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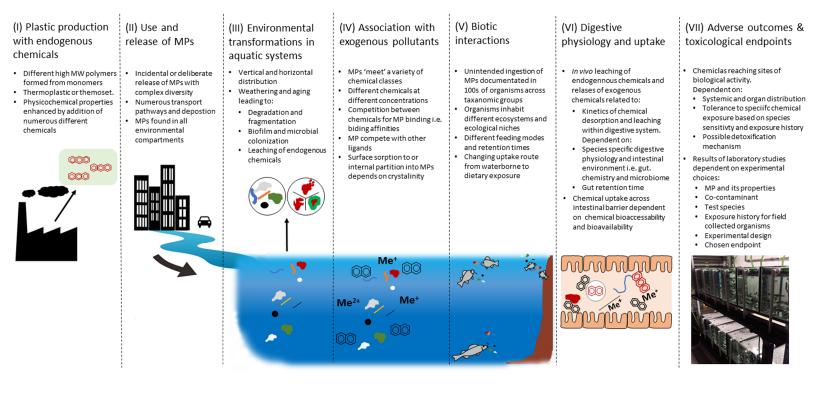
The ecotoxicological consequences of microplastics and co-contaminants in aquatic organisms: a mini-review

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- 1 Summary points (3–5 summary points)
- The transfer of endogenous or exogenous co-contaminants from microplastics to biota is one
 of the most studied aspects of plastic pollution.
- Consensus as to the validity and relevance of MPs as chemical carriers is still debated.
- A multitude of inter-connected factors from production and release, environmental
 transformations to biological and physiological interactions need to be considered.
- Greater environmental realism is needed to bridge the gap between lab studies and the realworld.
- 9 New particles such as nanoplastics, tire wear particles and bioplastics expand the scope for
 10 chemical transfer.

12 The ecotoxicological consequences of microplastics and co-contaminants in aquatic organisms: A

13 mini-review

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25

26 Abstract

27 Microplastics (MPs, <5 mm in size) are a grave environmental concern. They are a ubiquitous 28 persistent pollutant group that has reached into all parts of the environment - from the highest 29 mountain tops to the depths of the ocean. During their production, plastics have added to them 30 numerous chemicals in the form of plasticizers, colorants, fillers and stabilizers, some of which have 31 known toxicity to biota. When released into the environments, MPs are also likely to encounter 32 chemical contaminants, including hydrophobic organic contaminants, trace metals and 33 pharmaceuticals, which can sorb to plastic surfaces. Additionally, MPs have been shown to be 34 ingested by a wide range of organisms and it is this combination of ingestion and chemical 35 association that gives weight to the notion that MPs may impact the bioavailability and toxicity of 36 both endogenous and exogenous co-contaminants. In this mini-review we set the recent literature 37 within what has been previously published about MPs as chemical carriers to biota, with particular 38 focus on aquatic invertebrates and fish. We then present a critical viewpoint on the validity of lab-to-39 field extrapolations in this area. Lastly, we highlight the expanding 'microplastic universe' with the 40 addition of anthropogenic particles that have gained recent attention, namely, tire wear particles, 41 nanoplastics and, bio-based or biodegradable MPs, and highlight the need for future research in their 42 potential roles as vehicles of co-contaminant transfer.

Keywords: Plastic pollution; Additives; MP-associated chemicals; Vector effect; Bioavailability;
Bioaccessibility; Uptake; Toxicity

46

47 1. Introduction - The case for microplastics as a vehicle for chemicals

48 The term 'Microplastics' (MPs) is used as a catch-all term to represent a complex variety of 49 properties that arise during both the manufacturing process and following release into the 50 environment (1). Plastics are composed of different organic polymers to which an array of chemicals 51 (termed here as 'endogenous chemicals', e. g., plasticizers, colorants, fillers and stabilizers) are added 52 to enhance certain properties, such as rigidity, malleability, or thermal resistance, and prolonging life 53 (2). Depending on use, plastics are produced within the microplastic size range of < 5 mm (primary 54 MPs such as microbeads) or into larger products that can subsequently breakdown releasing MPs 55 (secondary MPs) (3,4). Microplastics in the environment exist as a heterogeneous mixture of physical 56 and chemical properties forms (1,4,5) and undergo several environmental transformations such as 57 weathering, fragmentation, and biofilm and microbial colonization (6-8).

58

59 These transformations will affect how MPs interact within their environment – an environment that 60 already contains a plethora of potential co-contaminants (termed here as exogenous chemicals). 61 Tens of thousands of chemical entities are found on the global market with approximately 2000 new 62 chemicals added each year (9). The widespread ingestion of MPs, documented across aquatic taxa 63 (10–12), provides a pathway for both endogenous and exogenous chemicals to enter the organism. 64 Yet the role of MPs as chemical vectors not only relies upon the association with co-contaminants 65 and the influence of the ambient environment, but considerations of the organism's biology and 66 physiology are also important. For instance, feeding modes, gut retention times and digestive 67 physiology will all play a role in whether the MP-associated chemical is bioaccessible and then 68 bioavailable to the organism (5,13). Toxicological consequences may result if the co-contaminants 69 reach and become available at sites of biological activity.

70

The role of MPs as chemical carriers has been the subject of much investigation, debate and speculation (see reviews (5,14–18)), but consensus remains elusive. The interactions of MPs, environment, chemicals, and biota are summarized in Figure 1, and whilst laboratory studies can only investigate a portion of this complexity at one time, it is precisely this complexity that has made the role MPs in co-contaminant transfer one of the most studied and divergent topics in MP research.

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77 2. Biotic effects of microplastics and co-contaminant exposure

78 2a. Endogenous chemicals

79 During the manufacturing of plastics various substances often termed as "additives" are combined 80 with the polymeric resins to improve properties of final applications and a few other reaction by-81 products will be further accidentally incorporated (2,19,20). When plastic debris reach aquatic 82 environments, these endogenous substances, some of which are known to be toxic to biota, can 83 migrate from the resin to the external medium, as substances are often physically, rather than 84 chemically, bonded to the main polymer matrix (20). The leaching of substances from MPs may occur 85 at higher rates compared to macroplastic litter due to their increased surface area to volume ratio. 86 The majority of organic additives have a low hydrophobicity, or low octanol-water partition coefficient (K_{ow}), and a low molecular weight (21). Therefore, exposure of biota can be low due to the 87 88 low diffusion of organic chemicals from the plastic to the water (21,22), as additives with a higher 89 potential for toxicity and bioaccumulation have a higher K_{ow} (19).

90

91 Some endogenous chemicals found in aquatic environments are known to have toxicological 92 properties, such as phthalates, bisphenol A (BPA), nonylphenol (NP), and brominated flame 93 retardants (BFR) (19), as well as trace metals such as cadmium (Cd), lead (Pb), antimony (Sb), and tin 94 (Sn) (2). Of concern are the leaching rates of endogenous chemicals from weathered MPs (20), in 95 particular when in gastrointestinal fluids during digestion where high levels of surfactants and lower 96 pH may facilitate the migration process of compounds from the plastic resin (23). Gut retention times 97 of MPs vary amongst invertebrate species depending on physiology and relative MP size and shape 98 and can last between a few hours and weeks. During this period, endogenous substances will 99 increase in their bioaccessibility and bioavailability due to a low pH environment which enhances 100 leaching from the polymeric matrix and further due to an affinity to fatty tissues of hydrophobic 101 endogenous chemicals (22). For example, Kühn et al. (2020) (24) demonstrated in vitro that 102 environmental MPs will leach additives to stomach oil of northern fulmars. Tanaka et al. (2020) (25) 103 demonstrated that feeding seabird chicks with MPs spiked in their resin with additives induced accumulation at $10^1 - 10^5$ times above baseline in the tested individuals. However, a modelling 104 105 exercise demonstrated that MPs may have a residual contribution to the accumulation and toxicity 106 induced by leachates in the intestinal tracts of lugworms and in the North Sea cod when compared to 107 sources of these contaminants such as water or sediments, but in vivo experimental validation is still 108 required (22).

109

110 2b. Exogenous chemicals

111 Microplastics enter an environment that already contain a chemical cocktail, including hydrophobic 112 organic contaminants (HOCs i.e., polycyclic aromatic hydrocarbons (PAHs) and polychlorinated 113 biphenyls (PCBs)) (26,27), trace metals (28,29) and pharmaceuticals (30–32), all of which have been 114 measured on MPs collected from the environment. These exogenous chemicals can 'sorb' (covering 115 both surface adsorption and internal partition) to the MP based on the structure of the polymer (i.e., 116 its ratio of crystalline and amorphous regions) (33). The ingestion of the MPs can thus change the 117 route of uptake compared to the dissolved form of the exogenous chemical from waterborne to 118 dietary exposure (34) and following desorption within an organisms' digestive system there is 119 potentially a greater level of chemical exposure (35). Accordingly, this so-called 'vector-effect' (3) has 120 been the subject of much research and speculation, but even the earliest investigations 121 demonstrated the varying nature of the vector phenomenon where MPs enhanced the bioavailability 122 and toxicity of co-contaminants (36,37), where the addition of MPs to the exposure scenario resulted 123 in negligible impacts (38), and cases where MPs reduced pollutant bioavailability (39).

124 The scientific literature in this area is too vast to comprehensively cover in this short review (see 125 recent reviews (5,14-18), however, recent descriptions of vector effects continue to vary. Two 126 approaches are used to determine vector effects, (i) to directly measure chemicals in tissues to 127 determine the influence of MPs to the exposure scenario and (ii) measure a toxicological marker of 128 exposure as an indicator of chemical bioavailability and biological reactivity. Numerous laboratory 129 studies have reported that exogenous contaminants sorbed to MPs are bioavailable and lead to a 130 measurable transfer into the tissue (40-42) and toxicological impacts based upon the assessment of 131 biomarkers (42–44), even if the tissue burdens did not correspondingly increase (45–47). Conversely, 132 several studies have demonstrated that at least for some measured endpoints, the role of MPs on 133 chemical-induced negative impacts is not significant (48–52). Using a novel 'feeding tube' method to 134 directly introduce polyethylene and polystyrene MPs loaded with PCB-153 into the digestive tract of 135 fish larvae showed no transfer of the PCB from MP into the tissue (53). In the exposures of Talitrus 136 saltator MPs were shown to carry HOCs into the tissue following ingestion, but when 137 uncontaminated MPs were fed to sand hoppers, then the MP scavenged the chemicals and reduced 138 the tissue burden (54). Thus, under some circumstances, MP ingestion can potentially perform a 139 "cleaning effect" (54,55). The transfer of PCBs from MPs under simulated gut fluid conditions was 140 demonstrated to be biphasic and reversible (55).

141

142 The combination of MPs and co-contaminants may be viewed as similar to the interactions within a 143 chemical mixture - independent or dependent action, or additivity, synergism or antagonism (56). 144 The joint exposure of MPs and the pharmaceutical triclosan to marine microalgae induced 145 antagonistic effects with increasing MP concentrations reducing the triclosan toxicity based on the 146 adsorption of the chemical to the plastic surface (57). However, the co-exposure of the marine 147 copepod Acartia tonsa to polyethylene microbeads and triclosan resulted in a relatively obscure 148 mixture effect known as potentiation in which the MP, without itself being toxic, enhanced the 149 toxicity of triclosan (56). Thus, even when assessed through a recognized framework designed to 150 disentangle the effects of single components within a mixture, the MP vector effect does not provide 151 consistent outcomes. Though little used in MP research, the analysis of mixtures may provide a 152 mechanistic insight into the individual roles of each competent within MP-co-contaminant 153 combination and further attention with this approach would be warranted.

154

155 2c. Focus on digestive physiology

156 Recognising that there are important differences between all the studies described in the preceding 157 sections that impact the outcome - choices relating to MP properties, co-contaminant, species, 158 experimental design, and biological endpoint (see Figure 1) – there remains disparity in descriptions 159 of the MPs a chemical carrier which goes beyond the interactions and sorptive behaviour in the test 160 media and needs to consider physiology, particularly that of the intestinal environment. If MP 161 ingestion is the assumed route of entry, then there are two possibilities for MPs and co-contaminants 162 to enter the gastrointestinal tract of aquatic animals - independently or with the co-contaminant 163 associated (sorbed) to the MP (58). In the gastrointestinal tract the fate of the MP and co-164 contaminant to remain independent, remain sorbed or desorb is largely driven by the gut lumen 165 environment. Perhaps the earliest study to investigate HOC desorption in simulated gut conditions 166 showed pH and temperature were important factors in determining desorption rates, suggesting that 167 warm blooded animals could be of greater threat of MP-facilitated HOC transfer, but desorption also 168 occurred in cold-blooded conditions representative of fish and invertebrates (59). Recent follow-up 169 studies have also demonstrated that both endogenous and exogenous chemicals separate from the 170 MP within intestinal and biological fluids and conditions (13,60,61).

171

However, the lumen of the gastrointestinal tract (GIT) is a dynamic environment that varies between
species and within species. For instance, the luminal pH of the polychaete worms *Lumbriculus*

174 variegatus and Arenicola marina are 5.4-6.5 and 6.8-7.2, respectively, whereas carnivorous fish, such 175 as rainbow trout (Oncorhynchus mykiss) exhibit a wider range pH 2.0-8.5 (62). In the latter, the GIT is 176 compartmentalised into different anatomical regions, with an acidic lumen in the stomach and alkali 177 lumen in the intestinal regions. The pH is a main driver for determining the partitioning of chemicals 178 onto the surface of MPs for ionisable organic chemicals (63) and dissolved metals (64,65), typically 179 with lower pH values causing less chemical to bind to the surface of the MPs. For fish, at least, the 180 variation in pH along the lumen of the gastrointestinal tract creates the potential for the cycling of 181 chemicals on and off the MP (55). Temperature, salinity and ionic strength have been shown to affect 182 the sorption behaviour of co-contaminants to MPs (15). However, determining the relative 183 contribution of each GIT parameter to the potential for vector effects and co-contaminant transfer is 184 difficult in vivo, but a greater understanding of the role of species-specific digestive physiology is 185 paramount to better understand the toxicological effects of MP and co-contaminant exposures.

186

187 3. Laboratory-to-field extrapolation of MP co-contaminant studies

188 The ecotoxicological consequences of MPs and co-contaminants have largely been studied within 189 laboratory settings. In extrapolating those findings to the natural world, two pertinent questions 190 need to be addressed: (i) do laboratory studies realistically reflect the complexity of MPs in the 191 environment and (ii) are MPs relevant chemical carriers compared to other potential sorbents? It is 192 now established that in the environment, MPs are neither just pristine nor just contaminated, but 193 rather exist in a continuum as a class of complex pollutants from different polymer types, shapes and 194 sizes, at different levels of environmental transformations, and which can leach or sorb a multitude 195 of chemicals (Figure 1) ((1,5,18,66). Despite this, most co-contaminant experimental studies employ 196 aspects that lack environmental relevance; the use of pristine MPs, single polymers and MP types 197 (e.g., the overuse of polystyrene spheres (67)) at levels above field concentrations coupled with 198 single pollutants, short equilibrium times or methods to artificially hasten sorption kinetics (5,33). 199 Thus, studies with different MP morphologies are needed to reflect environmental prevalence (68) 200 and as different types may exhibit different gut passage times which may affect chemical transfer 201 from MP to tissue. Natural ageing (i.e., weathering) of MPs increases their adsorption affinity 202 towards contaminants, but this parameter has seldom been considered in the effect assessments of 203 MPs. The weathering of plastics in environmental settings is affected by exposure to UV radiation 204 (sunlight), temperature shifts, humidity, and oxygen and ozone levels (69-71), and in turn the 205 weathering of MPs can further play an important role in the leachates released and toxicity to 206 organisms (20,72,73). Furthermore, as climatic conditions shift due to global change (e.g., lower pH,

increased temperature and fluctuating salinities) the impact of such parameters should be betterlinked to plastic pollution (66,74).

209

210 The aspect of relevance has been most comprehensively addressed by Koelmans et al. 2016 (22). 211 Briefly, the authors modelled analysis considered the whole mass of various compartments of the 212 ocean including plastics, and then in which compartment exogenous HOCs may preferentially reside 213 based upon partition coefficients. Ocean water would hold 98.3% of HOCs in the ocean and plastics 214 just 0.0002% - in last place of the nine compartments included in the model (22). Thus, when 215 assessing the relative importance of MPs as chemical carriers, other compartments namely food and 216 water, may be of greater importance as contaminant vectors, however, it is not possible to entirely 217 disregard the link between MP ingestion and chemical availability (18). Similarly, the transfer of 218 endogenous chemicals is not accounted for.

219

220 4. Expanding the microplastics universe

221 As the MP field progresses, new classes of anthropogenic particles are coming into focus and the 222 same questions regarding chemical transfer are being asked. Nanoplastics (defined as < 1 μ m by ISO 223 (75)) have been shown to be taken up by invertebrates (76,77) and translocated across the gastrointestinal membrane of fish in an ex-vivo gut sac model (78). Nano-sized particles have the 224 225 potential to achieve cellular internalization via endocytotic mechanism and with this exists the 226 possibility that endogenous and exogenous chemicals associated to nanoplastics may be carried into 227 the cell. Coupled with the greater biological reactivity at the nano-size, the overall hazard of 228 nanoplastics may be greater than MPs (79). However, clear demonstrations of this potential are 229 currently absent from the literature.

230

231 Concerns about plastic pollution and greater environmental sustainability have promoted 232 'bioplastics' as an alternative to conventional fossil-fuel based polymers. The term 'bioplastics' may 233 encompass both bio-based plastics made from renewable or natural sources (i.e. plant material) and 234 biodegradable plastics that are made from materials which can be subject to enzymatic degradation 235 of the polymeric matrix (20). Thus, whilst conventional wisdom would say that bioplastics are 236 designed to degrade faster than conventional plastics, there is specificity to the conditions of 237 degradation, such as the right medium (water, soil, compost), and the absence of such conditions 238 may result in a longer than expected residence time in the environment (80). Though generally

considered 'green' the bioplastic polyhydroxybutyrate (PHB) still contained a wide variety of exogenous chemicals and showed slight toxicity to sea urchin larvae (81). Also using PHB as a test bioplastic, Magara et al. (2019) (82) compared the effects of polyethylene and PHB MPs as a vector of fluoranthene to *Mytilus edulis* with the two polymers exhibiting similar minimal differences to fluoranthene-only exposures. Thus, whether such materials constitute toxicologically safer alternatives is not yet verified as the literature is limited.

245

246 Tire wear particles (TWP), tire and road wear particles (TRWP), recycled crumb rubber (RTC) and tire-247 repair-polished debris (TRD) are rubber-related additions to the MP field (83-85). Of these TWP is 248 perhaps the most discussed with estimates of release suggesting that TWP is a significant component 249 of MP pollution (86). The chemicals added to tires during manufacturing have been shown to readily 250 be released from the tire under laboratory conditions (87). This complex 'leachate' has been shown 251 to be toxic to a variety of aquatic organisms (87,88) with some specific chemicals now being 252 pinpointed as known toxic agents. For instance, 6PPD-quinone was responsible for the acute toxicity 253 of Pacific Northwest coho salmon observed in the field (89). Recent studies with TWP have focussed 254 on the particle and the leachate with several species ingesting TWP (84,90,91) and the two fractions 255 showing distinct toxicities (91,92). Thus, the role of the rubber particle delivering leachate in vivo 256 requires greater attention.

257

258 5. Conclusions

259 The role of MPs in effecting the bioavailability and toxicological consequences of endogenous and 260 exogenous co-contaminants has been a much-debated aspect of plastic pollution. There is a wealth 261 of in-depth literature on the subject (see reviews (2,5,15,16,18,19), but experimental studies often 262 display inconsistencies. This is not surprising since the delivery of chemicals by MPs is dependent on 263 multiple inter-connected factors (Figure 1) (5,47)). Thus, it remains difficult to judge whether MPs are 264 realistic carriers of chemicals and furthermore, based on modelled analysis, whether MPs are 265 relevant to study in this context given their relative contribution to oceanic mass compared to other 266 sorbents (22). The expansion of the field to include a greater range of particles, namely, nanoplastics, 267 TWP and 'bio-plastics', will increase focus to cellular-level vector effects, leachate-related toxicity 268 and 'benign-by-design', but future research should also consider the complex processes involved in 269 MP-facilitated chemical transfer (Figure 1) with greater attention needed for biological parameters. 270 Overall, greater environmental and physiological realism is needed to bridge the gap between the lab 271 and the real-world.

272

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274 All authors contributed to all aspects of this publication.

275

- 276 Declaration of Interests
- 277 The authors have no competing interests to declare.

278

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285 References

- Rochman CM, Brookson C, Bikker J, Djuric N, Earn A, Bucci K, et al. Rethinking microplastics as
 a diverse contaminant suite. Vol. 38, Environmental Toxicology and Chemistry. Wiley
 Blackwell; 2019. p. 703–11.
- Hahladakis JN, Velis CA, Weber R, Iacovidou E, Purnell P. An overview of chemical additives
 present in plastics: Migration, release, fate and environmental impact during their use,
 disposal and recycling. Vol. 344, Journal of Hazardous Materials. Elsevier B.V.; 2018. p. 179–
 99.
- Syberg K, Khan FR, Selck H, Palmqvist A, Banta GT, Daley J, et al. Microplastics: Addressing
 ecological risk through lessons learned. Environmental Toxicology and Chemistry. 2015 May
 1;34(5):945–53.
- Andrady AL. The plastic in microplastics: A review. Vol. 119, Marine Pollution Bulletin. Elsevier
 Ltd; 2017. p. 12–22.
- Khan FR, Patsiou D, Catarino AI. Pollutants Bioavailability and Toxicological Risk from
 Microplastics. In: Handbook of Microplastics in the Environment. Springer International
 Publishing; 2021. p. 1–40.
- Corcoran PL. Degradation of Microplastics in the Environment. In: Handbook of Microplastics
 in the Environment. Springer International Publishing; 2022. p. 531–42.
- Wu X, Liu P, Shi H, Wang H, Huang H, Shi Y, et al. Photo aging and fragmentation of
 polypropylene food packaging materials in artificial seawater. Water Research. 2021 Jan
 1;188.

306 8. Amaral-Zettler LA, Zettler ER, Mincer TJ. Ecology of the plastisphere. Vol. 18, Nature Reviews 307 Microbiology. Nature Research; 2020. p. 139–51. 308 9. Brander SM. Rethinking our chemical legacy and reclaiming our planet. Vol. 5, One Earth. Cell 309 Press; 2022. p. 316–9. 310 10. Phuong NN, Zalouk-Vergnoux A, Poirier L, Kamari A, Châtel A, Mouneyrac C, et al. Is there any 311 consistency between the microplastics found in the field and those used in laboratory 312 experiments? Vol. 211, Environmental Pollution. Elsevier Ltd; 2016. p. 111-23. 313 11. Du S, Zhu R, Cai Y, Xu N, Yap PS, Zhang Y, et al. Environmental fate and impacts of 314 microplastics in aquatic ecosystems: A review. Vol. 11, RSC Advances. Royal Society of 315 Chemistry; 2021. p. 15762-84. 316 12. Courtene-Jones W, Clark NJ, Fischer AC, Smith NS, Thompson RC. Ingestion of Microplastics by 317 Marine Animals. In: Andrady AL, editor. Plastics and the Ocean. Wiley; 2022. p. 349-66. 318 13. Bao ZZ, Chen ZF, Lu SQ, Wang G, Qi Z, Cai Z. Effects of hydroxyl group content on adsorption 319 and desorption of anthracene and anthrol by polyvinyl chloride microplastics. Science of the 320 Total Environment. 2021 Oct 10;790. 321 14. Hartmann NB, Rist S, Bodin J, Jensen LHS, Schmidt SN, Mayer P, et al. Microplastics as vectors 322 for environmental contaminants: Exploring sorption, desorption, and transfer to biota. Vol. 323 13, Integrated Environmental Assessment and Management. Wiley-Blackwell; 2017. p. 488-324 93. 325 Fred-Ahmadu OH, Bhagwat G, Oluyoye I, Benson NU, Ayejuyo OO, Palanisami T. Interaction of 15. 326 chemical contaminants with microplastics: Principles and perspectives. Vol. 706, Science of 327 the Total Environment. Elsevier B.V.; 2020. 328 16. Santos LHMLM, Rodríguez-Mozaz S, Barceló D. Microplastics as vectors of pharmaceuticals in 329 aquatic organisms - An overview of their environmental implications. Case Studies in 330 Chemical and Environmental Engineering. 2021 Jun 1;3. 331 17. Huang W, Song B, Liang J, Niu Q, Zeng G, Shen M, et al. Microplastics and associated 332 contaminants in the aquatic environment: A review on their ecotoxicological effects, trophic 333 transfer, and potential impacts to human health. Journal of Hazardous Materials. 2021 Mar 334 5;405. 335 18. Koelmans AA, Diepens NJ, Mohamed Nor NH. Weight of Evidence for the Microplastic Vector 336 Effect in the Context of Chemical Risk Assessment. In: Banks MS, editor. Microplastic in the 337 Environment: Pattern and Process. 2022. p. 155–97. 338 19. Hermabessiere L, Dehaut A, Paul-Pont I, Lacroix C, Jezequel R, Soudant P, et al. Occurrence and effects of plastic additives on marine environments and organisms: A review. Vol. 182, 339 340 Chemosphere. Elsevier Ltd; 2017. p. 781–93. 341 20. Curto M, le Gall M, Catarino AI, Niu Z, Davies P, Everaert G, et al. Long-term durability and 342 ecotoxicity of biocomposites in marine environments: A review. Vol. 11, RSC Advances. Royal 343 Society of Chemistry; 2021. p. 32917–41. 344 21. Kwon JH, Chang S, Hong SH, Shim WJ. Microplastics as a vector of hydrophobic contaminants: 345 Importance of hydrophobic additives. Vol. 13, Integrated Environmental Assessment and 346 Management. Wiley-Blackwell; 2017. p. 494-9.

347 22. Koelmans AA, Bakir A, Burton GA, Janssen CR. Microplastic as a Vector for Chemicals in the 348 Aquatic Environment: Critical Review and Model-Supported Reinterpretation of Empirical 349 Studies. Vol. 50, Environmental Science and Technology. American Chemical Society; 2016. p. 350 3315-26. 351 23. Bakir A, O'Connor IA, Rowland SJ, Hendriks AJ, Thompson RC. Relative importance of 352 microplastics as a pathway for the transfer of hydrophobic organic chemicals to marine life. 353 Environmental Pollution. 2016 Dec 1;219:56–65. 354 24. Kühn S, Booth AM, Sørensen L, van Oyen A, van Franeker JA. Transfer of Additive Chemicals 355 From Marine Plastic Debris to the Stomach Oil of Northern Fulmars. Frontiers in 356 Environmental Science. 2020 Aug 19;8. 357 25. Tanaka K, Watanuki Y, Takada H, Ishizuka M, Yamashita R, Kazama M, et al. In Vivo 358 Accumulation of Plastic-Derived Chemicals into Seabird Tissues. Current Biology. 2020 Feb 359 24;30(4):723-728.e3. 360 26. Syberg K, Knudsen CMH, Tairova Z, Khan FR, Shashoua Y, Geertz T, et al. Sorption of PCBs to 361 environmental plastic pollution in the North Atlantic Ocean: Importance of size and polymer 362 type. Case Studies in Chemical and Environmental Engineering. 2020 Sep 1;2. 363 27. Fred-Ahmadu OH, Tenebe IT, Ayejuyo OO, Benson NU. Microplastics and associated organic 364 pollutants in beach sediments from the Gulf of Guinea (SE Atlantic) coastal ecosystems. 365 Chemosphere. 2022 Jul 1;298. 366 28. Vedolin MC, Teophilo CYS, Turra A, Figueira RCL. Spatial variability in the concentrations of 367 metals in beached microplastics. Marine Pollution Bulletin. 2018 Apr 1;129(2):487–93. 368 29. Carbery M, MacFarlane GR, O'Connor W, Afrose S, Taylor H, Palanisami T. Baseline analysis of 369 metal(loid)s on microplastics collected from the Australian shoreline using citizen science. 370 Marine Pollution Bulletin. 2020 Mar 1;152. 371 30. Santana-Viera S, Montesdeoca-Esponda S, Torres-Padrón ME, Sosa-Ferrera Z, Santana-372 Rodríguez JJ. An assessment of the concentration of pharmaceuticals adsorbed on 373 microplastics. Chemosphere. 2021 Mar 1;266. 374 31. Wagstaff A, Lawton LA, Petrie B. Polyamide microplastics in wastewater as vectors of cationic 375 pharmaceutical drugs. Chemosphere. 2022 Feb 1;288. 376 32. McDougall L, Thomson L, Brand S, Wagstaff A, Lawton LA, Petrie B. Adsorption of a diverse 377 range of pharmaceuticals to polyethylene microplastics in wastewater and their desorption in 378 environmental matrices. Science of the Total Environment. 2022 Feb 20;808. 379 33. Velez JFM, Shashoua Y, Syberg K, Khan FR. Considerations on the use of equilibrium models 380 for the characterisation of HOC-microplastic interactions in vector studies. Vol. 210, 381 Chemosphere. Elsevier Ltd; 2018. p. 359-65. 382 34. Khan FR, Syberg K, Shashoua Y, Bury NR. Influence of polyethylene microplastic beads on the 383 uptake and localization of silver in zebrafish (Danio rerio). Environmental Pollution. 2015 Jul 384 4;206:73-9. 385 35. Sørensen L, Rogers E, Altin D, Salaberria I, Booth AM. Sorption of PAHs to microplastic and 386 their bioavailability and toxicity to marine copepods under co-exposure conditions. 387 Environmental Pollution. 2020 Mar 1;258.

- 388 36. Oliveira M, Ribeiro A, Hylland K, Guilhermino L. Single and combined effects of microplastics
 and pyrene on juveniles (0+ group) of the common goby Pomatoschistus microps (Teleostei,
 Gobiidae). Ecological Indicators. 2013;34:641–7.
- 37. Rochman CM, Kurobe T, Flores I, Teh SJ. Early warning signs of endocrine disruption in adult
 fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the
 marine environment. Science of the Total Environment. 2014 Sep 15;493:656–61.
- 38. Besseling E, Wegner A, Foekema EM, van den Heuvel-Greve MJ, Koelmans AA. Effects of
 microplastic on fitness and PCB bioaccumulation by the lugworm Arenicola marina (L.).
 Environmental Science and Technology. 2013 Jan 2;47(1):593–600.
- 397 39. Chua EM, Shimeta J, Nugegoda D, Morrison PD, Clarke BO. Assimilation of polybrominated
 398 diphenyl ethers from microplastics by the marine amphipod, allorchestes compressa.
 399 Environmental Science and Technology. 2014 Jul 15;48(14):8127–34.
- 40. González-Soto N, Hatfield J, Katsumiti A, Duroudier N, Lacave JM, Bilbao E, et al. Impacts of
 401 dietary exposure to different sized polystyrene microplastics alone and with sorbed
 402 benzo[a]pyrene on biomarkers and whole organism responses in mussels Mytilus
 403 galloprovincialis. Science of the Total Environment. 2019 Sep 20;684:548–66.
- 404 41. O'Donovan S, Mestre NC, Abel S, Fonseca TG, Carteny CC, Cormier B, et al. Ecotoxicological
 405 effects of chemical contaminants adsorbed to microplastics in the Clam Scrobicularia plana.
 406 Front Mar Sci. 2018 Apr 26;5(APR).
- 407 42. Lu K, Qiao R, An H, Zhang Y. Influence of microplastics on the accumulation and chronic toxic
 408 effects of cadmium in zebrafish (Danio rerio). Chemosphere. 2018 Jul 1;202:514–20.
- 409 43. Rainieri S, Conlledo N, Larsen BK, Granby K, Barranco A. Combined effects of microplastics and
 410 chemical contaminants on the organ toxicity of zebrafish (Danio rerio). Environmental
 411 Research. 2018 Apr 1;162:135–43.
- 44. Batel A, Borchert F, Reinwald H, Erdinger L, Braunbeck T. Microplastic accumulation patterns
 and transfer of benzo[a]pyrene to adult zebrafish (Danio rerio) gills and zebrafish embryos.
 Environmental Pollution. 2018 Apr 1;235:918–30.
- 45. Granby K, Rainieri S, Rasmussen RR, Kotterman MJJ, Sloth JJ, Cederberg TL, et al. The
 416 influence of microplastics and halogenated contaminants in feed on toxicokinetics and gene
 417 expression in European seabass (Dicentrarchus labrax). Environmental Research. 2018 Jul
 418 1;164:430–43.
- 46. Magara G, Elia AC, Syberg K, Khan FR. Single contaminant and combined exposures of
 420 polyethylene microplastics and fluoranthene: accumulation and oxidative stress response in
 421 the blue mussel, Mytilus edulis. Journal of Toxicology and Environmental Health Part A:
 422 Current Issues. 2018 Aug 18;81(16):761–73.
- 423 47. Ašmonaitė G, Tivefälth M, Westberg E, Magnér J, Backhaus T, Carney Almroth B. Microplastics
 424 as a Vector for Exposure to Hydrophobic Organic Chemicals in Fish: A Comparison of Two
 425 Polymers and Silica Particles Spiked With Three Model Compounds. Frontiers in
 426 Environmental Science. 2020 Jul 3;8.
- 42748.Beiras R, Tato T. Microplastics do not increase toxicity of a hydrophobic organic chemical to428marine plankton. Marine Pollution Bulletin. 2019 Jan 1;138:58–62.

429 49. Beiras R, Muniategui-Lorenzo S, Rodil R, Tato T, Montes R, López-Ibáñez S, et al. Polyethylene 430 microplastics do not increase bioaccumulation or toxicity of nonylphenol and 4-MBC to 431 marine zooplankton. Science of the Total Environment. 2019 Nov 20;692:1-9. 432 50. Wen B, Jin SR, Chen ZZ, Gao JZ, Liu YN, Liu JH, et al. Single and combined effects of 433 microplastics and cadmium on the cadmium accumulation, antioxidant defence and innate 434 immunity of the discus fish (Symphysodon aequifasciatus). Environmental Pollution. 2018 Dec 435 1;243:462-71. 436 51. Schmieg H, Burmester JKY, Krais S, Ruhl AS, Tisler S, Zwiener C, et al. Interacting effects of 437 polystyrene microplastics and the antidepressant amitriptyline on early life stages of brown 438 trout (Salmo trutta f. fario). Water (Switzerland). 2020 Sep 1;12(9). 439 52. Guven O, Bach L, Munk P, Dinh K v., Mariani P, Nielsen TG. Microplastic does not magnify the 440 acute effect of PAH pyrene on predatory performance of a tropical fish (Lates calcarifer). 441 Aquatic Toxicology. 2018 May 1;198:287-93. 442 53. Norland S, Vorkamp K, Bogevik AS, Koelmans AA, Diepens NJ, Burgerhout E, et al. Assessing 443 microplastic as a vector for chemical entry into fish larvae using a novel tube-feeding 444 approach. Chemosphere. 2021 Feb 1;265. 445 54. Scopetani C, Cincinelli A, Martellini T, Lombardini E, Ciofini A, Fortunati A, et al. Ingested 446 microplastic as a two-way transporter for PBDEs in Talitrus saltator. Environmental Research. 447 2018 Nov 1;167:411-7. 448 55. Mohamed Nor NH, Koelmans AA. Transfer of PCBs from Microplastics under Simulated Gut 449 Fluid Conditions Is Biphasic and Reversible. Environmental Science and Technology. 2019; 450 56. Syberg K, Nielsen A, Khan FR, Banta GT, Palmqvist A, Jepsen PM. Microplastic potentiates 451 triclosan toxicity to the marine copepod Acartia tonsa (Dana). Journal of Toxicology and 452 Environmental Health - Part A: Current Issues. 2017 Dec 17;80(23-24):1369-71. 453 57. Zhu Z lin, Wang S chun, Zhao F fei, Wang S guang, Liu F fei, Liu G zhou. Joint toxicity of 454 microplastics with triclosan to marine microalgae Skeletonema costatum. Environmental 455 Pollution. 2019 Mar 1;246:509–17. 456 58. Khan FR, Boyle D, Chang E, Bury NR. Do polyethylene microplastic beads alter the intestinal 457 uptake of Ag in rainbow trout (Oncorhynchus mykiss)? Analysis of the MP vector effect using 458 in vitro gut sacs. Environmental Pollution. 2017;231:200-6. 459 59. Bakir A, Rowland SJ, Thompson RC. Enhanced desorption of persistent organic pollutants from 460 microplastics under simulated physiological conditions. Environmental Pollution. 461 2014;185:16-23. 462 60. Coffin S, Huang GY, Lee I, Schlenk D. Fish and Seabird Gut Conditions Enhance Desorption of 463 Estrogenic Chemicals from Commonly-Ingested Plastic Items. Environmental Science and 464 Technology. 2019 Apr 16;53(8):4588–99. 465 61. Wu P, Tang Y, Jin H, Song Y, Liu Y, Cai Z. Consequential fate of bisphenol-attached PVC 466 microplastics in water and simulated intestinal fluids. Environmental Science and 467 Ecotechnology. 2020 Apr 1;2.

468 469 470	62.	van der Zande M, Jemec Kokalj A, Spurgeon DJ, Loureiro S, Silva P v., Khodaparast Z, et al. The gut barrier and the fate of engineered nanomaterials: A view from comparative physiology. Environmental Science: Nano. 2020 Jul 1;7(7):1874–98.
471 472 473	63.	Zhang H, Wang J, Zhou B, Zhou Y, Dai Z, Zhou Q, et al. Enhanced adsorption of oxytetracycline to weathered microplastic polystyrene: Kinetics, isotherms and influencing factors. Environmental Pollution. 2018 Dec 1;243:1550–7.
474 475 476	64.	Ahechti M, Benomar M, el Alami M, Mendiguchía C. Metal adsorption by microplastics in aquatic environments under controlled conditions: exposure time, pH and salinity. International Journal of Environmental Analytical Chemistry. 2022;102(5):1118–25.
477 478 479	65.	Wang Y, Liu C, Wang F, Sun Q. Behavior and mechanism of atrazine adsorption on pristine and aged microplastics in the aquatic environment: Kinetic and thermodynamic studies. Chemosphere. 2022 Apr 1;292.
480 481 482 483	66.	Catarino AI, Asselman J, Niu Z, Everaert G. Micro- and nanoplastics effects in a multiple stressed marine environment. Journal of Hazardous Materials Advances [Internet]. 2022 Aug;7:100119. Available from: https://linkinghub.elsevier.com/retrieve/pii/S2772416622000754
484 485 486 487	67.	Coffin S, Bouwmeester H, Brander S, Damdimopoulou P, Gouin T, Hermabessiere L, et al. Development and application of a health-based framework for informing regulatory action in relation to exposure of microplastic particles in California drinking water. Microplastics and Nanoplastics. 2022 Dec;2(1).
488 489 490	68.	Suaria G, Achtypi A, Perold V, Lee JR, Pierucci A, Bornman TG, et al. O C E A N O G R A P H Y Microfibers in oceanic surface waters: A global characterization [Internet]. Vol. 6, Sci. Adv. 2020. Available from: https://www.science.org
491 492 493	69.	Bhagat K, Barrios AC, Rajwade K, Kumar A, Oswald J, Apul O, et al. Aging of microplastics increases their adsorption affinity towards organic contaminants. Chemosphere. 2022 Jul 1;298.
494 495 496	70.	Liu G, Zhu Z, Yang Y, Sun Y, Yu F, Ma J. Sorption behavior and mechanism of hydrophilic organic chemicals to virgin and aged microplastics in freshwater and seawater. Environmental Pollution. 2019 Mar 1;246:26–33.
497 498 499	71.	Cormier B, Gambardella C, Tato T, Perdriat Q, Costa E, Veclin C, et al. Chemicals sorbed to environmental microplastics are toxic to early life stages of aquatic organisms. Ecotoxicology and Environmental Safety. 2021 Jan 15;208.
500 501	72.	Bejgarn S, MacLeod M, Bogdal C, Breitholtz M. Toxicity of leachate from weathering plastics: An exploratory screening study with Nitocra spinipes. Chemosphere. 2015 Aug 1;132:114–9.
502 503 504	73.	Jiang X, Lu K, Tunnell JW, Liu Z. The impacts of weathering on concentration and bioaccessibility of organic pollutants associated with plastic pellets (nurdles) in coastal environments. Marine Pollution Bulletin. 2021 Sep 1;170.
505 506 507	74.	Ford H v., Jones NH, Davies AJ, Godley BJ, Jambeck JR, Napper IE, et al. The fundamental links between climate change and marine plastic pollution. Vol. 806, Science of the Total Environment. Elsevier B.V.; 2022.

508 75. Allan J, Belz S, Hoeveler A, Hugas M, Okuda H, Patri A, et al. Regulatory landscape of 509 nanotechnology and nanoplastics from a global perspective. Regulatory Toxicology and 510 Pharmacology. 2021 Jun 1;122. 511 76. Lahive E, Cross R, Saarloos AI, Horton AA, Svendsen C, Hufenus R, et al. Earthworms ingest 512 microplastic fibres and nanoplastics with effects on egestion rate and long-term retention. 513 Science of the Total Environment. 2022 Feb 10;807. 514 77. Redondo-Hasselerharm PE, Vink G, Mitrano DM, Koelmans AA. Metal-doping of nanoplastics 515 enables accurate assessment of uptake and effects onGammarus pulex. Environmental 516 Science: Nano. 2021 Jun 1;8(6):1761-70. 517 78. Clark NJ, Khan FR, Mitrano DM, Boyle D, Thompson RC. Demonstrating the translocation of 518 nanoplastics across the fish intestine using palladium-doped polystyrene in a salmon gut-sac. 519 Environment International. 2022 Jan 15;159. 520 79. Koelmans AA, Besseling E, Shim WJ. Nanoplastics in the Aquatic Environment. Critical Review. 521 In: Bergman M, Gutow L, Klages M, editors. Marine Anthropogenic Litter. Cham: Springer 522 International Publishing; 2015. p. 325–40. 523 80. Tong H, Zhong X, Duan Z, Yi X, Cheng F, Xu W, et al. Micro- and nanoplastics released from 524 biodegradable and conventional plastics during degradation: Formation, aging factors, and 525 toxicity. Science of the Total Environment. 2022 Aug 10;833. 526 81. Uribe-Echeverría T, Beiras R. Acute toxicity of bioplastic leachates to Paracentrotus lividus sea 527 urchin larvae. Marine Environmental Research. 2022 Apr 1;176. 528 82. Magara G, Khan FR, Pinti M, Syberg K, Inzirillo A, Elia AC. Effects of combined exposures of 529 fluoranthene and polyethylene or polyhydroxybutyrate microplastics on oxidative stress 530 biomarkers in the blue mussel (Mytilus edulis). Journal of Toxicology and Environmental 531 Health - Part A: Current Issues. 2019 May 19;82(10):616–25. 532 83. Wagner S, Hüffer T, Klöckner P, Wehrhahn M, Hofmann T, Reemtsma T. Tire wear particles in 533 the aquatic environment - A review on generation, analysis, occurrence, fate and effects. Vol. 534 139, Water Research. Elsevier Ltd; 2018. p. 83–100. 535 84. Halle LL, Palmqvist A, Kampmann K, Khan FR. Ecotoxicology of micronized tire rubber: Past, 536 present and future considerations. Vol. 706, Science of the Total Environment. Elsevier B.V.; 2020. 537 538 85. Luo Z, Zhou X, Su Y, Wang H, Yu R, Zhou S, et al. Environmental occurrence, fate, impact, and 539 potential solution of tire microplastics: Similarities and differences with tire wear particles. 540 Vol. 795, Science of the Total Environment. Elsevier B.V.; 2021. 541 86. Boucher J, Friot D. Primary microplastics in the oceans: A global evaluation of sources. IUCN 542 International Union for Conservation of Nature; 2017. 543 87. Marwood C, McAtee B, Kreider M, Ogle RS, Finley B, Sweet L, et al. Acute aquatic toxicity of 544 tire and road wear particles to alga, daphnid, and fish. Ecotoxicology. 2011;20(8):2079-89. 545 88. Wik A, Nilsson E, Källqvist T, Tobiesen A, Dave G. Toxicity assessment of sequential leachates 546 of tire powder using a battery of toxicity tests and toxicity identification evaluations. 547 Chemosphere. 2009;77(7):922-7.

Tian Z, Zhao H, Peter KT, Gonzalez M, Wetzel J, Wu C, et al. A ubiquitous tire rubber–derived
chemical induces acute mortality in coho salmon. Science (1979). 2021 Jan 8;371(6525):185–
9.

- 90. Redondo-Hasselerharm PE, de Ruijter VN, Mintenig SM, Verschoor A, Koelmans AA. Ingestion
 and Chronic Effects of Car Tire Tread Particles on Freshwater Benthic Macroinvertebrates.
 Environmental Science and Technology. 2018 Dec 4;52(23):13986–94.
- 554 91. Khan FR, Halle LL, Palmqvist A. Acute and long-term toxicity of micronized car tire wear
 555 particles to Hyalella azteca. Aquatic Toxicology. 2019 Aug 1;213.
- Halle LL, Palmqvist A, Kampmann K, Jensen A, Hansen T, Khan FR. Tire wear particle and
 leachate exposures from a pristine and road-worn tire to Hyalella azteca: Comparison of
 chemical content and biological effects. Aquatic Toxicology. 2021 Mar 1;232.
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560 Figure Legends

561 Figure 1

562 Schematic diagram of the complexity involved in the association of MPs with co-contaminants. 563 Starting at the production stage where a range of endogenous chemical "additives" (schematic 564 aromatic rings in red) are incorporated into the polymeric resin (green) (I). MPs entering or produced 565 through breakdown in the environment are a complex suite of physico-chemical properties (II) which 566 are subject to the environmental transformations (III). MPs are known to sorb of exogenous 567 pollutants, such as hydrophobic organic pollutants (schematic aromatic rings, black) and metals (Me⁺ and Me²⁺ black) (IV) and be ingested by biota (V). Within the digestive system endogenous additives 568 (red) leach out of the MP and sorbed exogenous pollutants (black) desorb. The released chemicals 569 570 may then be available for uptake (VI). The onset of toxicological outcomes will depend on the further 571 transport of the chemicals to sites of biological activity and the ecotoxicological assessments of 572 effects will depend on experimental choices made (VII). The complexity and array of variables 573 presented here highlights why the role of MPs as chemical carriers is still debated. Schematic based 574 on Khan et al., 2021 (5).

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