0.07 mg/kg body weight/day of $\mathrm{NO_2}^$ was established by the European Food Safety Authority (EFSA) [19]. To the best of our knowledge, no previous study has investigated the $\rm NO_2^-$ concentration in commercial edible algae. This represents a significant gap in the literature that warrants further research.

In addition to NO_3^- and NO_2^- , edible algae can also serve as an important source of iodine. This is relevant because iodine can potentially interfere with NO_3^- uptake and its subsequent reduction to NO_2^- [20]. Moreover, iodine is essential for thyroid hormone synthesis, but an excess of iodine intake can also lead to thyroid gland dysfunction and goitre [21]. The recommended intake of iodine is 150 μ g/day for adults, with the tolerable upper intake level (UL) set at 600 µg/day by the Scientific Committee on Food (SCF) [22]. Following this, some manufacturers of edible algae recommend a maximum intake of 5 g/day to reduce the risk of iodine toxicity. However, most commercial edible algae do not provide information about their iodine concentration on the packaging. Current scientific data shows significant variation in the iodine concentration of commercial edible algae, ranging from 30 to 31,000 μ g/g [23].

Therefore, the main goal of this study was to analyse the NO_3^- , NO_2^- , and iodine concentrations in commercial macroalgae and microalgae available in the United Kingdom (UK). A secondary aim was to compare the NO_3^- and NO_2^- levels in these edible algae with the current ADI and UL levels for NO_3^- , NO_2^- , and iodine, respectively. We hypothesised that certain edible algae could contain substantial amounts of $\text{NO}_3^{\text{-}}$, potentially surpassing the levels found in terrestrial green leafy vegetables like rocket (6000–7000 mg/kg) or beetroot (3000–4000 mg/kg). Additionally, we also hypothesised that consuming standard portion sizes of edible algae (5 g/day) would not result in NO_3^- , NO_2^- , and iodine levels exceeding the ADI and UL thresholds.

2. Materials and Methods

The concentrations of NO_3^- , NO_2^- , and iodine in 23 commercially available edible algae in the UK (Table 1) were measured. These products were purchased in July 2022 and stored at −20 ◦C pending analyses.

Table 1. Edible algae analysed in this study.

Table 1. *Cont.*

"Date of packaging" and "Best before date" are expressed as DD.MM.YY or MM.YY, when available. * Searched online as it was not mentioned on the package.

2.1. Preparation of Edible Algae Extracts

The extraction of NO_3^- and NO_2^- from edible algae was performed according to Pinto et al. (2015) [24] with some modifications. Five grams of frozen edible algae were pulverised using a grinder (Krups F20342 Grinder, Krups, Germany), except for those edible algae already in powder form. Then, 1.5 g of pulverised/homogenised or powdered edible algae was diluted with 100 mL of hot (70–80 ◦C) ultrapure water (Purelab OptionQ, Oxford, UK) in a 125 mL conical flask. The flask was heated and shaken for 15 min in a boiling water bath. After cooling, 1 mL of each sample was transferred to an Eppendorf tube and stored at −80 ◦C pending biochemistry analysis.

2.2. Analysis of Nitrate ($NO₃⁻$) and Nitrite ($NO₂⁻$)

Eppendorf tubes were thawed and centrifuged at 13,000 rpm for 10 min and at 4 $^{\circ}$ C. The supernatant was then collected and $10 \mu L$ of each sample was injected into a dedicated HPLC analyser (ENO-30; Eicom, San Diego, CA, USA) to measure $\rm NO_3^-$ and $\rm NO_2^-$. Briefly, $NO₃⁻$ and $NO₂⁻$ were separated on a reverse-phase column packed with polystyrene polymer (NO-PAK 4.6 \times 50 mm, EICOM; Amuza, Inc., San Diego, CA, USA). NO₃⁻ was reduced to NO_2^- in a reduction column packed with copper-plated cadmium filings (NO-RED EICOM; Amuza, Inc.). NO_2 ⁻ was mixed with a Griess reagent to form a purple azo dye in a reaction coil. The separation and reaction columns, along with the reaction coil, were placed in a column oven set at 35 ◦C. The absorbance of the dye at 540 nm was measured with a flow-through spectrophotometer (NOD-30; Eicom).

The mobile phase (10% methanol, 0.15 M NaCl/NH4Cl, and 0.5 g/L 4Na-EDTA) and reactor phase (10% methanol, 1.25% HCl containing 5 g/L of sulfanilamide with 0.25 g/L of *N*-naphthylethylenediamine) were delivered at flow rates of 0.33 mL/min and 0.10 mL/min, respectively. A standard curve was produced by injecting 10 μ L of water with sodium NO_3^- (Na NO_3^- /7631-99-4; Sigma Aldrich, St. Louis, MO, USA) and sodium $\rm NO_2^-$ (Na $\rm NO_2^-$ /7632-00-0; Sigma Aldrich) at different concentrations (1.95 μM, 62.5 μM, and 250 μ M). The nori sample was diluted 1:100 using a carrier solution containing 10% methanol, 0.15 M NaCl/NH4Cl, and 0.5 g/L 4Na-EDTA.

2.3. Analysis of Iodine

The extraction of iodine was performed according to Fecher et al. (1998) [25] using an alkali-based extraction to ensure volatile iodine species were kept in solution. Around 100 mg of each homogenised sample was added to separate acid-washed vials $(n = 3$ /species, total = 69 samples). Procedural blanks (containing reagents only, $n = 3$) and a certified reference material (CRM; $n = 3$) with a known concentration of total iodine were also included to ensure the reliability and validity of measurements. Then, 5 mL of high-purity water and 1 mL of 25% tetramethylammonium hydroxide (TMAH) were added to each vial. The vials were sealed with Teflon-lined screw top caps and mixed before being placed in an oven at 90 \pm 5 °C, where the samples were left for 3 h to digest. Following digestion, the samples were transferred to 15 mL centrifuge tubes and diluted to 10 mL with high-purity water. To each sample, Indium and Iridium were spiked in as internal standards to monitor instrument drift throughout the analysis, ready for quantification using inductively coupled plasma mass spectrometry (ICP-MS). A series of dissolved standards were analysed to compare with the unknown samples. The instrument detection limit was 0.0021 µg iodine/L, which equates to 0.19 mg iodine/kg. Analysis of the CRMs showed a concentration of 106.9 ± 5.0 mg/kg, which is similar to other reports $(120 \pm 4$ mg/kg); [26], ensuring the validity and reliability of the measurement of the samples.

2.4. Percentage Contribution to ADI for NO³ [−] *and NO²* [−] *Levels*

The ADI for NO_3^- and NO_2^- is 3.7 mg/kg body mass/day [12] and 0.07 mg/kg body weight/day [19], respectively. Consequently, the percentage contribution to the ADI was calculated using the mean NO_3^- and NO_2^- concentrations obtained from the consumption

of 5 g of edible algae. These calculations were based on the estimated ADI for a 70 kg adult (259 mg/day for NO_3^- and 4.9 mg/day for NO_2^-).

2.5. Percentage Contribution to the UL for Iodine

The percentage contribution to the UL for iodine was calculated based on the iodine concentration obtained from consuming 5 g of edible algae. This calculation references the estimated UL of $600 \mu g/day$ of iodine [22].

2.6. Statistical Analyses

Data were presented as mean \pm standard deviation. The normality of data was assessed using the Shapiro-Wilk test. Differences in the NO_3^- , NO_2^- , and iodine concentrations between different edible algae were compared using the Kruskal-Wallis H test. Differences in the NO_3^- , NO_2^- , and iodine content between macroalgae were assessed using a Mann–Whitney U test. The association between NO_3^- , NO_2^- , and iodine concentrations was analysed using two-tailed Spearman's rank correlation analyses. Data were analysed using the statistical software SPSS (version 28). The level of significance was set at *p* < 0.05.

3. Results

The concentrations of NO_3^- , NO_2^- , and iodine are presented in Table 2. The coefficients of variation for the NO₃⁻ and NO₂⁻ analyses were 7.8 \pm 7.5% and 10.1 \pm 8.0%, respectively. Nori (N1) (17,191.33 \pm 980.89 mg/kg), kombu powder (KO2) (4475.33 \pm 42.36 mg/kg), and sweet kelp (SK1) (3183.00 \pm 147.00 mg/kg) exhibited the highest concentrations of NO³ [−] (*p* < 0.001) (Table 2). Both *Spirulina* samples (S1 and S2) (7.02 ± 0.13 mg/kg and 6.26 ± 0.08 mg/kg, respectively) had the highest concentrations of NO_2^- compared to other edible algae ($p < 0.001$). Regarding iodine, kelp (KE1) (6569.87 \pm 412.26 mg/kg) had the highest concentration, followed by kombu (KO1) (4061.30 \pm 271.66 mg/kg) and sweet kelp (SK1) (3882.07 \pm 214.96 mg/kg), with significantly higher levels than the other samples (p < 0.001). A moderate, negative, and significant association was observed between $\overline{\text{NO}}_2$ ⁻ and iodine ($r = -0.49$; $p = 0.018$). NO₃⁻ and iodine concentrations in macroalgae (NO₃⁻). 4050.13 ± 1925.01 mg/kg; iodine: 1925.01 ± 1455.80 mg/kg) were significantly higher than in microalgae (NO³ [−]: 55.73 ± 93.69 mg/kg; iodine: 17.61 ± 34.87 mg/kg; *p* < 0.001 for both).

Table 2. Concentrations of nitrate ($NO₃⁻$), nitrite ($NO₂⁻$), and iodine in commercial edible algae.

Table 2. *Cont.*

Values are mean \pm standard deviations. Abbreviations: NO₃⁻, nitrate; NO₂⁻, nitrite. ^a Significantly different (p < 0.05) compared to the other edible algae, except D2 and KE1. ^b Significantly different (p < 0.05) compared to N1, S1, and S2. ^c Significantly different ($p < 0.05$) compared to KE1, KK1, SK1, KO1, KO2, KO3, A1, and EG1. ^d Significantly different ($p < 0.05$) compared to the other edible algae, except D1, KE1, and W1. ^e Significantly different (*p* < 0.05) compared to D1, D2, N1, KE1, SK1, and KO2. ^f Significantly different (*p* < 0.05) compared to N1, S1, S2, and C1. ^g Significantly different (*p* < 0.05) compared to the other edible algae. ^h Significantly different $(p < 0.05)$ compared to the other edible algae, except D1, D2, U1, EG1, W1, and DS1. ⁱ Significantly different $(p < 0.05)$ compared to the other edible algae, except EG1. *i* Significantly different $(p < 0.05)$ compared to the other edible algae, except KO1. ^k Significantly different ($p < 0.05$) compared to D1, N1, SW1, and KO2. ¹ Significantly different (*p* < 0.05) compared to the other edible algae, except SK1. ^m Significantly different (*p* < 0.05) compared to the other edible algae, except KO3. ⁿ Significantly different (*p* < 0.05) compared to the other edible algae, except KO2. ^o Significantly different (*p* < 0.05) compared to D1, D2, N1, SK1, and KO2. ^P Significantly different (*p* < 0.05) compared to the other edible algae, except KK1. ^q Significantly different (p < 0.05) compared to the other edible algae, except S2. ^r Significantly different $(p < 0.05)$ compared to the other edible algae, except S1.

Table 3 shows the contributions of NO_3^- and NO_2^- to the ADI, as well as the contribution of iodine to the UL for the analysed samples. The consumption of standard portion sizes of edible algae (5 g/day) did not exceed the ADI levels for either NO_3^- or NO_2^- . However, the same standard portion size (5 g) of some edible algae surpassed the UL for iodine. Dulse (D2), pepper dulse fresh (PD1), wakame (W1), or bladderwrack (B1) exceeded the iodine UL by 132 and 250%. Even more significantly, kelp (KE1), kombu kelp (KK1), sweet kelp (SK1), kombu (KO1, KO2, and KO3), arame (A1), and egg wrack (EG1) exceeded the iodine UL by 625 and 5500%.

Table 3. The percentage (%) contribution to the acceptable daily intake (ADI) for both nitrate ($NO₃⁻$) and nitrite (NO₂⁻), and the percentage (%) contribution to the tolerable upper intake level (UL) for iodine in commercial edible algae.

Values are mean \pm standard deviations. Abbreviations: NO₃⁻, nitrate; NO₂⁻, nitrite; ADI, acceptable daily intake; UL, tolerable upper intake level. † The % contribution to the ADI was calculated using the mean NO_3^- and $NO_2^$ concentrations obtained from the consumption of 5 g of edible algae based on the estimated ADI for a 70 kg adult (259 mg/day for NO₃⁻ and 4.9 mg/day for NO₂⁻). \ddagger The % contribution to the UL for iodine was calculated based on the iodine concentration obtained from consuming 5 g of edible algae, with reference to the estimated UL of 600 µg/day of iodine.

4. Discussion

The main finding of this study was the elevated $NO₃⁻$ concentration in nori (N1) compared to the rest of the samples. The sweet kelp (SK1) and kombu powder (KO2) samples exhibited significant $\bar{\text{NO}_3}^-$ levels, although these were nearly four times lower than in nori (N1). Additionally, the NO_3^- and iodine content in edible macroalgae species was significantly higher than in microalgae species.

These results align with a previous study showing that nori species marketed in Spain had the highest NO_3^- concentration [15]. However, the NO_3^- concentration reported in that study was nearly four times lower (3183 \pm 2279 mg/kg) than in our study $(>17,000 \text{ mg/kg})$. Several environmental factors can significantly influence the absorption of $NO₃⁻$ by edible algae, including water temperature, sunlight exposure, water quality, and cultivation and harvesting practices [27]. Other factors that can affect the nutritional status of commercial edible algae include the origin, commercial status, and shelf life [28]. However, this study did not focus on these aspects. Our main goal was to investigate whether edible algae could be a natural source of NO_3^- . To achieve this, we used highly sensitive HPLC, which is considered one of the gold-standard approaches for measuring $NO₃⁻$ and $NO₂⁻$ in biological samples [16]. This was one of the main strengths of our study compared to previous research in this field [15].

We compared the NO_3^- content in edible algae with that in terrestrial vegetables such as rocket, spinach, and lettuce, which are known to accumulate substantial amounts of NO_3^- . The European Commission has set NO_3^- concentration limits of up to 5000 mg/kg in lettuce and 7000 mg/kg in rocket, as per Regulation (EC) No. 1258/2011, amending Regulation (EC) No. 1881/2006 [29,30]. These limits have been retained by the UK post-Brexit [31] and align with the ADI for NO_3^- of 3.7 mg/kg body mass/day established by the EFSA [12], which is based on the potential association between dietary NO_3^- intake and cancer risk [5]. However, recent evidence suggests that plant-based NO_3 ⁻ may actually be protective against cancer and cardiovascular disease [14]. Therefore, a comprehensive understanding of NO_3 ^{-'}s role in human health is crucial to harness its potential benefits.

To the best of our knowledge, there are no regulations limiting the amount of NO_3^- in commercially available edible algae. Our results showed that nori can concentrate higher NO_3^- levels compared to terrestrial vegetables, whereas sweet kelp and kombu powder samples contained amounts similar to those in lettuce and rocket [32]. However, the dietary portion size of edible algae is between $5-8$ g/day (dry weight) [33], while the portion size for rocket and lettuce is around 80 g [34]. Consequently, the average NO_3^- intake in a dietary portion of nori (5 g) could provide around 85 mg of $NO₃⁻$, whereas a portion of rocket (80 g) could provide over 500 mg of NO_3^- . Thus, it is very unlikely that dietary consumption of edible algae within the recommended levels will exceed the ADI for $NO₃⁻$.

On the other hand, current research suggests that consuming at least 250 mg (4 mmol) of $NO₃⁻$ from terrestrial vegetables can provide some physiological benefits, such as reduced blood pressure [4]. Based on our data, to achieve this amount, one would need to consume at least 15 g of nori (N1) and over 50 g of sweet kelp (SK1) or kombu powder (KO2) dry edible algae. However, some studies have reported a significant reduction in blood pressure levels with lower consumption of edible algae (2 g) [35,36], particularly with nori. Further research is needed to elucidate whether the antihypertensive effect of edible algae is least partially due to their $\mathrm{NO_3}^-$ concentration.

Regarding NO² [−], our analysis revealed that microalgae samples, particularly *Spirulina* (S1), had the highest concentration ($>7 \text{ mg/kg}$). This could be related to the presence of $NO₃⁻$ reductase enzymes in these samples, which reduce $NO₃⁻$ to $NO₂⁻$ [37]. In comparison, some terrestrial vegetables, like Swiss chard and wild rocket, have been reported to contain NO_2^- levels exceeding 50 mg/kg of NO_2^- , though such high levels are uncommon [38,39]. Most studies have reported $NO₂⁻$ values below 2 mg/kg in vegetables like lettuce and rocket [38,39]. Consistent with these findings, we recently reported NO_2^- values of approximately 1 mg/kg in fresh beetroot juice [40], which is seven times lower than the $\overline{NO_2}^-$ concentration observed in *Spirulina* in this study. The ADI level for NO_2^- is 0.07 mg/kg body weight/day, so nearly 5 mg/day for a standard 70 kg person. Consequently, consuming a standard portion size of *Spirulina* (5 g) would contribute significantly less (<0.04 mg) to this threshold.

Edible algae are also an important dietary source of iodine. In our study, the iodine concentration observed in nori (N1), kombu (KK1), and wakame (W1) was consistent with previously reported values for edible algae [41]. We found that kelp (KE1) had the highest iodine concentration, followed by kombu (KO1) and sweet kelp (SK1). Specifically, kelp (KE1) had the highest iodine concentration (6569 mg/kg). Consuming just 9 g of this algae would provide the recommended daily intake of 150 µg/day, and 20 g would meet the upper intake level of $600 \mu g/day$ for adults [22]. However, excessive and recurrent iodine intake can adversely affect thyroid function [21]. It is also important to note that the bioavailability of iodine from edible algae has been reported to be around 50% [42–44]. Further research is needed to better understand the impact of edible algae consumption on thyroid health.

We did not observe an association between the $NO₃⁻$ or $NO₂⁻$ concentration and iodine levels in the samples we analysed. In fact, we found that nori (N1), the sample with the highest NO_3^- concentration, had low iodine levels (16.3 mg/kg). To achieve the recommended daily intake of iodine, nearly 10 g of nori would be needed, and more than 35 g to reach the upper intake level [22]. This amount of nori would provide about 170 mg of $NO₃⁻$ (2.7 mmol), which is below the ADI levels for $NO₃⁻$, but also less than the amount of $NO₃⁻$ (4 mmol) suggested to provide physiological benefits [4]. Additionally, previous research has suggested that salivary NO_3^- uptake may compete with iodine [20]. However, this competition has not been demonstrated when iodine and $NO₃⁻$ were administered together in healthy adults [45].

Concerns have been raised about the accumulation of toxic metals in edible algae, particularly in areas with industrial contamination or poor sewage systems [46]. Elements such as cadmium (Cd), lead (Pb), or mercury (Hg) can be present in marine waters as pollutants [46]. However, this risk is not unique to edible algae; terrestrial vegetables can also accumulate heavy metals when soils are contaminated [47]. Regarding $\text{NO}_3^$ and NO_2^- , consuming small amounts of edible algae is unlikely to pose a higher risk of toxicity compared to terrestrial vegetables, except for some edible algae with high iodine concentrations. Nevertheless, monitoring the accumulation of harmful substances in marine vegetables is essential.

While this study focused on the use of edible algae in the form of food or supplements (powder) in the human diet, it is important to note that algae can also offer solutions to urgent challenges such as water pollution. By absorbing excess nutrients, including nitrates, from aquatic environments, algae can help mitigate the harmful effects of nutrient runoff and eutrophication, thereby contributing to improved water quality and ecosystem health [48]. Additionally, algae with high NO_3^- content can be utilised as organic fertilisers in agriculture, providing a sustainable alternative to chemical fertilisers and promoting soil health and productivity [49]. Thus, exploring the composition of algae is important for addressing environmental concerns and promoting sustainable practices across different sectors.

This study had several limitations worth discussing. Like terrestrial vegetables, the $NO₃⁻$ and $NO₂⁻$ concentrations in edible algae may vary throughout the year due to environmental and harvesting conditions. Consequently, future studies should aim to analyse the variability in the $\overline{\text{NO}_3}^-$ and $\overline{\text{NO}_2}^-$ concentrations in these commercial products across different seasons. Additionally, we did not differentiate between food products and edible algae-based supplements (powder) in this study, as both can be used to enhance the composition of recipes. Furthermore, the bioavailability of $NO₃⁻$, $NO₂⁻$, and iodine was not analysed since our main aim was to identify edible algae with the highest content of these molecules. We aim to conduct further studies using similar products to analyse the bioavailability of NO_3^- , NO_2^- and iodine in humans and to compare it with that of high-NO₃⁻ terrestrial vegetables. Besides NO₃⁻, NO₂⁻, and iodine, future studies should also analyse other nutrients and investigate the presence of potentially harmful compounds such as heavy metals, as these are significant concerns regarding the nutritional value of edible algae.

In summary, this study demonstrated that certain edible macroalgae species, particularly nori (N1) and kombu powder (KO2), can contain substantial quantities of $NO₃⁻$, surpassing levels found in terrestrial vegetables like rocket. However, consuming the

recommended portion size of commercial edible algae (5–8 g /day) is unlikely to exceed the $NO₃$ ⁻ ADI levels. $NO₂$ ⁻ was more abundant in edible microalgae species, especially Spirulina (S1 and S2). Nonetheless, similar to NO₃⁻, consuming standard portion sizes is unlikely to exceed the NO_2^- ADI levels. Macroalgae species such as kelp (KE1), sweet kelp (SK1), and kombu (KO1) exhibited higher iodine levels, and the consumption of small portions of these species $\left($ <1 g) could exceed the UL of iodine. We did not find an association between $\mathrm{NO_3}^-$ and iodine levels, but we did find a negative association between $\mathrm{NO_2}^$ concentration and iodine. Thus, edible algae rich in NO_3^- or NO_2^- did not necessarily contain large quantities of iodine. These findings are relevant to nutritionists, researchers, and commercial companies interested in the nutritional properties of edible algae.

Author Contributions: Conceptualization, P.C.-A. and R.B.; methodology, P.C.-A., N.J.C. and R.B.; software, P.C.-A. and R.B.; validation, P.C.-A., N.J.C. and R.B.; formal analysis, P.C.-A., J.M.H., O.S.B.N., L.V.H., N.J.C. and R.B.; investigation, P.C.-A., J.M.H., O.S.B.N., L.V.H., N.J.C. and R.B.; resources, P.C.-A., N.J.C. and R.B.; data curation, P.C.-A., N.J.C. and R.B.; writing—original draft preparation, P.C.-A. and R.B.; writing—review and editing, P.C.-A., J.M.H., O.S.B.N., L.V.H., N.J.C. and R.B.; visualization, P.C.-A.; supervision, P.C.-A., N.J.C., and R.B.; project administration, P.C.-A. and R.B.; funding acquisition, P.C.-A. and R.B. All authors have read and agreed to the published version of the manuscript.

Funding: This research received financial support from the University of Plymouth 2022 School of Health Professions Pump Priming Fund.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data that support the findings of this study are restricted for research use only. The data are not publicly available. Data are available from the authors upon reasonable request and with permission from the School of Health Professions, University of Plymouth at Plymouth, United Kingdom.

Acknowledgments: We thank Craig Beckley and Sarah E. Clough for their support in the data analysis and Rob Clough for the analytical support with the ICP-MS.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Wu, J.Y.; Tso, R.; Teo, H.S.; Haldar, S. The utility of algae as sources of high value nutritional ingredients, particularly for alternative/complementary proteins to improve human health. *Front. Nutr.* **2023**, *10*, 1277343. [\[CrossRef\]](https://doi.org/10.3389/fnut.2023.1277343) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37904788)
- 2. Figueroa, V.; Farfán, M.; Aguilera, J. Seaweeds as novel foods and source of culinary flavors. *Food Rev. Int.* **2023**, *39*, 1–26. [\[CrossRef\]](https://doi.org/10.1080/87559129.2021.1892749)
- 3. Pina-Pérez, M.C.; Brück, W.M.; Brück, T.; Beyrer, M. Microalgae as healthy ingredients for functional foods. In *The Role of Alternative and Innovative Food Ingredients and Products in Consumer Wellness*; Elsevier: London, UK, 2019; pp. 103–137.
- 4. Ashworth, A.; Bescos, R. Dietary nitrate and blood pressure: Evolution of a new nutrient? *Nutr. Res. Rev.* **2017**, *30*, 208–219. [\[CrossRef\]](https://doi.org/10.1017/S0954422417000063)
- 5. Bondonno, C.P.; Zhong, L.; Bondonno, N.P.; Sim, M.; Blekkenhorst, L.C.; Liu, A.; Rajendra, A.; Pokharel, P.; Erichsen, D.W.; Neubauer, O.; et al. Nitrate: The Dr. Jekyll and Mr. Hyde of human health? *Trends Food Sci. Technol.* **2023**, *135*, 57–73. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2023.03.014)
- 6. Pinaffi-Langley, A.C.d.C.; Dajani, R.M.; Prater, M.C.; Nguyen, H.V.M.; Vrancken, K.; Hays, F.; Hord, N.G. Perspective: Dietary nitrate from plant foods: A conditionally essential nutrient for cardiovascular health. *Adv. Nutr.* **2023**, *15*, 100158. [\[CrossRef\]](https://doi.org/10.1016/j.advnut.2023.100158)
- 7. Burleigh, M.; Liddle, L.; Muggeridge, D.J.; Monaghan, C.; Sculthorpe, N.; Butcher, J.; Henriquez, F.; Easton, C. Dietary nitrate supplementation alters the oral microbiome but does not improve the vascular responses to an acute nitrate dose. *Nitric Oxide* **2019**, *89*, 54–63. [\[CrossRef\]](https://doi.org/10.1016/j.niox.2019.04.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31051259)
- 8. du Toit, L.; Sundqvist, M.L.; Redondo-Rio, A.; Brookes, Z.; Casas-Agustench, P.; Hickson, M.; Benavente, A.; Montagut, G.; Weitzberg, E.; Gabaldón, T.; et al. The effect of dietary nitrate on the oral microbiome and salivary biomarkers in individuals with high blood pressure. *J. Nutr.* **2024**. [\[CrossRef\]](https://doi.org/10.1016/j.tjnut.2024.07.002)
- 9. Moran, S.P.; Rosier, B.T.; Henriquez, F.L.; Burleigh, M.C. The effects of nitrate on the oral microbiome: A systematic review investigating prebiotic potential. *J. Oral Microbiol.* **2024**, *16*, 2322228. [\[CrossRef\]](https://doi.org/10.1080/20002297.2024.2322228)
- 10. Vanhatalo, A.; Blackwell, J.R.; L'Heureux, J.E.; Williams, D.W.; Smith, A.; van der Giezen, M.; Winyard, P.G.; Kelly, J.; Jones, A.M. Nitrate-responsive oral microbiome modulates nitric oxide homeostasis and blood pressure in humans. *Free Radic. Biol. Med.* **2018**, *124*, 21–30. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2018.05.078)
- 11. Bryan, N.S.; Alexander, D.D.; Coughlin, J.R.; Milkowski, A.L.; Boffetta, P. Ingested nitrate and nitrite and stomach cancer risk: An updated review. *Food Chem. Toxicol.* **2012**, *50*, 3646–3665. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2012.07.062)
- 12. EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS); Mortensen, A.; Aguilar, F.; Crebelli, R.; Di Domenico, A.; Dusemund, B.; Frutos, M.J.; Galtier, P.; Gott, D.; Gundert-Remy, U.; et al. Re-evaluation of sodium nitrate (E 251) and potassium nitrate (E 252) as food additives. *Efsa J.* **2017**, *15*, e04787. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32625505)
- 13. Hord, N.G.; Tang, Y.; Bryan, N.S. Food sources of nitrates and nitrites: The physiologic context for potential health benefits. *Am. J. Clin. Nutr.* **2009**, *90*, 1–10. [\[CrossRef\]](https://doi.org/10.3945/ajcn.2008.27131) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19439460)
- 14. Bondonno, N.P.; Pokharel, P.; Bondonno, C.P.; Erichsen, D.W.; Zhong, L.; Schullehner, J.; Frederiksen, K.; Kyrø, C.; Hendriksen, P.F.; Hodgson, J.M.; et al. Source-specific nitrate intake and all-cause mortality in the Danish Diet, Cancer, and Health Study. *Eur. J. Epidemiol.* **2024**, 1–18. [\[CrossRef\]](https://doi.org/10.1007/s10654-024-01133-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38802612)
- 15. Martín-León, V.; Paz, S.; D'Eufemia, P.A.; Plasencia, J.J.; Sagratini, G.; Marcantoni, G.; Navarro-Romero, M.; Gutiérrez, Á.J.; Hardisson, A.; Rubio-Armendáriz, C. Human exposure to toxic metals (Cd, Pb, Hg) and nitrates (NO_3^-) from seaweed consumption. *Appl. Sci.* **2021**, *11*, 6934. [\[CrossRef\]](https://doi.org/10.3390/app11156934)
- 16. Bryan, N.S.; Grisham, M.B. Methods to detect nitric oxide and its metabolites in biological samples. *Free Radic. Biol. Med.* **2007**, *43*, 645–657. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2007.04.026)
- 17. Lobban, C.S.; Harrison, P.J. *Seaweed Ecology and Physiology*; Cambridge University Press: Cambridge, UK, 1994.
- 18. Liao, M.-L.; Seib, P.A. Chemistry of L-ascorbic acid related to foods. *Food Chem.* **1988**, *30*, 289–312. [\[CrossRef\]](https://doi.org/10.1016/0308-8146(88)90115-X)
- 19. EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS); Mortensen, A.; Aguilar, F.; Crebelli, R.; Di Domenico, A.; Dusemund, B.; Frutos, M.J.; Galtier, P.; Gott, D.; Gundert-Remy, U.; et al. Re-evaluation of potassium nitrite (E 249) and sodium nitrite (E 250) as food additives. *Efsa J.* **2017**, *15*, e04786. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32625504)
- 20. Edwards, D.; Fletcher, K.; Rowlands, E. Antagonism between perchlorate, iodide, thiocyanate, and nitrate for secretion in human saliva analogy with the iodide trap of the thyroid. *Lancet* **1954**, *263*, 498–499. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(54)91196-4)
- 21. Farebrother, J.; Zimmermann, M.B.; Andersson, M. Excess iodine intake: Sources, assessment, and effects on thyroid function. *Ann. N. Y. Acad. Sci.* **2019**, *1446*, 44–65. [\[CrossRef\]](https://doi.org/10.1111/nyas.14041)
- 22. EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). Scientific opinion on dietary reference values for iodine. *EFSA J.* **2014**, *12*, 3660.
- 23. Blikra, M.J.; Henjum, S.; Aakre, I. Iodine from brown algae in human nutrition, with an emphasis on bioaccessibility, bioavailability, chemistry, and effects of processing: A systematic review. *Compr. Rev. Food Sci. Food Saf.* **2022**, *21*, 1517–1536. [\[CrossRef\]](https://doi.org/10.1111/1541-4337.12918) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35233943)
- 24. Pinto, E.; Almeida, A.A.; Aguiar, A.A.; Ferreira, I.M. Comparison between the mineral profile and nitrate content of microgreens and mature lettuces. *J. Food Compos. Anal.* **2015**, *37*, 38–43. [\[CrossRef\]](https://doi.org/10.1016/j.jfca.2014.06.018)
- 25. Fecher, P.A.; Goldmann, I.; Nagengast, A. Determination of iodine in food samples by inductively coupled plasma mass spectrometry after alkaline extraction. *J. Anal. At. Spectrom.* **1998**, *13*, 977–982. [\[CrossRef\]](https://doi.org/10.1039/a801671b)
- 26. Gaudry, A.; Zeroual, S.; Cherkaoui el Moursli, R.; Guessous, A.; Chouak, A.; Mouradi, A.; Givernaud, T.; Moskura, M.; Delmas, R. Neutron activation analysis applied to the study of heavy metal marine pollution observed through bioaccumulation in macroscopic algae near El Jadida, Morocco. *J. Radioanal. Nucl. Chem.* **2007**, *271*, 165–171. [\[CrossRef\]](https://doi.org/10.1007/s10967-007-0124-2)
- 27. Khan, M.I.; Shin, J.H.; Kim, J.D. The promising future of microalgae: Current status, challenges, and optimization of a sustainable and renewable industry for biofuels, feed, and other products. *Microb. Cell Factories* **2018**, *17*, 36. [\[CrossRef\]](https://doi.org/10.1186/s12934-018-0879-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29506528)
- 28. Wells, M.L.; Potin, P.; Craigie, J.S.; Raven, J.A.; Merchant, S.S.; Helliwell, K.E.; Smith, A.G.; Camire, M.E.; Brawley, S.H. Algae as nutritional and functional food sources: Revisiting our understanding. *J. Appl. Phycol.* **2017**, *29*, 949–982. [\[CrossRef\]](https://doi.org/10.1007/s10811-016-0974-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28458464)
- 29. European Commission. Regulation (EU) No 1258/2011 of 2 December 2011 amending Regulation (EC) No. 1881/2006 as regards maximum levels for nitrates in foodstuffs. *Off. J. Eur. Union L* **2011**, *320*, 15–17.
- 30. Signore, A.; Bell, L.; Santamaria, P.; Wagstaff, C.; Van Labeke, M.-C. Red light is effective in reducing nitrate concentration in rocket by increasing nitrate reductase activity, and contributes to increased total glucosinolates content. *Front. Plant Sci.* **2020**, *11*, 604. [\[CrossRef\]](https://doi.org/10.3389/fpls.2020.00604)
- 31. The Agricultural Products, Food and Drink (Amendment) (EU Exit) Regulations. Statutory Instruments, 2020(1661). 2020. Available online: https://www.legislation.gov.uk/uksi/2020/1661/pdfs/uksi_20201661_en.pdf (accessed on 7 February 2024).
- 32. Zhong, L.; Blekkenhorst, L.C.; Bondonno, N.P.; Sim, M.; Woodman, R.J.; Croft, K.D.; Lewis, J.R.; Hodgson, J.M.; Bondonno, C.P. A food composition database for assessing nitrate intake from plant-based foods. *Food Chem.* **2022**, *394*, 133411. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2022.133411)
- 33. MacArtain, P.; Gill, C.I.; Brooks, M.; Campbell, R.; Rowland, I.R. Nutritional value of edible seaweeds. *Nutr. Rev.* **2007**, *65*, 535–543. [\[CrossRef\]](https://doi.org/10.1111/j.1753-4887.2007.tb00278.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18236692)
- 34. Public Health England. *The Eatwell Guide: Helping You Eat a Healthy, Balanced Diet*; Public Health England: London, UK, 2016.
- 35. Wada, K.; Nakamura, K.; Tamai, Y.; Tsuji, M.; Sahashi, Y.; Watanabe, K.; Ohtsuchi, S.; Yamamoto, K.; Ando, K.; Nagata, C. Seaweed intake and blood pressure levels in healthy pre-school Japanese children. *Nutr. J.* **2011**, *10*, 83. [\[CrossRef\]](https://doi.org/10.1186/1475-2891-10-83) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21827710)
- 36. Wada, K.; Tsuji, M.; Nakamura, K.; Oba, S.; Nishizawa, S.; Yamamoto, K.; Watanabe, K.; Ando, K.; Nagata, C. Effect of dietary nori (dried laver) on blood pressure in young Japanese children: An intervention study. *J. Epidemiol.* **2021**, *31*, 37–42. [\[CrossRef\]](https://doi.org/10.2188/jea.JE20190176) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32201400)
- 37. Yabuki, Y.; Mori, E.; Tamura, G. Nitrite reductase in the cyanobacterium Spirulina platensis. *Agric. Biol. Chem.* **1985**, *49*, 3061–3062. [\[CrossRef\]](https://doi.org/10.1271/bbb1961.49.3061)
- 38. Aires, A.; Carvalho, R.; Rosa, E.A.; Saavedra, M.J. Effects of agriculture production systems on nitrate and nitrite accumulation on baby-leaf salads. *Food Sci. Nutr.* **2013**, *1*, 3–7. [\[CrossRef\]](https://doi.org/10.1002/fsn3.1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24804008)
- 39. Bahadoran, Z.; Mirmiran, P.; Jeddi, S.; Azizi, F.; Ghasemi, A.; Hadaegh, F. Nitrate and nitrite content of vegetables, fruits, grains, legumes, dairy products, meats and processed meats. *J. Food Compos. Anal.* **2016**, *51*, 93–105. [\[CrossRef\]](https://doi.org/10.1016/j.jfca.2016.06.006)
- 40. Bescos, R.; Rollason, M.L.; Davies, T.S.; Casas-Agustench, P. Content of nitrate and nitrite in commercial and self-made beetroot juices and the effect of storage temperature. *Food Sci. Nutr.* **2023**, *11*, 6376–6383. [\[CrossRef\]](https://doi.org/10.1002/fsn3.3575) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37823101)
- 41. Public Health England. *Composition of Foods Integrated Dataset (CoFID): McCance and Widdowson's Composition of Foods Integrated Dataset on the Nutrient Content of the UK Food Supply*; Public Health England: London, UK, 2021.
- 42. Andersen, S.; Noahsen, P.; Rex, K.F.; Florian-Sørensen, H.C.; Mulvad, G. Iodine in edible seaweed, its absorption, dietary use, and relation to iodine nutrition in Arctic people. *J. Med. Food* **2019**, *22*, 421–426. [\[CrossRef\]](https://doi.org/10.1089/jmf.2018.0187)
- 43. Aquaron, R.; Delange, F.; Marchal, P.; Lognoné, V.; Ninane, L. Bioavailability of seaweed iodine in human beings. *Cell. Mol. Biol.* **2002**, *48*, 563–569. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12146713)
- 44. Combet, E.; Ma, Z.F.; Cousins, F.; Thompson, B.; Lean, M.E. Low-level seaweed supplementation improves iodine status in iodine-insufficient women. *Br. J. Nutr.* **2014**, *112*, 753–761. [\[CrossRef\]](https://doi.org/10.1017/S0007114514001573)
- 45. Bailey, S.J.; Blackwell, J.R.; Wylie, L.J.; Emery, A.; Taylor, E.; Winyard, P.G.; Jones, A.M. Influence of iodide ingestion on nitrate metabolism and blood pressure following short-term dietary nitrate supplementation in healthy normotensive adults. *Nitric Oxide* **2017**, *63*, 13–20. [\[CrossRef\]](https://doi.org/10.1016/j.niox.2016.12.008)
- 46. Filippini, M.; Baldisserotto, A.; Menotta, S.; Fedrizzi, G.; Rubini, S.; Gigliotti, D.; Valpiani, G.; Buzzi, R.; Manfredini, S.; Vertuani, S. Heavy metals and potential risks in edible seaweed on the market in Italy. *Chemosphere* **2021**, *263*, 127983. [\[CrossRef\]](https://doi.org/10.1016/j.chemosphere.2020.127983) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32841878)
- 47. Zhou, H.; Yang, W.-T.; Zhou, X.; Liu, L.; Gu, J.-F.; Wang, W.-L.; Zou, J.-L.; Tian, T.; Peng, P.-Q.; Liao, B.-H. Accumulation of heavy metals in vegetable species planted in contaminated soils and the health risk assessment. *Int. J. Environ. Res. Public Health* **2016**, *13*, 289. [\[CrossRef\]](https://doi.org/10.3390/ijerph13030289) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26959043)
- 48. Racine, P.; Marley, A.; Froehlich, H.E.; Gaines, S.D.; Ladner, I.; MacAdam-Somer, I.; Bradley, D. A case for seaweed aquaculture inclusion in US nutrient pollution management. *Mar. Policy* **2021**, *129*, 104506. [\[CrossRef\]](https://doi.org/10.1016/j.marpol.2021.104506)
- 49. Illera-Vives, M.; Labandeira, S.S.; Fernández-Labrada, M.; López-Mosquera, M.E. Agricultural uses of seaweed. In *Sustainable Seaweed Technologies*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 591–612.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.