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Research Article

Cardiopulmonary Exercise Capacity and Preoperative Markers of Inflammation

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Explanatory mechanisms for the association between poor exercise capacity and infections following surgery are underexplored. We hypothesized that aerobic fitness—assessed by cardiopulmonary exercise testing (CPET)—would be associated with circulating inflammatory markers, as quantified by the neutrophil-lymphocyte ratio (NLR) and monocyte subsets. The association between cardiopulmonary reserve and inflammation was tested by multivariable regression analysis with covariates including anaerobic threshold (AT) and malignancy. In a first cohort of 240 colorectal patients, AT was identified as the sole factor associated with higher NLR (P = 0.03) and absolute and relative lymphopenia (P = 0.01). Preoperative leukocyte subsets and monocyte CD14⁺ expression (downregulated by endotoxin and indicative of chronic inflammation) were also assessed in two further cohorts of age-matched elective gastrointestinal and orthopaedic surgical patients. Monocyte CD14⁺ expression was lower in gastrointestinal patients (n = 43) compared to age-matched orthopaedic patients (n = 31). The circulating CD14⁺CD16⁻ monocyte subset was reduced in patients with low cardiopulmonary reserve. Poor exercise capacity in patients without a diagnosis of heart failure is independently associated with markers of inflammation. These observations suggest that preoperative inflammation associated with impaired cardiorespiratory performance may contribute to the pathophysiology of postoperative outcome.

1. Introduction

Immune dysregulation is a key feature of low cardiac output states. Absolute numbers, as well as function, of monocytes and T-cells are markedly altered in cardiac failure [1]. Severe heart failure is associated with higher levels of circulating endotoxin [2] and lymphopenia [3]. Systemic inflammation driven by exposure to endotoxin in patients with heart failure results in downregulation of monocyte CD14⁺ expression and increased soluble CD14 through shedding of this receptor from the cellular membrane. Alterations in three distinct CD14⁺ monocyte subsets occur in various pathophysiological states, as defined by CD16 (Fc γ RIII) expression. The function of these subsets appears to be highly context-specific, with CD16⁺ subsets frequently expanded in chronic inflammatory conditions. Notably, in patients with chronic systolic heart failure, the nonclassical CD14^{dim}CD16⁺ subset is more prevalent and inversely associated with worsening cardiac performance [4]. By contrast, reduced levels of the classical CD14⁺⁺CD16⁻ monocyte subset are reported in heart failure, compatible with remodelling roles for different subsets [5].

Poor exercise capacity in surgical patients, as measured objectively by preoperative cardiopulmonary exercise [6– 12], is associated with poorer postoperative outcomes including infection—following surgery [13]. Plausible mechanisms that explain the link between poor exercise capacity and poorer postoperative outcomes remain underexplored. Whether preoperative exercise capacity in surgical patients is similarly independently linked to markers of inflammation, as reported in heart failure, remains unclear. We therefore hypothesized that impaired cardiovascular performance (as defined by cardiopulmonary exercise testing) would be associated with biomarkers for inflammation (neutrophillymphocyte ratio and absolute and relative lymphopenia) and/or evidence consistent with surface markers indicative of changes in monocyte function by circulating endotoxin.

2. Materials and Methods

2.1. Patient Populations. Patients undergoing major elective colorectal surgery, including those who were screened for the COMPETE-C randomized controlled trial [14], underwent CPET at Derriford Hospital, Plymouth, UK, as approved by the Cornwall and Plymouth Research Ethics Committee (Medical Research Ethics Committee (MREC): 08/H0203/159). Full details of this trial (including CON-SORT information) have been reported previously [14]. Further preoperative immune analysis was undertaken in two Research Ethics Committee approved studies at University College London Hospitals enrolling elective orthopaedic patients, healthy volunteers (MREC: 09/H0805/59), and patients undergoing major gastrointestinal surgery (MREC: 09/H0805/58). Adherence to STROBE guidelines is documented in Table S1 in Supplementary Material available online at http://dx.doi.org/10.1155/2014/727451.

2.2. Cardiopulmonary Exercise Testing. Patients completed symptom-limited maximal cardiopulmonary exercise testing (CPET) as part of their routine preoperative assessment. Following a period of 2 min rest and 2 min unloaded pedalling, patients began ramped, continuous incremental, and symptom limited exercise on a stationary ergometer (Zan, nSpire, CO, USA), with a ramp gradient 15–20 W/minute.

CPET was stopped as determined by the patients' tolerance. Ventilation and gas exchange variables were measured using a metabolic cart (Zan, nSpire, CO, USA). 12-lead electrocardiogram, noninvasive blood pressure, and pulse oximetry were monitored throughout the test period. Anaerobic threshold (AT) was used as the marker of aerobic fitness. AT was determined by two independent assessors according to the published guidelines using V-slope and confirmed by ventilatory equivalents [15].

Experiment 1 (differential leukocyte counts). Preoperative blood samples were used to assess leukocyte subsets in both centres (Sysmex XE2100 analyzer, Sysmex, Milton Keynes, UK).

Experiment 2 (flow cytometry assessment of monocytes). Heparinised blood samples were collected in heparin from preoperative patients at the same time of day, who had fasted for at least 6 h. All patients were undergoing elective surgery and were free from infection. Flow cytometry (Cyan ADP cytometer, Beckman Coulter, High Wycombe, UK) was performed using 100 μ L of freshly obtained whole blood samples placed on ice immediately after collection and processed in accordance with published guidelines [16]. Samples were processed within 2 h of collection. Control beads were used prior to each run. CD14⁺ expression was quantified in monocytes identified by forward and side scatter characteristics, combined with HLA-DR surface expression. Antibodies for CD14, CD16, and HLA-DR were purchased from Miltenyi Biotec, UK. Respective murine isotype

controls (IgG2a and IgM, resp.; Miltenyi Biotec, UK) were used for each surface marker. Fc receptor blocker was used routinely. After incubation for 20 minutes, samples were fixed with 2% paraformaldehyde and red blood cells were lysed using isotonic ammonium chloride buffer. At least 10000 gated events were captured. Flow cytometry data were analyzed using Kaluza software (version 1.2; Beckman Coulter, High Wycombe, UK). Using an established gating strategy [17], three distinct subsets that are altered in various pathophysiological states of CD14⁺ monocyte subsets were identified by CD16 (Fc γ RIII) expression.

2.3. Statistics. To test the association between cardiopulmonary limitation and preoperative inflammation, multiple regression analysis (1-way hierarchical switching model) was undertaken with NLR as the dependent variable. AT was entered as a continuous independent variable with the following categorical independent variables: risk factors for cardiovascular disease [18], presence of malignancy, age, and gender. The relationship between a preoperative neutrophillymphocyte ratio >2.8—which is predictive of overall survival following colorectal surgery—and AT was also assessed (Fisher's exact test) [19]. For continuous data, variables were analysed with ANCOVA. Nonparametric data were analysed with the Mann-Whitney U test. All reported P values are two-sided. Statistical analyses were performed using NCSS 8 (Kaysville, UT, USA).

2.4. Sample Size Calculations. We powered the primary outcome (NLR) on the basis that ~30% colorectal patients with low AT (<11 mL·kg⁻¹·min⁻¹) [13] would exhibit a prognostically significant difference in NLR ≥ 0.5 [20–23], with an anticipated standard deviation of 1.0. Thus, ≥ 185 patients with AT > 11 mL·kg⁻¹·min⁻¹, compared with ≥ 56 patients with low AT (alpha = 0.05; power = 90%). Based on preceding studies in the cardiac failure literature measuring soluble CD14 [24], we sought to find an absolute difference in monocyte CD14 median fluorescence intensity of 25 arbitrary units (anticipated standard deviation of 25 arbitrary units), thus requiring a sample size of ≥ 32 cases per group ($\alpha = 0.01$; power = 90%).

3. Results

Experiment 1 (CPET physiological characteristics and leukocyte subsets). Demographics and associated cardiopulmonary test parameters of patients undergoing preoperative CPET are shown in Table 1. The majority of anaerobic threshold values were consistent with those reported for NYHA Classes 3-4 (Figure 1). Impaired cardiovascular performance was associated independently with higher NLR (P = 0.04) and absolute (P = 0.007) and relative lymphopenia (P =0.004), adjusted for the presence of malignancy. Unadjusted for malignancy, low AT was associated with higher NLR (low AT: +0.54 (95% CI: 0.1–0.98); P = 0.01) and absolute (low AT: -0.20 lymphocytes 10⁹ mL⁻¹ (95% CI: 0.01–0.40); P =0.04) and relative lymphopenia (low AT: -3.4% (95% CI: 1.05–5.79); P = 0.005; Figure 2).

	$V_{\rm AT} < 11 {\rm mL kg^{-1} min^{-1}}$	$V_{\rm AT} > 11 {\rm mL kg^{-1} min^{-1}}$	<i>P</i> value
Number	79	161	
Age (y)	71 (69–73)	63 (61–66)	0.0001
Gender (male:female ratio)	1.05	1.47	0.30
Cancer (<i>n</i> ; %)	66 (86%)	114 (72%)	0.02
Chemotherapy (<i>n</i> ; %)	21 (13%)	11 (14%)	0.82
Body mass index (kg m ²)	29.6 (28.5–30.6)	27.4 (26.7–28.0)	0.0004
$\dot{V}_{E}/\dot{V}CO_{2}$	30.3 (28.9–31.7)	27.8 (27.2–28.5)	0.0004
Peak $\dot{V}O_2$ (mL kg ⁻¹ min ⁻¹)	14.8 (13.9–15.6)	22.5 (21.5–23.4)	< 0.0001

TABLE 1: CPET demographics in colorectal surgery cohort, stratified according to the prognostically relevant anaerobic threshold $<11 \, mL \, kg^{-1} \min^{-1}$ and $>11 \, mL \, kg^{-1} \min^{-1}$.

Data are presented as mean (95% confidence intervals), unless stated otherwise.

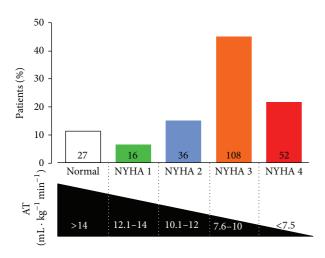


FIGURE 1: Histogram showing numbers of patients (n = 240) stratified by AT-defined NYHA class. Cutoff values for AT estimated from recent published series [25, 26].

Controlling for the presence of malignancy (present in 77% of patients), low AT remained associated with higher NLR (low AT: +0.54 (95% CI: 0.1–0.98); P = 0.01) and absolute (low AT: -0.20 lymphocytes $10^9 \cdot mL^{-1}$ (95% CI: 0.01–0.40); P = 0.04) and relative (low AT: -3.4% (95% CI: 1.05–5.79); P = 0.005) lymphopenia, with no significant interaction observed between malignancy and AT < 11 mL·kg⁻¹·min⁻¹ (P = 0.29). Multiple regression analysis identified AT as the sole factor associated with higher NLR (P = 0.033). An AT < 11 mL·kg⁻¹·min⁻¹ was strongly associated with a preoperative NLR reported to be predictive of survival postoperatively (relative risk: 1.9 (95% confidence intervals: 1.4–2.5); P = 0.01) [19].

Experiment 2 (preoperative monocyte CD14⁺ expression). Next, we assessed whether age-matched patients free of malignancy showed any evidence for systemic inflammation as indicated by biomarker levels found in patients undergoing colorectal surgery. We reasoned that monocyte CD14 surface expression would be reduced in patients with low AT,

compared to age-matched controls with no clinical evidence for cardiac failure. Preoperative neutrophil-lymphocyte ratio was higher (P = 0.01), and monocyte CD14⁺ expression was lower (-112 median fluorescence units (95% CI: 49–176); P = 0.002) in 38 patients undergoing major surgery for gastrointestinal malignancy (median AT: 10 mL·kg⁻¹·min⁻¹ (IQR 9–11)), compared to age-matched orthopaedic patients (n = 31) without overt clinical heart failure (Figures 3(a) and 3(b)). CD14⁺ monocyte subset analysis (Figure 3(c)) showed that the CD14⁺CD16⁻ subset was reduced in patients with low cardiopulmonary reserve (Figure 3(d)).

4. Discussion

Our data show that impaired cardiovascular performance as measured by CPET is associated with changes in leukocyte subsets that have emerged as robust prognostic markers of colorectal surgical outcome. Consistent with these data, we also found that downregulation of monocyte CD14⁺ expression and lymphopenia were associated with impaired cardiorespiratory reserve.

Maintenance, or enhancement, of exercise capacity has been associated with a range of beneficial immune responses [27]. Habitual exercise is associated with improved immune responses to vaccination [28] and outcomes following viral infections [29] and malignancy [30]. Immune dysregulation is a pivotal feature of poor aerobic capacity [1], with the most severe grades of heart failure associated with higher levels of circulating endotoxin [2] and lymphopenia [3]. Our data suggest that in higher-risk surgical patients similar immune dysregulation may occur, even though such patients have never received a formal diagnosis of cardiac failure. Since the presence of overt cardiac failure is an important prognostic factor in determining postoperative outcome [31], our data also suggest that dysregulation of inflammation is associated with poor cardiopulmonary reserve in preoperative patients. Preexisting inflammation could contribute to increased morbidity and mortality after surgery—in addition to any putative mechanistic role for impaired perioperative cardiovascular performance. Interestingly, several studies have reported an association between preoperative NLR and postoperative complications [19-23] but have not made the

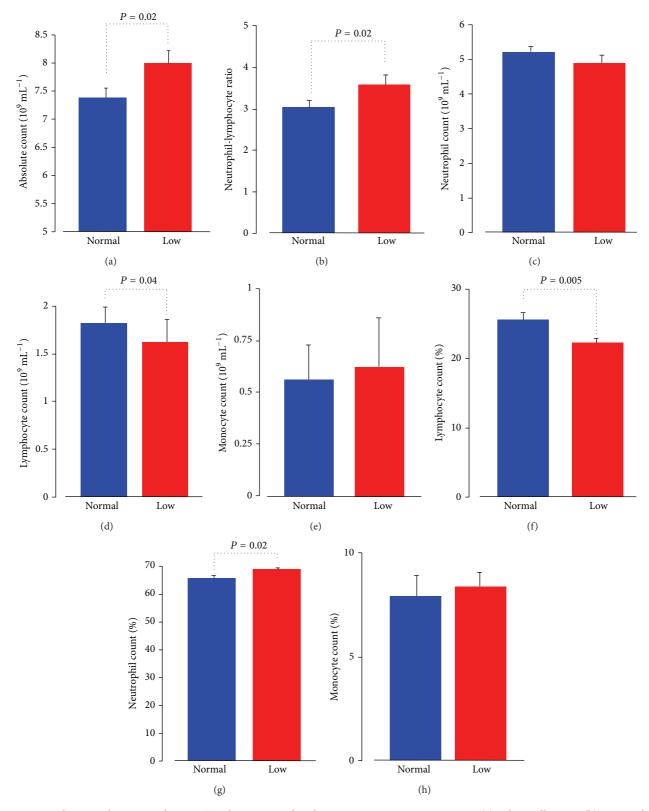


FIGURE 2: Leukocyte subsets according to AT value associated with poorer postoperative outcomes. (a) White cell count. (b) Neutrophillymphocyte ratio. (c) Absolute neutrophil count. (d) Absolute lymphocyte count. (e) Absolute monocyte count. (f) Proportion of lymphocytes. (g) Proportion of neutrophils. (h) Proportion of monocytes. All data are represented as mean \pm SD; n = 240 patients.

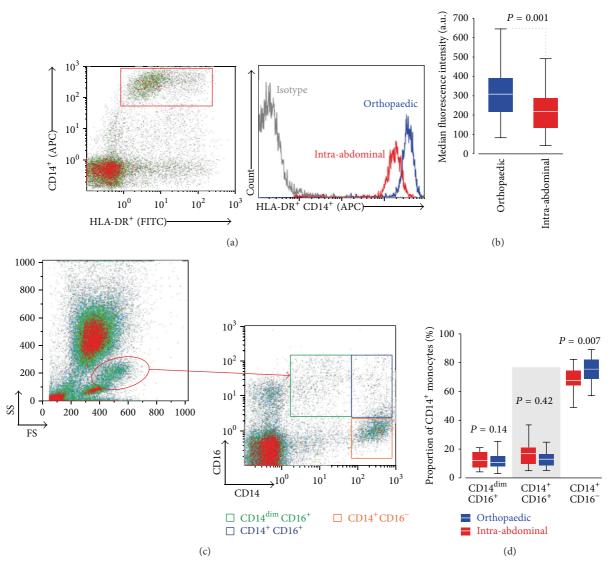


FIGURE 3: Monocyte CD14⁺ expression. (a) Representative HLA-DR⁺ CD14⁺ expression (median fluorescence intensity) in orthopedic (blue) and gastrointestinal (red) surgical patients. Isotype control shown in grey. (b) Summary data for orthopedic (n = 43 patients) and gastrointestinal malignancy (n = 31 patients). (c) Gating strategy to define HLA-DR⁺ CD14⁺ monocyte subsets, including CD16⁺ cells. (d) Proportions of monocyte CD14⁺ subsets according to CD16⁺ expression. All data are represented as median (IQR).

link with another strong predictor of postoperative morbidity—aerobic performance. Taken together, these data suggest that low AT may therefore be a marker, rather than a direct mediator, of poorer perioperative outcomes.

Experimental and clinical data suggest that established immune dysfunction is instrumental in driving impaired cardiovascular function, rather than merely being a marker of disease progression [1]. Thus, targeting immune function is an increasingly important line of investigation in efforts to reduce the substantial mortality from the increasing prevalence of cardiac failure. Strikingly, similar data from the perioperative literature supports this concept: patients with evidence for preoperative depletion of antibodies to endotoxin exhibit higher levels of proinflammatory cytokines and sustain more perioperative morbidity [32, 33]. Downregulation of CD14⁺ is consistent with the hypothesis that chronic systemic exposure to low plasma levels of endotoxin occurs as a result of (occult) cardiac failure [34, 35]. Alternatively, these data may reflect gastrointestinal pathology and endotoxin "leak" [36]. Nevertheless, our results are consistent with previous studies reporting that the CD14⁺⁺CD16⁻ monocyte subset was reduced in patients with chronic systolic heart failure [34].

Monocyte-macrophage function is altered by exerciseinduced activation of both sympathetic and hypothalamicpituitary axes [37]. Absolute monocyte count transiently increases in response to acute exercise, accompanied by preferential demarginalization of the proinflammatory CD14⁺CD16⁺ subset [38] and downregulation of Toll-like receptors [39]. After cessation of exercise, the CD14⁺CD16⁺ subset declines-indicative of remarginalization or recruitment to tissue [40]. Sustained exercise is associated with reduced circulating monocyte inflammatory responses to lipopolysaccharide, lower TLR4 expression, and a lower percentage of the proinflammatory CD14⁺CD16⁺ [41]. Human data regarding the impact of chronic exercise on, as well as tissue resident function of, monocyte function is limited. However, murine models suggest that exercisetrained mice demonstrate enhanced splenic macrophage proliferative capacity and peritoneal macrophages phagocytic activity [42]. Patients with more advanced heart failure, as reflected by low AT, demonstrate an increased number of neutrophils. This relative neutrophilia is accompanied by a decreased number of lymphocytes, characterised by higher proportions of terminally differentiated CD4⁺ and CD8⁺ T-subsets [43]. These data are consistent with molecular mechanisms driving a T-cell immunosenescent state [44].

We acknowledge several limitations of this exploratory study. Further detailed analyses of leukocyte subset functionality stratified by AT could be instructive; in particular, CD14⁺ subsets associated with an augmented proinflammatory phenotype or different migratory properties require further elucidation. Measurement of endotoxin levels would add further insight. The association between NLR and coronary artery disease [45] may reveal mechanistic links to perioperative outcomes, including myocardial injury. Ultimately, detailed outcome data—including propensity to infectious complications—will test the hypothesis further that low AT may be associated with defective innate and/or adaptive immune functionality.

In summary, impaired cardiovascular performance is associated with changes in leukocyte subsets that have emerged as robust prognostic markers of colorectal surgical outcome. Consistent with these data, we also find that downregulation of monocyte CD14⁺ expression and lymphopenia are associated with impaired cardiorespiratory reserve. These observations suggest that preoperative inflammation, associated with impaired cardiorespiratory performance, may contribute a pathophysiological role in determining postoperative outcome.

Abbreviations

AT: Anaerobic threshold.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Authors' Contribution

Gareth L. Ackland carried out the study design, data collection, flow cytometry, and data analysis. Pervez Sultan took part in the data collection, flow cytometry, and data analysis. Mark R. Edwards, Ana Gutierrez del Arroyo, and David Cain participated in the data collection and flow cytometry. Richard Struthers, Robert Sneyd and Gary Minto assisted in the CPET data collection and data analysis.

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References

- V. K. Topkara, S. Evans, W. Zhang et al., "Therapeutic targeting of innate immunity in the failing heart," *Journal of Molecular and Cellular Cardiology*, vol. 51, no. 4, pp. 594–599, 2011.
- [2] J. Niebauer, H. D. Volk, M. Kemp et al., "Endotoxin and immune activation in chronic heart failure: a prospective cohort study," *The Lancet*, vol. 353, no. 9167, pp. 1838–1842, 1999.
- [3] M. Vaduganathan, A. P. Ambrosy, S. J. Greene et al., "Predictive value of low relative lymphocyte count in patients hospitalized for heart failure with reduced ejection fraction: insights from the EVEREST trial," *Circulation: Heart Failure*, vol. 5, no. 6, pp. 750–758, 2012.
- [4] O. Amir, I. Spivak, I. Lavi, and M. A. Rahat, "Changes in the monocytic subsets CD14^{dim}CD16⁺ and CD14⁺⁺CD16⁻ in chronic systolic heart failure patients," *Mediators of Inflammation*, vol. 2012, Article ID 616384, 9 pages, 2012.
- [5] S. Apostolakis, G. Y. H. Lip, and E. Shantsila, "Monocytes in heart failure: Relationship to a deteriorating immune overreaction or a desperate attempt for tissue repair?" *Cardiovascular Research*, vol. 85, no. 4, pp. 649–660, 2010.
- [6] M. Colson, J. Baglin, S. Bolsin, and M. P. W. Grocott, "Cardiopulmonary exercise testing predicts 5 yr survival after major surgery," *British Journal of Anaesthesia*, vol. 109, no. 5, pp. 735– 741, 2012.
- [7] R. A. Hartley, A. C. Pichel, S. W. Grant et al., "Preoperative cardiopulmonary exercise testing and risk of early mortality following abdominal aortic aneurysm repair," *British Journal of Surgery*, vol. 99, no. 11, pp. 1539–1546, 2012.
- [8] F. Ausania, C. P. Snowden, J. M. Prentis et al., "Effects of low cardiopulmonary reserve on pancreatic leak following pancreaticoduodenectomy," *British Journal of Surgery*, vol. 99, no. 9, pp. 1290–1294, 2012.
- [9] M. A. Junejo, J. M. Mason, A. J. Sheen et al., "Cardiopulmonary exercise testing for preoperative risk assessment before hepatic resection," *British Journal of Surgery*, vol. 99, no. 8, pp. 1097– 1104, 2012.

- [10] M. Swart and J. B. Carlisle, "Case-controlled study of critical care or surgical ward care after elective open colorectal surgery," *British Journal of Surgery*, vol. 99, no. 2, pp. 295–299, 2012.
- [11] P. Murray, P. Whiting, S. P. Hutchinson, R. Ackroyd, C. J. Stoddard, and C. Billings, "Preoperative shuttle walking testing and outcome after oesophagogastrectomy," *British Journal of Anaesthesia*, vol. 99, no. 6, pp. 809–811, 2007.
- [12] J. T. Lee, E. J. Chaloner, and S. J. Hollingsworth, "The role of cardiopulmonary fitness and its genetic influences on surgical outcomes," *British Journal of Surgery*, vol. 93, no. 2, pp. 147–157, 2006.
- [13] R. J. T. Wilson, S. Davies, D. Yates, J. Redman, and M. Stone, "Impaired functional capacity is associated with all-cause mortality after major elective intra-abdominal surgery," *British Journal of Anaesthesia*, vol. 105, no. 3, pp. 297–303, 2010.
- [14] C. Challand, R. Struthers, J. R. Sneyd et al., "Randomized controlled trial of intraoperative goal-directed fluid therapy in aerobically fit and unfit patients having major colorectal surgery," *British Journal of Anaesthesia*, vol. 108, no. 1, pp. 53– 62, 2012.
- [15] H. L. Pina, G. J. Balady, P. Hanson, A. J. Labovitz, D. W. Madonna, and J. Myers, "Guidelines for clinical exercise testing laboratories: a statement for healthcare professionals from the committee on exercise and cardiac rehabilitation, American Heart Association," *Circulation*, vol. 91, no. 3, pp. 912–921, 1995.
- [16] G. Monneret, F. Venet, C. Meisel, and J. C. Schefold, "Assessment of monocytic HLA-DR expression in ICU patients: analytical issues for multicentric flow cytometry studies," *Critical Care*, vol. 14, no. 4, article 432, 2010.
- [17] J. Cros, N. Cagnard, K. Woollard et al., "Human CD14dim monocytes patrol and sense nucleic acids and viruses via TLR7 and TLR8 receptors," *Immunity*, vol. 33, no. 3, pp. 375–386, 2010.
- [18] G. L. Ackland, S. Harris, Y. Ziabari, M. Grocott, and M. Mythen, "Revised cardiac risk index and postoperative morbidity after elective orthopaedic surgery: a prospective cohort study," *British Journal of Anaesthesia*, vol. 105, no. 6, pp. 744–752, 2010.
- [19] L. Jankova, O. F. Dent, C. Chan, P. Chapuis, and S. J. Clarke, "Preoperative neutrophil/lymphocyte ratio predicts overall survival but does not predict recurrence or cancer-specific survival after curative resection of node-positive colorectal cancer," *BMC Cancer*, vol. 13, article 442, 2013.
- [20] M. J. Proctor, D. C. McMillan, D. S. Morrison, C. D. Fletcher, P. G. Horgan, and S. J. Clarke, "A derived neutrophil to lymphocyte ratio predicts survival in patients with cancer," *British Journal of Cancer*, vol. 107, no. 4, pp. 695–699, 2012.
- [21] P. Ding, X. An, R. Zhang et al., "Elevated preoperative neutrophil to lymphocyte ratio predicts risk of recurrence following curative resection for stage IIA colon cancer," *International Journal of Colorectal Disease*, vol. 25, no. 12, pp. 1427–1433, 2010.
- [22] K. J. Halazun, A. Aldoori, H. Z. Malik et al., "Elevated preoperative neutrophil to lymphocyte ratio predicts survival following hepatic resection for colorectal liver metastases," *European Journal of Surgical Oncology*, vol. 34, no. 1, pp. 55–60, 2008.
- [23] E. J. Cook, S. R. Walsh, N. Farooq, J. C. Alberts, T. A. Justin, and N. J. Keeling, "Post-operative neutrophil-lymphocyte ratio predicts complications following colorectal surgery," *International Journal of Surgery*, vol. 5, no. 1, pp. 27–30, 2007.
- [24] S. D. Anker, K. R. Egerer, H. D. Volk, W. J. Kox, P. A. Poole-Wilson, and A. J. S. Coats, "Elevated soluble CD14 receptors and altered cytokines in chronic heart failure," *American Journal of Cardiology*, vol. 79, no. 10, pp. 1426–1430, 1997.

- [25] J. E. Hansen, X. G. Sun, and W. W. Stringer, "A simple new visualization of exercise data discloses pathophysiology and severity of heart failure," *Journal of the American Heart Association*, vol. 1, no. 3, Article ID e001883, 2012.
- [26] C. C. Lang, P. Karlin, J. Haythe, T. K. Lim, and D. M. Mancini, "Peak cardiac power output, measured noninvasively, is a powerful predictor of outcome in chronic heart failure," *Circulation: Heart Failure*, vol. 2, no. 1, pp. 33–38, 2009.
- [27] N. Mathur and B. K. Pedersen, "Exercise as a mean to control low-grade systemic inflammation," *Mediators of Inflammation*, vol. 2008, Article ID 109502, 6 pages, 2008.
- [28] AR. Pascoe, "The effects of exercise on vaccination responses: a review of chronic and acute exercise interventions in humans," *Brain, Behavior, and Immunity*, pp. 33–41, 2014.
- [29] R. J. Simpson, "Aging, persistent viral infections, and immunosenescence: can exercise "make space"?" *Exercise and Sport Sciences Reviews*, vol. 39, no. 1, pp. 23–33, 2011.
- [30] R. Ballard-Barbash, C. M. Friedenreich, K. S. Courneya, S. M. Siddiqi, A. McTiernan, and C. M. Alfano, "Physical activity, biomarkers, and disease outcomes in cancer survivors: a systematic review," *Journal of the National Cancer Institute*, vol. 104, no. 11, pp. 815–840, 2012.
- [31] B. G. Hammill, L. H. Curtis, E. Bennett-Guerrero et al., "Impact of heart failure on patients undergoing major noncardiac surgery," *Anesthesiology*, vol. 108, no. 4, pp. 559–567, 2008.
- [32] E. Bennett-Guerrero, M. H. Panah, G. Robin Barclay et al., "Decreased endotoxin immunity is associated with greater mortality and/or prolonged hospitalization after surgery," *Anesthesiology*, vol. 94, no. 6, pp. 992–998, 2001.
- [33] E. Bennett-Guerrero, L. Ayuso, C. Hamilton-Davies et al., "Relationship of preoperative antiendotoxin core antibodies and adverse outcomes following cardiac surgery," *The Journal of the American Medical Association*, vol. 277, no. 8, pp. 646–650, 1997.
- [34] O. Amir, I. Spivak, I. Lavi, and M. A. Rahat, "Changes in the monocytic subsets CD14^{dim}CD16⁺ and CD14⁺⁺CD16⁻ in chronic systolic heart failure patients," *Mediators of Inflammation*, vol. 2012, Article ID 616384, 9 pages, 2012.
- [35] C. Barisione, S. Garibaldi, G. Ghigliotti et al., "CD14CD16 monocyte subset levels in heart failure patients," *Disease Markers*, vol. 28, no. 2, pp. 115–124, 2010.
- [36] B. M. Charalambous, R. C. M. Stephens, I. M. Feavers, and H. E. Montgomery, "Role of bacterial endotoxin in chronic heart failure: the gut of the matter," *Shock*, vol. 28, no. 1, pp. 15–23, 2007.
- [37] N. P. Walsh, M. Gleeson, R. J. Shephard et al., "Position statement part one: immune function and exercise," *Exercise Immunology Review*, vol. 17, pp. 6–63, 2011.
- [38] S. Hong and P. J. Mills, "Effects of an exercise challenge on mobilization and surface marker expression of monocyte subsets in individuals with normal vs. elevated blood pressure," *Brain, Behavior, and Immunity*, vol. 22, no. 4, pp. 590–599, 2008.
- [39] R. J. Simpson, B. K. McFarlin, C. McSporran, G. Spielmann, B. Ó. Hartaigh, and K. Guy, "Toll-like receptor expression on classic and pro-inflammatory blood monocytes after acute exercise in humans," *Brain, Behavior, and Immunity*, vol. 23, no. 2, pp. 232–239, 2009.
- [40] B. Steppich, F. Dayyani, R. Gruber, R. Lorenz, M. Mack, and H. W. L. Ziegler-Heitbrock, "Selective mobilization of CD14+ CD16+ monocytes by exercise," *American Journal of Phy*siology—Cell Physiology, vol. 279, no. 3, pp. C578–C586, 2000.

- [41] M. Gleeson, N. C. Bishop, D. J. Stensel, M. R. Lindley, S. S. Mastana, and M. A. Nimmo, "The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease," *Nature Reviews Immunology*, vol. 11, no. 9, pp. 607–615, 2011.
- [42] J. Woods, Q. Lu, M. A. Ceddia, and T. Lowder, "Special feature for the Olympics: effects of exercise on the immune system: exercise-induced modulation of macrophage function," *Immunology & Cell Biology*, vol. 78, no. 5, pp. 545–553, 2000.
- [43] M. A. Moro-Garcia, A. Echeverria, M. C. Galan-Artimez et al., "Immunosenescence and inflammation characterize chronic heart failure patients with more advanced disease," *International Journal of Cardiology*, vol. 174, no. 3, 2014.
- [44] R. Macaulay, A. N. Akbar, and S. M. Henson, "The role of the T cell in age-related inflammation," *Age*, vol. 35, no. 3, pp. 563–572, 2013.
- [45] A. J. Fowler and R. A. Agha, "Neutrophil/lymphocyte ratio is related to the severity of coronary artery disease and clinical outcome in patients undergoing angiography—the growing versatility of NLR," *Atherosclerosis*, vol. 228, no. 1, pp. 44–45, 2013.