PEARL

Faculty of Science and Engineering

School of Biological and Marine Sciences

2022-05-18

Impacts of Artificial Light at Night (ALAN) in marine ecosystems a review

Marangoni, L

http://hdl.handle.net/10026.1/19086

10.1111/gcb.16264 Global Change Biology Wiley

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Please cite only the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author.

SCHOLARONE[™] Manuscripts

1 Impacts of Artificial Light at Night (ALAN) in marine ecosystems – a review

2

3 Abstract

4 The globally widespread adoption of Artificial Light at Night (ALAN) began in the mid-20th 5 century. Yet, it is only in the last decade that a renewed research focus has emerged into its impacts on ecological and biological processes in the marine environment that are guided by natural 6 intensities, moon phase, natural light and dark cycles and daily light spectra alterations. The field 7 8 has diversified rapidly from one restricted to impacts on a handful of vertebrates, to one in which impacts have been quantified across a broad array of marine and coastal habitats and species. Here 9 10 we review the current understanding of ALAN impacts in diverse marine ecosystems. The review 11 presents the current state of knowledge across key marine and coastal ecosystems (sandy and 12 rocky shores, coral reefs and pelagic) and taxa (birds and sea turtles), introducing how ALAN can 13 mask seabirds and sea turtles navigation, cause changes in animals predation patterns and failure of coral spawning synchronization, as well as inhibition of zooplankton Diel Vertical Migration. 14 Mitigation measures are recommended, however, while strategies for mitigation were easily 15 16 identified, barriers to implementation are poorly understood. Finally, we point out knowledge gaps 17 that if addressed would aid in the prediction and mitigation of ALAN impacts in the marine realm.

18

Laura F.B. Marangoni^{1*}, Thomas Davies^{2*}, Tim Smyth³, Airam Rodríguez^{4,5,6}, Mark
 Hamann⁷, Cristian Duarte⁸, Kellie Pendoley⁹, Jørgen Berge^{10,11,12}, Elena Maggi¹³, Oren Levy¹⁴

¹Smithsonian Tropical Research Institute, Smithsonian Institution, Ciudad de Panamá,
 0843-03092, Panamá.

²School of Biological and Marine Sciences, University of Plymouth, Drake Circus,
 Plymouth, Devon, United Kingdom, PL4 8AA

25 ³*Plymouth Marine Laboratory, Prospect Place, Plymouth, Devon, UK, PL1 3DH*

⁴Grupo de Ornitología e Historia Natural de las islas Canarias, GOHNIC, Canary
 Islands, Spain.

⁵Terrestrial Ecology Group, Department of Ecology, Universidad Autónoma de Madrid,
 Madrid, Spain.

⁶Centro de Investigación en Biodiversidad y Cambio Global (CIBC-UAM), Universidad
 Autónoma de Madrid, Madrid, Spain.

⁷College of Science and Engineering, Marine Biology, James Cook University, Townsville,
 Australia

⁸Departamento de Ecología y Biodiversidad, Facultad de Ciencias de la Vida,
 Universidad Andres Bello, Santiago, Chile.

36	⁹ Pendoley	Environmental	Pty Ltd,	12A	Pitt	Way,	Booragoon	Western	Australia,	6154,
37	Australia									

- ¹⁰ UiT The Arctic University of Norway, Faculty for Biosciences, Fisheries and Economics,
 Department for Arctic and Marine Biology. 9037 Tromsø, Norway
- 41 ¹¹ University Centre in Svalbard, Pb 156, N-9171 Longyearbyen, Norway

¹² Centre of Autonomous Marine Operations and Systems, Dept of Biology and
 Technology, Norwegian University of Science and Technology, N-7491, Trondheim, Norway
 45

¹³ Dip. di Biologia, CoNISMa, Università di Pisa, 56126 Italy.

¹⁴ Mina and Everard Goodman Faculty of Life Sciences, Bar-Ilan University, Ramat Gan,
52900, Israel.

- 49 *These authors contributed equally to this work.
- 50

42

46

51 **1. Introduction**

Artificial Light at Night (ALAN) is a widespread, pervasive, and expanding form of pollution (Gaston et al., 2021) that has come to be recognized as a major 21st century global change issue (Davies & Smyth, 2018). Its impacts span the biological hierarchy ranging from those on organism physiology through to changes in the composition of ecological communities (Sanders et al., 2021). It's now broadly accepted that ALAN has been reshaping nature for more than a century.

While research into the prevalence and impacts of ALAN in marine ecosystems has somewhat 57 58 lagged behind terrestrial, the last five years have seen a dramatic advance in our understanding. We now know that at least 22% of coastal regions are exposed to ALAN (Davies et al., 2014), 59 and the light from cities is sufficient to elicit biological responses in animals on the seafloor in 60 adjacent habitats (Davies et al., 2020, Ayalon et al., 2021a). 1.9 million km² of the world's coastal 61 seas are exposed to ALAN at 1m depth, 1.6 million km² at 10m depth, and 840,000 km² at 20m 62 depth (Fig. 1A) (Smyth et al., 2021). The most exposed regions include the Mediterranean (Fig. 63 1B), the Red Sea and Persian Gulf (Fig. 1C), and the seas of South-East Asia (Fig. 1D). ALAN is 64 even prevalent across those areas of the ocean most valued by humanity, with 20% of the world's 65 contiguous Marine Protected Areas exposed across 100% of their range (Davies et al., 2016). 66

67 The potential for ALAN to impact the wide array of organisms, processes, and habitats in the 68 sea for which light cycles are critical had remained largely unexplored until recently (Longcore & Rich, 2004; Davies et al., 2014). These include: diel vertical migrations (Berge et al., 2020) -69 70 the largest migration of biomass on the planet (Hayes, 2003); coral spawning (Ayalon et al., 71 2021a), and symbiosis (Ayalon et al., 2021b) – which are key for the maintenance of coral reefs; 72 consumer-resource interactions (Bolton et al., 2017; Underwood et al., 2017; Maggi et al., 2020a) 73 that are known to drive top down structuring of marine ecosystems (Paine, 1966); migrations and orientation of marine organisms - critical for their survival (Navarro-Barranco & Hughes, 2015; 74 75 Torres et al., 2020); and the recruitment of sessile invertebrate larvae into marine habitats (Davies 76 et al., 2015; Lynn et al., 2021b), (Fig 2). All these processes are fundamental to the health of

marine ecosystems, and all are known to depend on the cycles, spectra or intensity of sun or 77 78 moonlight.

79 Here, we bring together recognized experts in marine ALAN across habitats and taxonomic groups to review these recent advances, with the aim of providing a gateway to research in the 80 81 field. First, we review progress in sandy beaches, rocky intertidal shores, shallow water coral 82 reefs, and pelagic environments (Fig. 2). Second, we evaluate the current state of litigation and management options available to conservation practitioners. Finally, we identify key knowledge 83

84 gaps and highlight key questions for future research.

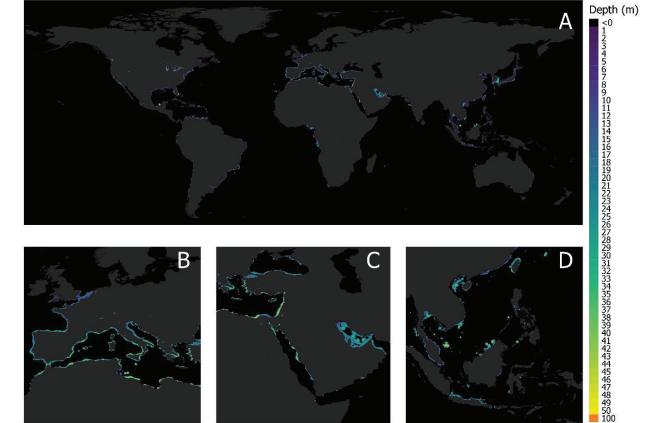




Figure 1. The depth of biologically important artificial light at night (ALAN) (Zc): around the 86 world's coastlines (A); in the Mediterranean and Northeast Atlantic (B); in the Black Sea, the 87 Red Sea and the Persian Gulf (C); and in the Gulf of Thailand, Andaman Sea, South China Sea 88 89 and the Java Sea (D). The legend inset details the depths (m) to which biologically important ALAN penetrates the sea. The data is derived from the relationship between The New World 90 91 Atlas of Artificial Night Sky Brightness (Falchi et al., 2016) and sea surface irradiances (Davies et al., 2020), coupled with the monthly climatologies of globally inherent optical water 92 93 properties and validated against in situ data collected from the Persian Gulf (Tamir et al., 2017). "Biologically important ALAN" is defined as the minimum irradiances of white light that elicit 94 diel vertical migration in female Calanus copepods (Båtnes et al., 2013) [see Smyth et al. 95 (2021) for further details]. Maps are representative of average ALAN penetration over a typical 96 97 vear. Full dataset is available to download from 98 https://doi.pangaea.de/10.1594/PANGAEA.929749

99 2. Biological effects of ALAN in the marine biota

100 In this section we have compiled the most relevant information obtained on key species belonging to different marine habitats across sandy beaches (including sea turtles), rocky intertidal shores, 101 shallow water coral reefs, and pelagic environments. A special section was created for seabirds 102 103 due to their high mobility and therefore presence in different marine realms. Despite the research gaps and limitations - to name a few e.g., lack of multistressor experiments including ALAN as a 104 factor, and difficulties to acquire data to define more precisely ecological relevant light intensities 105 to be tested (see Box 2) (Aulsebrook et al. 2022) - ALAN can be recognized as major sensory 106 107 pollutant of concern due it's obvious and widespread effects on pathways associated with natural circadian regulations in the marine biota. It is also important to note that experimental approaches 108 using high ALAN levels (not considered environmental realistic) in many of the short-term 109 110 experiments presented here it's an important step to understanding the mechanisms and long-term 111 effects of the chronic disturbance caused by ALAN.





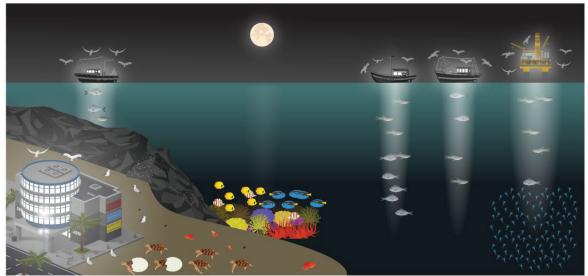


Figure 2. (A) Different marine environments not affected by Artificial Light Pollution at Night 113 (ALAN), and (B) marine environments under the potential impacts of ALAN: i) Sandy beaches 114 effects on invertebrate species day-night activity rhythms and biodiversity, effects in the on-beach 115 116 orientation of adults and hatchling turtles, and seabirds fledgling grounded by ALAN; ii) Rocky 117 intertidal shores - influence in metabolic activity/behavior of primary producers, sessile and 118 mobile animals; iii) Shallow water coral reefs – effects on gametogenesis and the synchronization 119 of gamete release in prominent coral species, and negative impacts over fish reproduction, iv) 120 Pelagic environment - inhibition of vertically migrating zooplankton, and disorientation and mortality of seabirds. 121

122

- 123 2.1. Sandy beach ecosystems
- 124

125 Many sandy beach species are known for their day/night activity rhythms controlled by

natural light cycles. Consequently, the dramatic ongoing expansion of ALAN in these
 ecosystems is expected to exert a significant effect on sandy beach biodiversity.

127 128



129 130

Figure 3. Los Choros Sandy beach, coast of Coquimbo, North of Chile (Credit; JosuéNavarrete).

133

Exposed sandy beach ecosystems represent over 80% of the ice-free coastline (Bascom, 1980). These ecosystems are considered highly valuable from an ecologic, economic and cultural points of view (King & Symes, 2004; Pendleton et al., 2006; McLachlan et al., 2013). However, the growing urbanization and the increase of human population near sandy shores has become a threat to these ecosystems (Jaramillo et al., 2021). One of the most important environmental stressors associated with the urbanization of sandy beaches is ALAN (Gonzalez et al., 2014; Schlacher et al., 2016).

Sandy beaches support an abundant and diverse fauna, being crustaceans (mainly talitrid amphipods, cirolanid and oniscoid isopods and hippid crabs), polychaetes and bivalves the main taxonomic groups (Brown & McLachlan, 1990; Jaramillo et al., 2003). Several sandy beach species such as amphipods, isopods, and insects are known for their dynamic day/night activity rhythms (Fallaci et al., 1999, 2002; Kennedy et al., 2000, Jaramillo et al., 2003), which are

Global Change Biology

controlled by natural geophysical cycles of day and night, and moonlight (Ugolini et al., 2003; 146 Jaramillo et al., 2003; Meschini et al., 2008). For example, these invertebrates rely on darkness to 147 use natural light signals and return to their burrows after down shore migrations (Fallaci et al., 148 2002). Consequently, the ongoing expansion of ALAN in these ecosystems is expected to exert 149 150 both a significant and negative effect on sandy beach biodiversity (e.g., Luarte et al., 2016; Duarte 151 et al., 2019). However, despite growing concerns on the impacts of ALAN, studies evaluating its effects on sandy beach organisms, other than sea turtles (e.g., Rivas et al., 2015; Dimiatris et al., 152 2018, see specific section 2.5. for this group), remain very scarce (e.g., Quintanilla-Ahumada et 153 al., 2021; Lynn et al., 2021a). The studies carried out so far have evaluated the effects of ALAN 154 on locomotor activity, feeding rates, absorption efficiency and growth rate, physiology, 155 distribution, and abundance of different species of invertebrates (see below). 156

One of the first studies evaluating the ALAN effects on locomotor activity of sandy beach 157 animals was conducted by Bregazzy & Naylor (1972). Those authors showed that in laboratory 158 159 conditions ALAN (~ 200 lux) almost entirely inhibited the locomotor activity of the talitrid amphipod *Talitrus saltator*. Interestingly, these individuals recovered their locomotor activity 160 pattern immediately after the light was removed. Similar results were registered by Luarte et al. 161 162 (2016), who using laboratory experiments showed that the locomotor activity of the talitrid 163 amphipod Orchestoidea tuberculata was significantly reduced in the presence of ALAN (60 lux). Similarly, Duarte et al. (2019) found that in laboratory conditions (120 lux), ALAN reduced the 164 locomotor activity of the oniscoid isopod *Tylus spinulosus*. More recently, Lynn et al. (2021a,b) 165 also found that ALAN (80 lux) disrupted the locomotor activity of the talitrid amphipod 166 Americorchestia longicornis. Consistent with the results reported by Bregazzy & Naylor (1972), 167 168 A. longicornis was found to recover its natural rhythm of activity shortly after ALAN was removed from the system (Lynn et al., 2021a). By contrast, Fanini et al. (2016) found that the locomotor 169 activity of the amphipod *Platorchestia smithi* was similar in a beach exposed to ALAN with 170 171 respect to another that was not exposed. However, this last study was correlational and should be 172 considered with caution. More recently 0.2 lx white lighting lower in brightness than a full moon (equivalent to artificial sky glow) has been demonstrated to reduce locomotor activity and 173 disorientate the migration behavior of Talitrus saltator (Torres et al. 2020). 174

175 Only a scarce number of studies have evaluated ALAN effects on aspects such as feeding rates, absorption efficiency and growth rates (Luarte et al., 2016; Lynn et al., 2021a; Quintanilla-176 Ahumada et al., 2022). The presence of ALAN reduced the consumption rate, absorption 177 efficiency and growth rate in the amphipods O. tuberculata (Luarte et al., 2016) and A. longicornis 178 (Lynn et al., 2021a), meanwhile ALAN (from 0 to 100 lx) did not affect T. spinulosus's growth 179 rate but increased its absorption efficiency (Quintanilla-Ahumada et al., 2022). The absorption 180 181 efficiency results should be considered with precaution, because of methodological restrictions, 182 as the animals had to be maintained without sand, which, eventually, could modify the ALAN effects on this biological trait). The RNA:DNA ratio is a relatively new indicator of the 183 physiological or nutritional condition of organisms (Buckley et al., 1999; Chícharo & Chícharo, 184 185 2008). Recent studies by Quintanilla-Ahumada et al. (2021, 2022), used this molecular tool to evaluate ALAN effects in sandy beach ecosystems, found that RNA:DNA ratio in the insect 186 Phalerisida maculata and in the isopod T. spinulosus, declined in the presence of ALAN, 187 188 indicating detrimental physiological effects. ALAN also shows important effects on the 189 abundance and distribution of sandy beach organisms. González et al. (2014) applied correlative

190 analyses and found that the abundance of the insect P. maculata was negatively related with the night sky quality (an indirect indicator of ALAN). Duarte et al. (2019) registered that the 191 192 distribution of T. spinulosus was modified by ALAN (120 lux and less), with individuals avoiding 193 the lit areas and therefore restricting their habitat availability. Importantly, ALAN effects 194 decreased with increasing distance from the light sources (Duarte et al., 2019). Despite the work 195 conducted this far, direct evidence of ALAN effects on sandy beach organisms remains limited 196 only to studies focusing on this stressor upon single species (e.g., Luarte et al., 2016; Duarte et 197 al., 2019).

198 It is important to note that the light intensities used in some of these studies were those recorded under the light source (mainly in Chilean sandy beaches) in the promenade area, located very 199 close to the beach. Such light intensities could be higher than those occurring in the intertidal area, 200 mainly in the middle and lower intertidal zone. Therefore, future studies should consider using 201 202 the light intensity directly recorded at the intertidal zone. However, at least for the Chilean coast, 203 in extreme heavy polluted beaches, light intensity values in the intertidal zone can be as high or higher than those used in these studies, mainly in the upper intertidal zone (Duarte unpublished 204 205 data). Another important consideration should be to expand ALAN studies to beaches located in 206 different geographic areas, for example tropical ones, which with the exception of sea turtle species, have not received special attention regarding to ALAN. 207

208 **2.1.2 Sea-Turtles**

209



- 210
- Figure 4. Sea turtles hatching in Heron Island, Australia (Credit; Levy, O).
- 212

It is well established that ALAN, even at low levels, is a threat influencing all seven species of marine turtle, primarily as hatchlings or nesting adult females (Witherington & Bjorndal, 1991a,b; Salmon, 2003). Artificial light pollution is known to influence (i) the on-beach orientation and nest site selection of adult female turtles, (ii) the on-beach orientation and sea finding behavior of hatchling turtles, and (iii) the at-sea dispersal of hatchling turtles. The degree to which species, or populations are exposed to artificial light pollution, and thus its influence as a threatening process, varies across the world with populations nesting at sites closer to areas of urban or industrial development being more exposed (e.g., Kamrowski et al. 2012; 2014; Colmon
et al., 2020; Shimada et al., 2021).

Artificial light primarily impacts marine turtles during nesting or hatchling life stages, and 222 consequently it is predominantly linked to early life stage mortality. Thus, it could, along with 223 224 other threats, cause gradual decline in the reproductive output of a nesting area. However, while 225 there are several studies examining beach specific exposure, or short-term (season) impacts, quantifying the long-term impacts of artificial light pollution-caused mortality are difficult 226 because marine turtles can take decades to reach maturity, and impacts of light pollution contribute 227 to other pressures over turtles' lifetimes to influence population viability. Hence, understanding 228 the degree to which nesting sites for each population of turtles are exposed to artificial light 229 pollution is important, so that site-specific interventions can be implemented. 230

Typically, female turtles lay their clutches above the high-water mark, and often in the 231 primary dune systems or high up on the beach. Hatchlings generally emerge from nests at night 232 233 and orient themselves from the nest site to the ocean, ideally as fast as possible (Wyneken & Salmon, 1992). After emerging they generally show a preference of moving towards horizons 234 which are low and bright, and moving away from horizons which are dark and elevated (e.g., 235 236 Lucas et al., 1992; Salmon et al., 1995; Limpus & Kamrowski, 2013; Pendoley & Kamrowski, 2015) and using these cues they can navigate across the beach to the water. Exposure to coastal 237 light pollution disrupts the natural orientation cues and leads to the disorientation and 238 239 misorientation of hatchlings because lights obscure the natural horizons (Witherington & Bjorndal, 1991a,b). 240

Hatchling sea finding ability is influenced by both the wavelength and intensity of artificial 241 242 light (Witherington & Bjorndal, 1991b; Cruz et al., 2018). Such ability is significantly compromised by exposure to shorter wavelength lights, even at lower intensity (Salmon, 2003; 243 Celano et al. 2018). Importantly, it is becoming clear that the thresholds of concern for both 244 245 wavelength and intensity of artificial light are likely to vary within and among species (Fritsches, 246 2012). The impacts of exposure can be influenced by the presence or absence of other natural (dune height and structure, vegetation) or unnatural cues (presence of buildings or artificial 247 structures) (Salmon, 2003; Kamrowski et al., 2015), highlighting the need for site-specific 248 research on orientation thresholds and light-reduction interventions. 249

Once at sea, hatchling turtles will swim actively for around 24 to 48 hours (Wyneken & 250 Salmon, 1992). This period, known as the swim frenzy, enables hatchlings to move quickly from 251 252 nearshore to offshore waters (Wyneken & Salmon, 1992). During the swim frenzy the hatchlings are using multiple cues to enable their directional swimming – these include swimming towards 253 254 the low, light, horizon, and swimming perpendicular to wave fronts (Salmon & Wyneken, 1987; 255 Lohmann et al., 2017; Wilson et al., 2021). According to Salmon & Wyneken (1987) light cues 256 are important for at sea dispersal, however, there is likely to be a distance offshore where the cue is either not available or not used. While this distance is currently unknown, there is a growing 257 empirical basis demonstrating that offshore dispersal for marine turtle hatchlings is compromised 258 259 by light pollution originating from land-based or marine structures, such as infrastructure like 260 jetties (Truscott et al., 2017; Cruz et al., 2018; Wilson et al., 2018). Continuing to advance knowledge on how the at sea dispersal is influenced by artificial lights from shore, or offshore 261 262 infrastructure is a key avenue for further research as human developments expand along the coasts.

For nesting turtles there has been substantial research on factors that influence nest site 263 selection (i.e., the placement of clutches on a beach). Factors including distance from vegetation 264 (most nests being laid closer to the vegetation line – Hays et al., 1995; Kelly et al., 2017), elevation 265 and beach slope (Wood & Bjorndal, 2000; Patrício et al., 2018) and exposure to artificial light 266 267 pollution (Salmon, 2003; Windle et al., 2018) have all been associated with nest site selection. 268 There is also variation among species and region as to the relative importance of each. However, 269 less research has been conducted to examine the influence of light pollution on nest site selection 270 by nesting turtles. Among the studies, Salmon (2003) used data from a main nesting area in Florida 271 to test nest site selection in relation to the degree females were exposed to artificial light spill onto beaches; and Windle et al. (2018) used a combination of light pollution data and turtle density 272 273 data to examine the influence of artificial light pollution on nest site selection. Both studies 274 concluded that turtles use darker beaches and select darker sections of beaches.

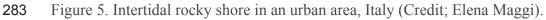
275 276

277 **2.2.** Rocky intertidal shores

278On a global scale, rocky intertidal shores are inhabited by diverse assemblages largely279influenced by light for metabolic activities and behaviors at different life stages. It is280reasonable to expect that the presence of ALAN may influence microphytobenthic and281macroalgal primary producers, as well as sessile and mobile animals.



282



Intertidal habitats represent a thin line demarcating the boundary between land and marinemasses. Among intertidal environments, rocky shores are characterized by large variations in

Global Change Biology

abiotic conditions, including strong gradients in wave exposure, temperature, and desiccation. 286 Despite the importance of such extreme factors, biotic interactions have shown strong 287 determinants of distribution patterns and abundance of organisms. These features have historically 288 289 made rocky shore intertidal habitats a natural laboratory to explore physiological and ecological 290 processes and mechanisms, which have been responsible for key conceptual advancements on 291 ecosystem functioning (Menge & Branch, 2001). ALAN, however, represents a guite novel and peculiar source of disturbance, whose effects are hardly predictable from knowledge on other 292 293 stressors. In fact, it is expected to act on pathways associated with natural circadian regulations, 294 which are related to different light optima and sensitivities among species and life stages (Davies & Smyth, 2018). Although to date the literature on effects of ALAN on intertidal rocky shores is 295 296 not particularly numerous, effects at the scale of both the individual and the ecosystem have already been highlighted. 297

298 Intertidal rocky shores host a diverse array of organisms, which can be primarily categorized 299 into sessile and vagile. Sessile species are those most diverse in terms of both size, taxonomy and trophic category. A large percentage of these species belongs to microbial biofilm. Despite low 300 visibility, its autotrophic component microphytobenthos, MPB) represents one of the main groups 301 302 of primary producers in intertidal habitats and, in association with heterotrophic microorganisms, 303 a source of food for a plethora of grazers species (Jenkins et al., 2001; Nagarkar et al., 2004). Seaweeds are the macroscopic primary producers on intertidal rocky shores, sometimes present 304 as dense macroalgal beds able to modify the abiotic conditions on the shore through their canopies. 305 Sessile species also include a variety of animals, such as bivalves, barnacles, ascidians, bryozoans, 306 hydrozoans, gastropods and polychaetes, among others. Mobile animals are abundant and 307 308 diversified as well; these include either herbivores, carnivores and omnivores almost freely living 309 on the different vertical portions of the intertidal habitats (e.g., limpets, gastropods, crabs, sea stars, sea urchins, small fish), or smaller species more strictly associated with larger ones, such 310 311 amphipods, isopods and crustaceans.

312 In the microtidal Mediterranean system, where intertidal biofilm is dominated by bacteria, 313 the presence of lit areas at night at intensities typically found along the coast (27 lux) was able to increase the biomass of MPB (here dominated by cyanobacteria) and its photosynthetic efficiency 314 (Maggi & Benedetti-Cecchi, 2018). Subsequent studies indicated that ALAN effect was likely 315 316 mediated by temporal changes in composition of mature assemblages (Maggi et al., 2020 a,b) and possibly related to different light optima among (groups of) species. Potential consequences of 317 effects on autotrophic microorganisms are not limited to net oxygen emissions and indirect 318 impacts on their consumers. In fact, the composition of microbial biofilm plays a key role in the 319 settlement of larval stages of invertebrates and spores (Keough & Raimondi, 1995; Oian et al., 320 321 2007). Indeed, the first study investigating the role of ALAN (either 30 or 19 lux) for intertidal 322 organisms focused on sessile larval stages of animals, revealing variable effects on settlement 323 rates on PVC panels attached on wooden floating rafts in the UK (Davies et al., 2015). Results showed that 39% of analyzed taxa were influenced by ALAN, either positively or negatively; it 324 325 was not surprising, given the importance of natural light as a cue for guiding larval recruitment 326 and later survival (Thorson, 1964; Mundy & Babcock, 1998). More recently, negative effects of ALAN on late settlers of barnacles have been documented both along natural shores in Chile (97-327 328 11 lux) (Notochthamalus scabrosus and Jehlius cirratus; Manriquez et al., 2021) and in the North 329 Atlantic on man-made structures (212-11 lux) (Semibalanus balanoides; Lynn et al. 2021a); in

these studies, lack of effects on early settlers suggested that, in presence of lit nights,
 metamorphosis was delayed or early mortality was increased in comparison to natural dark
 conditions.

333 As for sessile intertidal species, a big knowledge gap on effects on autotrophs still exists, with 334 a complete lack of studies on macroalgal species. In this case, it is worth mentioning that ALAN 335 effects could also influence non-trophic interactions mediated by algal canopies, such as facilitative effects exerted through the reduction of artificial light intensities for understory 336 337 assemblages (Bruno & Bertness, 2001). In addition to sessile species or life stages, intertidal rocky reefs are inhabited by mobile individuals. Mobility is a great advantage in a habitat characterized 338 by high abiotic variation. For example, many organisms have evolved predominantly nocturnal 339 behaviors to avoid energy expenditure related to thermal stress or to reduce the risk of predation 340 linked to visual stimuli (Wells, 1980; Manriquez et al., 2009). Different organs are involved in the 341 342 perception of circadian changes in light intensity and of prey or predators, from relatively simple 343 photoreceptors capable of forming sharp images in air in gastropods (Newell, 1965), to proper eyes in fish. Along the Chilean coasts, the abalone Concholepas concholepas (Bruguière, 1789), 344 or "Loco", is an ecologically and economically key species. Like most mollusks, it uses both 345 346 chemical and visual stimuli during sensory perception (Manríquez et al., 2014; Domenici et al., 347 2017). Chemoreception of odor cues through the osphradium is the main tool to monitor the presence of food items or predators; while information detected through its simple eyes modulates 348 phototaxis behavior and locomotion, and detection of forms. In addition, intertidal populations of 349 the "Loco" have been observed to prey mainly at night. This has led scientists to hypothesize 350 physiological and behavioral responses to the presence of ALAN. Indeed, field monitoring and 351 352 laboratory experiments have shown an increase in metabolic rate and self-righting time in juveniles in presence of LED illumination at night (~ 330 lux), as well as a preference for dark 353 areas to choose their prey (Manriquez et al., 2019). Similarly, ALAN (~ 100 lux) reduced feeding 354 355 activity in adult individuals, which were also more likely to be in a refuge than those under control 356 conditions (Manriquez et al., 2021). These impacts have clear implications for the long-term 357 sustainability and productivity of a keystone intertidal species that is able (among others) to consume the dominant mussel Perumytilus purpuratus, and therefore enhance rocky intertidal 358 biodiversity and functioning. These results do not, however, appear to be generalizable across 359 360 rocky shore predators. The common Atlantic dog whelk Nucella lapillus, forages more under ALAN even in the presence of a predator cue (Underwood et al. 2017) possibly due to increased 361 metabolic stress and the ability to visually perceive there is no predator threat. 362

363 Like some invertebrate species, many fish species have evolved endogenous clock systems which regulate tidally and circadian organized behavioral rhythms (Helfman et al., 2009). Among 364 them is the "Baunco", the rockfish Girella laevifrons, one of the most abundant fish in the littoral 365 366 zone of Southeastern Pacific. A recent study showed an increase in oxygen consumption and activity of this fish under ALAN conditions (70 lux). Importantly, loss of a dark night period was 367 able to modify or even stop the daily peak of activity of the Baunco, posing serious questions 368 369 about the sustainability of intertidal fish populations in urban areas (Pulgar et al., 2019). Results 370 are in accordance with those obtained on fish by manipulating ALAN under a wharf in Sydney Harbour (Bolton et al., 2017). The latter showed that, under ALAN, predatory behavior was 371 372 dramatically greater while abundance was reduced, a similar condition observed during daylight 373 conditions. Interestingly, authors observed a concomitant change in the structure of prev

Global Change Biology

374 (epifaunal) communities, which suggested an indirect top-down effect of ALAN. It is interesting to note that the presence of night lighting of artificial structures might elicit variable effects on 375 fish depending on the positioning of the light source. Lamps positioned over the structure, for 376 example, create an unnatural high contrast between illuminated areas and darker surroundings, 377 378 these latter found to attract higher densities of bogues (*Boops boops*) possibly seeking for a refuge 379 (Georgiadis et al. 2014; Mavraky et al. 2016). Further variability is likely related to speciesspecific responses, as suggested by the almost lack of effects on juvenile bonefish (Szekeres et al. 380 2017) or on most fish assemblage inhabiting shallow coastal seagrass beds (Martin et al. 2021). 381

The mentioned studies suggest that the effect of ALAN on single organisms can easily 382 reverberate on entire communities through trophic and non-trophic interactions. Further, 383 manipulative experiments in the Mediterranean rocky intertidal on biofilm assemblages and their 384 main consumers (i.e., littorinid gastropods) highlighted quite complex and temporally variable 385 effects. On the short term (~ 1 month), the increase in consumer pressure by grazers compensated 386 387 for the positive effect of ALAN (27 lux) on microbial primary producers (MPB), indicating that trophic interactions can provide a stabilizing mechanism against the effects of light pollution on 388 early colonizing autotrophs (Maggi & Benedetti-Cecchi, 2018). On the contrary, on longer terms 389 390 (~ 3 months) the presence of ALAN negatively affected the density of grazers. Adding to the 391 positive effect on cyanobacteria, ALAN eventually promoted a shift in the composition of epilithic assemblages, characterized by a higher photoautotrophic diversity at the expense of heterotrophic 392 393 bacteria (Maggi et al., 2020a). Although it's clear how pervasive the role of light pollution on 394 rocky shore intertidal communities is, biological and ecological consequences on different species and communities are still largely unknown. 395

396

2.3. Shallow water coral reef ecosystems

Reef building corals are highly photosensitive, and it is already clear that ALAN is a
 major emerging sensory pollutant concern for shallow coral reef ecosystems. Yet, ALAN is
 one of the most understudied threats to corals.







Figure 6. Coral reef 10-meter depth, Eilat, Red Sea (Credit; Shachaf Ben Ezra).

Tropical coral reefs are one of the most biodiverse and productive ecosystems on Earth. Their complex framework offers a unique habitat for thousands of associated species, supporting more species per unit area than any other marine ecosystem. In addition, coral reefs also provide important ecosystem services to millions of people (Hughes et al., 2017, 2018); yet, they have been heavily deteriorated worldwide due to poorly managed anthropogenic activities, habitat loss, and climate change (Hughes et al., 2018).

ALAN has been detected in fringing reefs localized in strongly urbanized locations. Mean night sky brightness levels at reef locations (see Ayalon et al., 2021a) show that many coral reef areas worldwide are affected by ALAN. An applicable example is the coast in the Gulf of Eilat/Aqaba in the Red Sea, where most of the studies on the effects of ALAN on reef building corals have been conducted (e.g., Tamir et al., 2017; Ayalon et al., 2019; Levy et al., 2020). This region is heavily light polluted, and the light reflected from the cities surrounding the reef can be seen from space (Tamir et al., 2017).

416 Scleractinian corals in symbiosis with dinoflagellates (Symbiodiniaceae) are the foundation 417 species for the formation of shallow water tropical coral reefs. Because their endosymbiotic 418 partners perform photosynthesis - and up to 95% of the photosynthates can be translocated to the coral host for its metabolic needs - this symbiosis is at the basis of the success of such tropical 419 environments (Muscatine et al., 1981; Tremblay et al., 2012; LaJeunesse et al., 2018). The aspects 420 of such symbiosis contribute to a higher susceptibility to ALAN, since corals and their symbionts 421 422 are highly photosensitive and are mostly found in shallow, clear water with relatively high light 423 levels (Rosenberg et al., 2019).

Light is detected by corals through light-sensing molecules, such as cryptochromes (CRY)proteins that can convert light leading to changes in intracellular levels of important second

Global Change Biology

426 messengers (Levy et al., 2007). The light/dark cycle regulates many cellular processes, the dark 427 period being crucial for stress recovery and repair, especially for the photosynthetic symbionts 428 (Hill et al., 2011; Levy et al., 2020). Natural periodic illumination (both solar and lunar) is a critical factor in cueing important processes of coral reproduction (Ayalon et al., 2021a; Lin et al., 429 430 2021). Also, the blue light spectrum – e.g., present in LED lights - play a key role in coral growth, 431 symbiont density, Chlorophyll a content, and photosynthetic rates (D'Angelo et al., 2008; Wijgerd et al., 2014). Therefore, it's not surprising that the exposure of corals to ALAN has been shown 432 433 to have detrimental effects on coral/symbionts metabolism and reproduction (see below). Yet, ALAN is one of the most understudied threats to corals. 434

Molecular evidence in corals on the effects of ALAN generally match those of more complex organisms, mainly mammalians. Rosenberg et al. (2019) used transcriptomic analysis to compare corals of the species *Acropora eurystoma* growing under natural light cycles and under ALAN (50-40 lux). Authors demonstrated many pathways that were altered, with approximately 25 times more differentially expressed genes that regulate cell cycle, cell proliferation, cell growth and protein synthesis under ALAN.

Physiological and biochemical investigations on Red Sea corals reported significant 441 442 deleterious effects of ALAN. Ayalon et al. (2019) first demonstrated that Acropora eurystoma 443 and *Pocillopora damicornis* experienced oxidative stress and photosynthetic impairment after exposure to different wavelengths of ALAN (40-35 lux). Subsequently, a more detailed study by 444 445 Levy et al. (2020) showed that the extent of deleterious effects of ALAN (40-30 lux) on the symbiotic association (loss of symbionts and Chlorophyll content) and physiology of Turbinaria 446 *reniformis* and *Stylophora pistillata* was aligned with the severity of the oxidative stress condition 447 448 experienced by the species. The same study also presented preliminary evidence that corals presenting higher basal levels of antioxidant capacity, such as *Turbinaria reniformis*, may be more 449 resistant to ALAN. Ayalon et al. (2021b) also reported ALAN can influence Symbiodinaceae 450 451 cultures and demonstrated different physiological responses according to the algae type. More 452 specifically, *Clodocopium* type showed to be generally more sensitive compared to *Durusdinium* type, presenting decreases in the maximum quantum efficiency of PSII, in the mitotic index, and 453 454 in total chlorophyll content after exposure to ALAN with illumination level up to 5 lux.

Regarding the effects on reproduction, recent studies reported large effects of ALAN on the 455 456 gametogenesis and the synchronization of gamete release in prominent coral species from the Indo-Pacific Ocean. The gametogenesis cycle of Acropora millepora and Acropora digitofera was 457 delayed or masked by exposure to ALAN (~ 15 lux), leading to unsynchronized gamete release 458 (Ayalon et al., 2021a). Dim light during the night also suppressed spawning in coral Dipsastrea 459 specisosa (Lin et al., 2021). Importantly, this later study showed that the period of darkness 460 461 between sunset and moonrise is essential to trigger synchronized mass spawning. Additionally, 462 Tamir et al. (2020) reported a 30% decrease in coral settlement success due to ALAN (~ 20 lux). Such results are alarming for the future of coral reefs. More than 80% of Scleractinian corals are 463 broadcasting spawners (Baird et al., 2009), and asynchronous reproduction caused by ecological 464 465 speciation could lead to reproductive isolation and prevent gene flow between differential lit coral communities (Rosenberg et al., 2019). Further, successful gamete production and fertilization, 466 development of viable offspring, and survival of coral recruits are undoubtedly some of the most 467 468 relevant processes for replenishing degraded reefs (Harii et al., 2010; Ayalon et al., 2021a).

In addition to the symbiosis established with endosymbiotic dinoflagellates, corals are 469 associated with prokaryotic symbionts. In fact, the coral host, and their microbiome (microalgal 470 471 and prokaryotic symbionts) show a tightly intertwined metabolic activity (Thompson et al., 2015). Coral-associated prokaryotic microbes are taxonomically and functionally diverse and are key for 472 473 maintaining the health of the holobiont (Krediet et al., 2013; Hernandez-Agreda et al., 2018; Olson 474 et al., 2009). To date, only one study explored the effects of ALAN on the coral microbiome. Baquiran et al. (2020) showed that the overall microbial community structure of the coral 475 476 Acropora digitifera remained stable under ALAN (~15 lux). However, it is important to note that bacteria that could use light for energy production (chlorophototrophic members of the phylum 477 Proteobacteria), as well as those that are associated with the phototrophic symbionts of the coral, 478 increased in abundance under light pollution conditions. Possibly, the higher abundance of 479 symbiont-associated microbes is linked to greater abundance and activity of the dinoflagellate 480 symbionts under short-term exposure to ALAN (Baguiran et al., 2020). 481

482 As for other species inhabiting coral reefs, only effects on fishes have been investigated so far. Fobert et al. (2019, 2020) showed a negative impact of ALAN (~ 15 lux) on the reproductive 483 success of the common clownfish Amphiprion ocellaris, with an increased interval between 484 485 spawning events and a smaller size of eggs in comparison to dark conditions; in addition, hatching 486 success was affected both by the presence of ALAN and by its spectral composition, with a more negative effect of cool-white in comparison to warm-white light. After hatching, other life stages 487 might represent critical intervals for the persistence of fish populations under light pollution. 488 O'Connor et al. (2019) highlighted larvae of Acanthurus triostegus living 10 days post-settlement 489 under ALAN conditions (~ 20-25 lux) experienced higher mortality rates by the end of the 490 491 experiment, although growing faster and heavier than control ones. Furthermore, a long-term study conducted in the wild (Schligler et al. 2021) showed that environmentally relevant intensity 492 of ALAN (~ 4.3 lux) is also able to reduce survival and growth of juveniles of the anemonefish 493 494 Amphiprion chrysopterus), compared to individuals exposed to natural night illuminance (by 495 moonlight). Finally, a recent study on the blue green *Chromis viridis* (Hyllyer et al. 2021) highlighted complex sub-acute effects of ALAN on adult fish, with predator threat able to alter 496 the increased metabolism of both specific tissues and whole organism observed under ALAN 497 conditions (~100 lux). Although an evolving area of study, it's already clear that ALAN is a major 498 499 emerging sensory pollutant of concern for shallow coral reef ecosystems. ALAN acts as a chronic disturbance, and corals under such pressure may not be able to perform their normal cyclic 500 501 behaviors (Rosemberg et al., 2019). Therefore, ALAN may impact the future of coral reefs, eventually contributing to global coral reef decline. 502

- 503
- 504 **2.4.** Pelagic environment organisms

505Oceans are vast, three-dimensional, and mostly influenced by currents. Pelagic506organisms are not attached to a substrate, hence the direct effects from ALAN in the open507oceans are likely to be different from those on a beach or on the seafloor. Still, recent work508suggests that lights from ships may have an impact on organisms in both the epipelagic and509even mesopelagic zones.



510

511 Figure 7. Lights from a ship working in the dark (Credit; Mike Snyder)

512 Light influences pelagic organisms in many ways, and artificial light may have a strong impact in their behavior (Blaxter & Currie, 1967). Prey availability, limiting the initiation and 513 magnitude of phytoplankton blooms and mortality through visual predation are some examples of 514 how artificial light may have an impact. In general, most zooplankton are negatively phototactic 515 (Forward, 1988), migrating to depth during daylight to avoid the threat of visual predation, and 516 517 surfacing at night to feed. This behavior is called Diel Vertical Migration (DVM) (Brierley, 2014). DVM is a characteristic feature of all the world's oceans and is considered the largest 518 synchronized movement of biomass on the planet (Hays, 2003). It is thus an important factor in 519 520 structuring pelagic communities. At the same time, as this process is mediated by light 521 (Ringelberg, 2010), it may potentially also be strongly affected by artificial light (Berge et al., 2020). At latitudes characterized by midnight sun during summer and polar night during winter, 522 DVM was generally assumed not to occur except during spring and autumn periods when there is 523 a clear day-night cycle (Berge et al., 2009). This view, however, was recently challenged. Instead 524 525 of an ecosystem that enters a resting state during the polar night, we now recognize a highly active ecosystem characterized by continuous activity and biological interactions across all trophic levels 526 527 and taxonomic groups (Berge et al., 2015). Importantly, even at the darkest periods of the year, light is still the primary regulative factor for most of these interactions, including vertically 528 529 migrating zooplankton (Last et al., 2016; Ludvigsen et al., 2018).

530 It is important to note that behavioral responses to artificial light vary among taxa. While 531 some species are known to be attracted to light, with herring *Clupea harengus* (Stickney, 1969), 532 krill (Utne-Palm et al., 2018, Krafft & Krag, 2021), snow crab Chionoecetes opilio (Nguyen et al., 2020), and grey mullet Mugil cephalus (Marchesan et al., 2005) as well-known examples, 533 534 others are known to avoid light. North Atlantic and Arctic copepods (Ludvigsen et al., 2018), 535 Atlantic cod Gadus morhua (Utne-Palm et al., 2018), and sea bream Sparus auratus (Marchesan et al., 2005), all species that are commercially important, have been shown to avoid artificial light 536 537 at night. A study from the Red Sea, in which a ROV equipped with LED lamps were used to herd mesopelagic scattering layers, similarly concluded that artificial light attracted some species and 538

repelled others (Kaartvedt et al., 2019). Also, recent studies from the high Arctic Archipelago of 539 Svalbard have shown that artificial light from both ships and instrumentation may have a strong 540 impact on organisms down to at least 200m depth (Berge et al. 2020). The artificial lights in this 541 case were measured to 2.2 µmol photons m⁻² s⁻¹ at the sea surface. However, with field 542 543 experiments carried out across nearly 8 degrees of latitude, differences in response to artificial 544 light varied both qualitatively and quantitatively in a way that could not be explained by species composition alone. Hence, in addition to interspecific differences in responses to light (Ryer et al. 545 546 2009), intraspecific variation could also complicate interpretations of responses to artificial light 547 (Berge et al 2020).

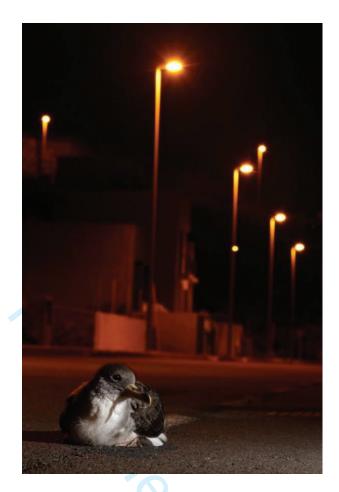
Effect of light pollution and ALAN in the open ocean is difficult to assess. By default, 548 sampling in the open ocean is often biased, as organisms are not attached or restricted to a 549 specific "site" or physical habitat. And except for acoustic instruments, most sampling 550 technologies include the use of either artificial light or the instrument itself creates a shadow 551 552 that might influence the organisms (see Box 1). To examine the potential effect of artificial light is thus very difficult to do with traditional methodologies that use artificial light to 553 554 function. A significant "effect" of ALAN in the open ocean is therefore not restricted to direct 555 effects but will also to a large degree include accuracies and artifacts in the measurements itself.

556

557

558 **2.5. Seabirds**

559Light pollution causes massive mortality events on seabird fledglings, involving more560than 70 species, some of them severely threatened. More subtle effects of ALAN are still561poorly understood.



562

Figure 8. Cory's shearwater *Calonectris borealis* fledgling grounded by artificial light in
Tenerife, Canary Islands (Credit; Beneharo Rodríguez).

Seabirds are defined as avian species for which a large proportion of the population relies on 565 the marine environment for at least part of the year (Croxall et al., 2012). For example, petrels 566 567 spend most of their life at sea and only visit land for breeding (Brooke, 2004), while some gull species can spend most of their lives outside the ocean. Within marine biota, seabirds are the only 568 animals mastering the three environments: marine, terrestrial, and aerial. Thus, there is a trade-off 569 among the adaptations of seabirds to cope with different environments. Seabirds are one of the 570 571 most threatened groups of birds (Dias et al., 2019). According to the International Union for 572 Conservation of Nature (IUCN) Red List criteria, around 31% of all seabird species are globally threatened and 47% have declining population trends. 573

574 Environmental conditions, such as light, can rapidly change. Many seabird species nest 575 underground or visit their breeding colonies at night while foraging during daylight. In addition, 576 many seabirds forage by diving in the water column, where the light spectrum changes rapidly with depth (Regular et al., 2011). Seabirds must adapt to rapid changes in intensity and spectra of 577 light to survive. In fact, visual systems of diving seabirds are more sensitive to blue light, the light 578 that penetrates more in-depth (Martin, 2017). The increase of artificial light levels has been 579 identified as a threat for seabirds and, particularly, petrels (Dias et al., 2019; Rodríguez et al., 580 581 2019).

Seabirds encounter the most light-polluted areas on land, mostly coastal areas close to or 582 within their nesting colonies. These light sources, such as streetlights, road lights and lighthouses, 583 584 attract and disorient birds during their flights between colony and foraging sites at sea. Many seabirds visit colonies at night presumably to avoid predation by diurnal predators (Bourgeois et 585 586 al., 2008; Rubolini et al., 2015). Visit frequency of breeders is influenced by moon cycles probably 587 because of moonlight level variation (Riou & Hamer, 2008; Rodríguez et al., 2016). Thus, breeders could be deterred from visiting colonies when lights are turned on close to their nests or 588 589 colonies. A recent experimental study on breeders of Manx shearwater Puffinus puffinus demonstrated that adult breeders flying over the colony were deterred by blue and green light in 590 comparison to red light (Syposz et al., 2021). Also, the number of birds flying over the colony 591 decreased with the duration and intensity of light treatments (Syposz et al., 2021). However, 592 experimental studies on the effect of color and intensity of light (3, 15 and 100 lux) on the colony 593 594 attendance of the smallest and only penguin whose activity on land is strictly nocturnal, the little 595 Eudyptula minor, demonstrated that penguins preferred lit paths over dark paths to reach their nests (Rodríguez et al., 2018). 596

From a conservation point of view and in a short-scale term, the most negative consequence 597 598 of light pollution is direct mortality caused mainly on fledglings of underground-nesting seabird 599 species (Rodríguez et al., 2017b). This phenomenon is known as fallout, and it occurs in all the oceans and seas across the world. Although reasons are unknown (Atchoi et al., 2020), fledglings 600 601 of many petrel species, but also Alcids and some sea ducks, are attracted and/or disorientated by 602 lights during their maiden flights from their nests toward the ocean (Rodríguez et al., 2017b). This leads to grounding and hitting infrastructures, e.g., wires, antennas, trees, buildings, or even the 603 604 ground, causing injuries and fatal victims. If they survive the first collisions, they are vulnerable to other threats, such as vehicle collisions, predation by domestic animals, or traps where they die 605 of inanition or dehydration, because they are usually unable to take off. Rescue programs are 606 607 initiatives aiming to mitigate light-induced mortality. To reach this, they call for the public 608 implication in rescuing and reporting birds grounded at lit areas, main towns, and cities. Most birds admitted to rescue programs survive (~ 90%), but the fraction of grounded birds that never 609 are found or reported through these collaborative initiatives is unknown. Some studies indicate 610 that around 40% of birds are never rescued (Ainley et al., 2001, Rodríguez et al., 2014). Artificial 611 612 light could also affect the natural colony attendance of breeders visiting colonies at night.

Increasing ALAN levels in and around nesting colonies could impact seabird's breeding behavior and, consequently, chick provisioning. In an overnight weight gain study, Scopoli's shearwater *Calonectris diomedea* chicks situated closer to a high-light intensity disturbance (i.e., disco event) gained less weight compared to conspecifics from nests further away (Cianchetti-Benedetti et al., 2018). Such effects were not perceivable at fledging, but it is expected that a higher frequency of disturbance events could affect chicks' fitness and, even, breeding success.

Beyond the coastline, seabirds can also encounter extremely light-polluted areas associated
to offshore oil and gas platforms, vessels, or light-enhanced fisheries, for example. Our knowledge
on light-induced mortality at sea is quite limited (Merkel & Johansen, 2011; Glass & Ryan, 2013;
Ronconi et al., 2015; Gjerdrum et al., 2021; Ryan et al., 2021), although we know that adults can
also be involved in attraction episodes. Clear monitoring protocols and independent trained
observers, who could rely on technological advances, such as telemetry, thermal cameras, acoustic

recordings, and radar, are essential to record episodic seabird-light interactions at sea (Ronconi et al., 2015).

The increase of light pollution levels under water (see above) widens the photic zone at night,
but also during dawn and dusk. Thus, both at neritic and oceanic waters the increase of light levels
by ALAN could enhance the foraging of pursuit-diving visual predator seabirds, such as murres
and penguins (Cannell & Cullen, 1998; Regular et al., 2011; Elliott & Gaston, 2015).

Artificial lights can also concentrate prey, which seabirds then take advantage of. Several gull 631 632 species have been reported to increase their foraging opportunities on marine, coastal and terrestrial lit areas. For example, fishing vessels usually use light to concentrate fish and squid. In 633 the Mediterranean Sea, lights of fishing vessels favor the capture of fish by the Audouin's gull 634 Ichthyaetus audouinii by illuminating the sea surface, concentrating fish, and locating shoals 635 (Arcos & Oro, 2002). Similarly, Brown-hooded gulls Larus maculipennis predate on Isopoda, 636 Polychaeta, fish larvae, and crustaceans, which are concentrated under artificial lights on 637 638 Argentinian coastal piers (Leopold et al., 2010). On land, Black-backed gull Larus dominicanus can take advantage of Cerambicidae beetles attracted to artificial lights at sawmills (Pugh & 639 Pawson, 2016). Gulls can also prey on seabirds and such predation can be facilitated by artificial 640 641 light. At Benidorm Island (western Mediterranean), Yellow-legged gulls Larus michahellis 642 increased predation on European storm-petrels Hydrobates pelagicus after light levels increased 643 by a new light installation in the nearby Benidorm city (Oro et al., 2005).

- 644 Although there is a certain consensus about the higher pervasiveness of blue light for wildlife and, particularly, seabirds (Rodríguez et al., 2017a; Longcore et al., 2018; Syposz et al., 2021), 645 more research is needed on the spectrum of light in the perception of seabirds. Similarly, 646 647 reductions in the duration of lights, by means of smart-lighting or part-night lighting, could help to mitigate light pollution impacts, but current understanding is insufficient to underpin sound 648 recommendations for all species. For example, more research is needed to assess the threshold 649 650 levels from which a response is triggered as well as the relative intensity with ambient light levels 651 (e.g., during full and new moon nights). The distance from light sources and seabirds at which they are attracted or disorientated must be influenced by light intensity. Thus, determining 652 distances at which individuals are safe is crucial for managing breeding colonies and corridors 653 between colonies and the ocean for inland breeding species. GPS tracking has revealed that most 654 655 of Cory's shearwater Calonectris borealis fledglings are grounded on areas distant less than 16 656 km from their nests (Rodríguez et al., 2015; 2022).
- 657

658 **3.** Conservation guidelines and strategies

Some few countries today - Spain (Catalonia), Chile, France, and Italy (Piedmont) - are trying 659 660 to establish laws to regulate light pollution. Most of the applicable documents addressing ALAN 661 are guidelines, Codes or Standards issued by regulators, advisory committees, non-government organizations (NGO) or professional bodies with no legal basis for enforcement of 662 recommendations. Many of the professional body guidance documents are targeted at lighting 663 664 engineers or designers, provide little detail regarding ALAN management and mitigation for the protection of sensitive receptors and must be purchased at a substantial cost (e.g., AS/NZS 665 666 4282:2019). Also, regulators, advisory committees, and NGOs typically focus on a single

sensitive receptor such as bats, birds or dark sky conservation for astronomy or star gazing (Voigt
et al., 2018; City of Calgary, 2011; NSW 2016).

Following the adoption of the Australian National Light Pollution Guidelines for Wildlife, 669 including Marine Turtles, Seabirds and Migratory Shorebirds (Commonwealth, 2020; CMS, 2020 670 a,b) by the UNEP Convention of Migratory Species in 2020, the CMS Secretariat has expanded 671 672 on the issue with the release of a follow-up review of ALAN impacts on migratory species not covered by the Australian guidelines (CMS, 2021). This document summarized some of the 673 674 available international laws and guidelines that address ALAN. Except for the CMS guidelines (CMS, 2020b), laws, standards and codes all relate to terrestrial based ALAN and sensitive 675 676 receptors. Guidance for addressing the impacts and management of artificial light in the marine environment does not currently exist in the (English) grey literature or publications. Recognizing 677 678 the difficulties in setting specific assessment trigger values for the broad range of variables influencing the impact of light on wildlife, the CMS guidelines recommend a case-by-case 679 680 approach to ALAN impact assessment, management, and mitigation. The conservation strategies 681 adopted will vary depending on the sensitive receptor (foraging and migrating birds, hatchling sea turtles migrating offshore, plankton, fish, marine mammals etc.), the sensitivity of the receptor to 682 683 different wavelengths of light (e.g., whales do not see color), the light sources (ships at anchor 684 offshore, offshore oil and gas facilities and flaring, ports and marinas, slow moving dredge vessels etc.), the light features (wavelength/color, intensity, shielding, flaring gas flow rates, elevation) 685 686 and variables such as turbidity, water depth, clouds, dust, aerosols, moon phase, fog, day length, all acting in combination to influence the visibility of the light. Standard best practice guidelines 687 for outdoor lighting design for the protection of sensitive receptors including wildlife have been 688 689 published (CMS, 2020b; IDA, 2021; ADSA, 2021) and include: (i) Start with natural darkness and only add light for specific purposes; (ii) use adaptive light controls to manage light timing, 690 691 intensity, and color; (iii) light only the object or area intended, (iv) use the lowest intensity lighting 692 appropriate for the task; (v) use non-reflective, dark-colored surfaces; and (vi) use lights with 693 reduced or filtered blue, violet, and ultraviolet wavelengths. Of these, the most important is avoiding short wavelength blue light due to its ubiquitous visibility across a wide range of taxa, 694 695 as well as its higher capacity to penetrate the water column, shielding light to prevent light spill 696 into the water or sky, and minimizing light intensity.

697 A correct application of such rules, for the management of 'dark nights' in the marine environment, needs the consultation of appropriately qualified biologists, which should be 698 included in the lighting design process as well as light management guidance documents and 699 regulations. At a larger scale, qualified researchers should help identify appropriate 'dark 700 701 sanctuary areas' within MPAs and providing for specific regulations in different marine habitats. 702 including long-term monitoring programs. Considering the best practices guidelines and their 703 correct application, we propose here "Ten golden rules for dark night conservation for marine 704 habitats" (See Fig. 9).

10 GOLDEN RULES FOR USING ARTIFICIAL LIGHT AT NIGHT

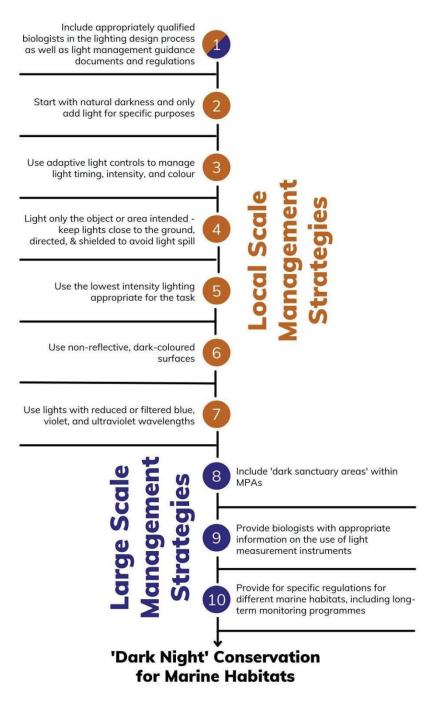




Figure 9. Ten golden rules for dark night conservation for marine habitats.

709 4. Conclusion and perspectives

Progress in our understanding of ALAN impacts in marine ecosystems has accelerated dramatically over the last five years. The number of species, habitats and ecological processes with documented responses now present a compelling case for ALAN as a globally widespread pollutant that is reshaping nature across our coastlines. The field remains however, recently emergent, and numerous knowledge gaps exist that if addressed would aid in the prediction and mitigation of ALAN impacts in the sea. Here we highlight key questions for future research to be addressed:

717 1. What is the impact of artificial light at sea on marine wildlife populations?

2. What is the contribution of marine traffic (i.e., mobile light sources from vessels) and traffic
management (i.e., fixed light sources from navigation markers) to marine light pollution, and what
is their potential to impact marine biodiversity?

3. What are the indirect impacts of ALAN in marine ecosystems through species interactionsand trophic cascades?

723 *4. Are there intergenerational impacts of ALAN?*

5. What are the best practice techniques to monitor and measure biologically meaningful
(i.e., radiometric as opposed to photometric) light, both underwater and on land, at both fine
(meters) and broad (kilometers) scales?

6. What are the thresholds (intensity, wavelength, exposure time) that elicit biological
responses in marine species?

729 7. How does the disruption of moonlight cycles by artificial skyglow impact circalunar730 rhythms in marine organisms?

731 8. What is the impact of coastal ALAN on marine ecosystem services?

732 9. Does ALAN impact long distance mass migrations of marine megafauna?

10. Is there temporal variability (e.g., monthly, or seasonal) in ALAN impacts on marineecosystems?

11. Are there additive/interactive effects between ALAN and other anthropogenic disturbances?

Addressing these questions will provide insight into the full extent of ALAN impacts inmarine ecosystems, their consequences for society, and options for mitigating them.

- 739
- 740

741

743 **References**

744

771

772

745	ADSA (2021). https://www.australasiandarkskyalliance.org/best-practice-
746	lighting AS/NZS 4282:2019 (2019) Control of the obtrusive effects of outdoor
747	lighting. Standards Australia https://infostore.saiglobal.com/en-us/standards/as-nzs-
748	<u>4282-2019-1141358_saig_as_as_2703687/</u>

- Ainley, D.G., Podolsky, R., Deforest, L., Spencer, G. (2001). The status and
 population trends of the Newell's Shearwater on Kaua'i: Insights from modeling. *Stud. Avian. Biol.*, 22, 108–123.
- Arcos, J.M., Oro, D. (2002). Significance of nocturnal purse seine fisheries for
 seabirds: a case study off the Ebro Delta (NW Mediterranean). *Mar. Biol.*, 141, 277–
 286.
- Atchoi, E., Mitkus, M., Rodríguez, A. (2020). Is seabird light-induced mortality
 explained by the visual system development? *Conserv. Sci. Pract.*, 2, e195.
 doi:10.1111/csp2.195.
- Aulsebrook, A.E., Jechow, A., Krop-Benesch, A., Christopher, C.M., Longcore,
 T., Perkin, E.K., van Grunsven, R.H.A. (2022). Nocturnal lighting in animal research
 should be replicable and reflect relevant ecological conditions. *Biol. Lett.*, *18*,
 20220035.
- Ayalon, I., Benichou, J.I.C., Avisar, D., Levy, O. (2021b). The endosymbiotic
 coral algae Symbiodinaceae are sensitive to a sensory pollutant: Artificial Light at
 Night, ALAN. *Front. Physiol.*, *12*, 695083.
- Ayalon, I., Marangoni, L.F.B, Benichou, J.I.C., Avisar, D., Levy, O. (2019). Red
 Sea corals under artificial light pollution at night (ALAN) undergo oxidative stress
 and photosynthetic impairment. *Glob. Change Biol.*, 25, 4194–4207.
- Ayalon, I., Rosenberg, Y., Benichou, J. I., Campos, C. L. D., Sayco, S. L. G.,
 Nada, M. A. L., et al. (2021a). Coral gametogenesis collapses under artificial light
 pollution. *Curr. Biol.*, *31*, 413–419.
 - Baird, A.H., Guest, J.R., Willis, B.L. (2009) Systematic and biogeographical patterns in the reproductive biology of scleractinian corals. *Annu. Rev. Ecol. Evol. Syst.*, *40*, 551–571.
- Baquiran, J. I. P., Nada, M. A. L., Campos, C. L. D., Sayco, S. L. G., Cabaitan,
 P. C., Rosenberg, Y., et al. (2020). The Prokaryotic Microbiome of Acropora digitifera
 is stable under short-term artificial light pollution. *Microorganisms*, *8*,1566.

777 778	Barentine, J.C. (2019). Methods for assessment and monitoring of light pollution around ecologically sensitive sites. <i>Journal of Imaging</i> , 5, e5050054.
110	around ecologically sensitive sites. <i>Journal of Imaging, 5</i> , e5050054.
779	Bascom, W. (1980). Waves and beaches: The dynamics of the ocean surface. New
780	York: Doubleday & Co.
781	Basedow, S. L., Tande, K. S., Norrbin, M. F., Kristiansen, S. A. (2013). Capturing
782	quantitative zooplankton information in the sea: Performance test of laser optical
783	plankton counter and video plankton recorder in a Calanus finmarchicus dominated
784	summer situation. Prog. Oceanogr., 108, 72–80.
785	Båtnes, A. S., Miljeteig, C., Berge, J., Greenacre, M., Johnsen, G. (2013).
786	Quantifying the light sensitivity of Calanus spp. during the polar night: potential for
787	orchestrated migrations conducted by ambient light from the sun, moon, or aurora
788	borealis? <i>Polar Biol.</i> , 38, 51-65.
789	Benoit-Bird, K. J., Moline, M. A., Schofield, O. M., Robbins, I. C., Waluk, C. M.
790	(2010). Zooplankton avoidance of a profiled open-path fluorometer. J. Plank. Res.,
791	32, 1413-1419.
792	Berge, J. et al. (2009) Diel vertical migration of Arctic zooplankton during the
793	polar night. <i>Biol. Lett.</i> , 5, 69–72.
794	Berge, J., Daase, M., Renaud, P.E. et al. (2015). Unexpected levels of biological
795	activity during the polar night offer new perspectives on warming. Arctic. Curr. Biol.,
796	25, 2555–2561.
797	Berge, J., Geoffroy, M., Daase, M., Cottier, F., Priou, P., Cohen, J. H., Johnsen,
798	G., et al. (2020). Artificial light during the polar night disrupts Arctic fish and
799	zooplankton behaviour down to 200 m depth. Commun. Biol., 3, 102.
800	Bicknell, A. W. J., Godley, B. J., Sheehan, E. V., Votier, S. C., Witt, M. J. (2016).
801	Camera technology for monitoring marine biodiversity and human impact. Front.
802	<i>Ecol. Environ., 14</i> , 424–432.
803	Blaxter, J.H.S., Currie, R.I. (1967). The effect of artificial lights on acoustic
804	scattering layers in the ocean. In: Symposia of the Zoological Society of London, 19,
805	1-14.
806	Boldt, J. L., Williams, K., Rooper, C. N., Towler, R. H., Gauthier, S. (2018).
807	Development of stereo camera methodologies to improve pelagic fish biomass
808	estimates and inform ecosystem management in marine waters. Fish. Res., 198, 66-
809	77.
810	Bolton, D., Mayer-Pinto, M., Clark, G. F., Dafforn, K. A., Brassil, W. A.,
811	Becker, A., Johnston, E. L. (2017). Coastal urban lighting has ecological

Global Change Biology

812 813	consequences for multiple trophic levels under the sea. <i>Sci. Total Environ.</i> , <i>576</i> , 1-9.
814 815 816	Bourgeois, K., Dromzée, S., Vidal, É., Legrand, J. (2008). Yelkouan shearwater <i>Puffinus yelkouan</i> presence and behaviour at colonies: not only a moonlight question. <i>C. R. Biol.</i> , <i>331</i> , 88–97.
817 818	Bregazzi, P.K., Naylor, E. (1972). The locomotor activity rhythm of <i>Talitrus</i> saltutor (Montagu) (Crustacea, Amphipoda). J. Exp. Biol. 57, 375-391
819	Brierley, A.S. (2014). Diel vertical migration. Curr. Biol., 24, R1074-R1076.
820 821	Brooke, M. (2004). <i>Albatrosses and Petrels across the World</i> . Oxford, Oxford University Press.
822 823 824	Bruno, J.F., Bertness, M.D. (2001). Habitat modification and facilitation in benthic marine communities. In, <i>Marine Community Ecology</i> (M. D. Bertness, S. D. Gaines, M. E. Hay, eds.), pp. 221–251. Sunderland, MA: Sinauer Associates.
825 826 827	Buckley, L., Caldarone, E., & Ong, T.L. (1999). RNA-DNA ratio and other nucleic acid-based indicators for growth and condition of marine fishes. <i>Hydrobiologia</i> , 401, 265-277.
828 829	Cannell, B.L., Cullen, J.M. (1998). The foraging behavior of Little Penguins Eudyptula minor at different light levels. <i>Ibis</i> , 140, 467–471.
830 831 832	Celano, L., Sullivan, C., Field, A., Salmon, M. (2018). Seafinding revisited: how hatchling marine turtles respond to natural lighting at a nesting beach. <i>J. Comp. Physiol. A</i> , 204, 1007-1015.
833 834	Chícharo, M.A., Chícharo, L. (2008). RNA:DNA ratio and other nucleic acid derived indices in marine ecology. <i>Int. J. Mol. Sci.</i> , <i>9</i> , 1453-1471.
835 836 837	Cianchetti-Benedetti, M., Becciu, P., Massa, B., Dell'Omo, G. (2018). Conflicts between touristic recreational activities and breeding shearwaters: short-term effect of artificial light and sound on chick weight. <i>Eur. J. Wildl. Res.</i> , <i>64</i> , 19.
838 839	CIE. ISO23539:2005(E)/ CIE S 010/E:2004. Photometry – The CIE System of Physical Photometry. Standard; CIE; Vienna, Austria; 2005.
840 841 842	City of Calgary (2011). Bird-Friendly Urban Design Guidelines. Land Use Planning & Policy. https://www.calgary.ca/pda/pd/current-studies-and-ongoing-activities/urban-design.html
843 844	CMS (2020a). Resolution 13.5 Light Pollution Guidelines for Wildlife. Adopted by the Conference of the Parties at its 13th Meeting (Gandhinagar, February 2020).

https://www.cms.int/sites/default/files/document/cms cop13 res.13.5 light-845 pollutionguidelines e.pdf 846 CMS (2020b). Annex to Resolution 13.5 Light Pollution Guidelines. National 847 848 Light Pollution Guidelines for Wildlife including marine turtles, seabirds and 849 migratory shorebirds. https://www.cms.int/sites/default/files/document/cms_cop13_res.13.5_annex_e.pdf 850 851 CMS (2021). Convention on the Conservation of Migratory Species of Wild 852 Animals 5th Meeting of the Sessional Committee of the CMS Scientific Council (ScC-SC5) Online, 28 June – 9 July 2021 UNEP/CMS/ScC-SC5/Inf.7 IMPACT OF LIGHT 853 POLLUTION ON DIFFERENT TAXA OF MIGRATORY SPECIES (Prepared by 854 behalf of the CMS 855 Ms. Laetitia Nunny on Secretariat) 87 pp 856 https://www.cms.int/sites/default/files/document/cms scc-sc5 inf.7 impact-of-lightpollution-on-migratory-species e.pdf 857 858 Colman, L. P., Lara, P. H., Bennie, J. et al. (2020). Assessing coastal artificial light and potential exposure of wildlife at a national scale: the case of marine turtles 859 in Brazil. Biodivers. Conserv. 29, 1135-1152. 860 Commonwealth of Australia (2020). National Light Pollution guidelines for 861 Wildlife, including marine turtles, seabirds and migratory shorebirds. 111pp. 862 863 https://www.environment.gov.au/biodiversity/publications/national-light-pollutionguidelines-wildlife 864 Croxall, J.P., Butchart, S.H.M., Lascelles, B., Stattersfield, A.J., Sullivan, B., et 865 866 al. (2012). Seabird conservation status, threats and priority actions: a global assessment. Bird Conserv. Int., 22, 1-34. 867 Cruz, L. M., Shillinger, G. L., Robinson, N. J., Tomillo, P. S., Paladino, F. V. 868 (2018). Effect of light intensity and wavelength on the in-water orientation of olive 869 870 ridley turtle hatchlings. J. Exp. Mar. Biol. Ecol., 505, 52-56. D'Angelo, C. et al. (2008). Blue light regulation of host pigment in reef-871 building corals. Mar. Ecol. Prog. Ser., 364, 97-106. 872 873 Davies, T. W., Smyth, T. (2018). Why artificial light at night should be a focus for global change research in the 21st century. Glob. Change Biol., 24, 872-882. 874 Davies, T. W., Coleman, M., Griffith, K. M., Jenkins, S. R. (2015). Night-time 875 lighting alters the composition of marine epifaunal communities. Biol. Lett., 11, 876 20150080. 877 Davies, T. W., Duffy, J. P., Bennie, J., Gaston, K. J. (2014). The nature, extent, 878 and ecological implications of marine light pollution. Front. Ecol. Environ. 12, 347-879 355. 880

881 882 883	Davies, T. W., Duffy, J. P., Bennie, J., Gaston, K. J. (2016). Stemming the Tide of Light Pollution Encroaching into Marine Protected Areas. <i>Conserv. Lett.</i> , <i>9</i> , 164-171.
884 885	Davies, T. W., McKee, D., Fishwick, J., Tidau, S., Smyth, T. (2020). Biologically important artificial light at night on the seafloor. <i>Sci. Rep.</i> , <i>10</i> , 12545.
886 887 888	Davies, T.W., Bennie, J., Cruse, D., Blumgart, D., Inger, R., Gaston, K.J. (2017). Multiple night-time light-emitting diode lighting strategies impact grassland invertebrate assemblages. <i>Glob. Change Biol.</i> , <i>23</i> , 2641-2648.
889 890	Dias, M.P., Martin, R., Pearmain, E.J., Burfield, I.J., Small, C., et al. (2019). Threats to seabirds: A global assessment. <i>Biol. Conserv.</i> , <i>237</i> , 525–537.
891 892 893 894	Dimitriadis, C., Fournari e Konstantinidou, I., Sourbes, L., Koutsoubas, D., Mazaris, A.D. (2018). Reduction of sea turtle population recruitment caused by nightlight: evidence from the Mediterranean region. <i>Ocean Coast Manag.</i> , <i>153</i> , 108-115.
895 896 897	Domenici, P., Torres, R., Manríquez, P.H. (2017). Effects of elevated carbon dioxide and increased temperature on locomotion and the repeatability of lateralization of a keystone marine mollusk. <i>J. Exp. Biol.</i> , <i>220</i> , 667–676.
898 899 900	Doya, C. et al. (2014). Diel behavioral rhythms in sablefish (<i>Anoplopoma fimbria</i>) and other benthic species, as recorded by the Deep-sea cabled observatories in Barkley canyon (NEPTUNE-Canada). <i>J. Mar. Syst.</i> , <i>130</i> , 69–78.
901 902 903 904	Duarte, C., Quintanilla-Ahumada, D., Anguita, C., Manríquez, P.H., Widdicombe, S., Pulgar, J., Silva-Rodríguez, E.A., Miranda, C., Manríquez, K., Quijón, P.A. (2019). Artificial light pollution at night (ALAN) disrupts the distribution and circadian rhythm of a sandy beach isopod. <i>Environ. Pollut., 248</i> , 565-573.
905 906 907	Emily K. Fobert, E.K., Schubert, K.P., Burke da Silva, K. (2021). The influence of spectral composition of artificial light at night on clownfish reproductive success, J. Exp. Mar. Biol. Ecol., 540, 151559.
908 909	Elliott, K.H., Gaston, A.J. (2015). Diel vertical migration of prey and light availability constrain foraging in an Arctic seabird. <i>Mar Biol.</i> , 162, 1739–1748.
910 911	Falchi, F., Cinzano, P., Duriscoe, D., Kyba, C. <i>et al.</i> (2016). The new world atlas of artificial night sky brightness. Sci. Adv., 2, 1-26.
912 913 914 915	Fallaci, M., Aloia, A., Audoglio, M., Colombini, I., Scapini, F., Chelazzi, L. (1999). Differences in behavioral strategies between two sympatric talitrids (Amphipoda) inhabiting an exposed sandy beach of the French Atlantic Coast. <i>Estuar</i> . <i>Coast Shelf Sci.</i> , <i>48</i> , 469-482.

916 917 918	Fallaci, M., Aloia, A., Colombini, I., Chelazzi, L. (2002). Population dynamics and life history of two Phaleria species (Coleoptera, Tenebrionidae) living on the Tyrrhenian sandy coast of central Italy. <i>Acta Oecol.</i> , <i>23</i> , 69-79.
919 920 921	Fanini, L., Hughes, L.E., Springthorpe, R., Tosetto, L., Lowry, J.K. (2016). Surface activity patterns of macrofauna on pocket, tidal beaches: insights into the role of wrack and artificial lighting. <i>Reg. Stud. Mar. Sci.</i> , <i>7</i> , 63-71.
922 923	Fobert, E.K., Burke da Silva, K., Swearer, S.E. (2019). Artificial light at night causes reproductive failure in clownfish. <i>Biol. Lett.</i> 15, 20190272.
924 925	Forward Jr, R.B.J. (1988). Diel vertical migration: zooplankton photobiology and behaviour. <i>Oceanogr. Mar. Biol. Annu. Rev.</i> , <i>26</i> , 361–393.
926 927	Fritsches, K. A. (2012). Australian loggerhead sea turtle hatchlings do not avoid yellow. <i>Mar. Fresh. Behav. Physiol.</i> 45, 79-89.
928 929 930	Gaston, K. J., Ackermann, S., Bennie, J., Cox, D. T., Phillips, B. B., de Miguel, A. S., Sanders, D. (2021). Pervasiveness of biological impacts of artificial light at night. <i>Integr. Comp. Biol.</i> , <i>61</i> , 1098-1110.
931 932	Gaston, K.J., Bennie, J., Davies, T.W., Hopkins, J. (2013). The ecological impacts of nighttime light pollution: a mechanistic appraisal. <i>Biol. Rev.</i> , <i>88</i> , 912-927.
933 934 935	Geoffroy, M., Langbehn, T., Priou, P., Varpe, Ø., Johnsen, G., Le Bris, A., Fisher, J. a. D., Daase, M. et al. (2021). Pelagic organisms avoid white, blue, and red artificial light from scientific instruments. <i>Sci. Rep.</i> , <i>11</i> , 14941.
936 937 938 939	Georgiadis, M., Mavraki, N., Koutsikopoulos, C. et al. (2014). Spatio-temporal dynamics and management implications of the nightly appearance of Boops boops (Acanthopterygii, Perciformes) juvenile shoals in the anthropogenically modified Mediterranean littoral zone. <i>Hydrobiologia</i> , 734, 81–96.
940 941 942	Gjerdrum, C., Ronconi, R., Turner, K., Hamer, T. (2021). Bird strandings and bright lights at coastal and offshore industrial sites in Atlantic Canada. <i>Avian Conserv. Ecol. Publ.</i> , <i>16</i> , 22.
943 944	Glass, J., Ryan, P. (2013). Reduced seabird night strikes and mortality in the Tristan rock lobster fishery. <i>African J. Mar. Sci., 35</i> , 589–592.
945 946 947	Gonzalez, S.A., Yañez-navea, K., Muñoz, M. (2014). Effect of coastal urbanization on sandy beach coleoptera <i>Phaleria maculata</i> (Kulzer, 1959) in northern Chile. <i>Mar. Pollut. Bull., 83,</i> 265-274.
948 949	Grubisic, M., van Grunsven, R.H.A., Manfrin, A., Monaghan, M.T., Hölker, F. (2018). A transition to white LED increases ecological impacts of nocturnal

950 951	illumination on aquatic primary producers in a lowland agricultural drainage ditch. <i>Env. Poll.</i> , <i>240</i> , 630-638.
952	Hänel, A., Posch, T., Ribas, S.J., Aubé, M., Duriscoe, D., Jechow, A., Kollath, Z.,
953	Lolkema, D.E., Moore, C., Schmidt, N., Spoelstra, H. (2018). Measuring night sky
954	brightness: methods and challenges. Journal of Quantitative Spectroscopy and
955	Radiative Transfer, 205, 278-90.
956	Harii, S., Yamamoto, M., Hoegh-Guldberg, O. (2010). The relative
957	contribution of dinoflagellate photosynthesis and stored lipids to the survivorship
958	of symbiotic larvae of the reef-building corals. Mar. Biol., 157, 1215–1224.
959	Hayes, G. C. (2003). A review of the adaptive significance and ecosystem
960	consequences of zooplankton diel vertical migrations. <i>Hydrobiologia</i> , 503, 163-170.
961	Hays, G. C., Mackay, A., Adams, C. R., Mortimer, J. A., Speakman, J. R.,
962	Boerema, M. (1995). Nest site selection by sea turtles. J. Mar. Biol. Assoc. United
963	Kingdom., 75, 667-674.
964	Helfman, G., Collette, B., Facey, D., Bowen, B. (2009). The Diversity of Fish:
965	Biology, Evolution and Ecology. Willey-Blackwell. A John Wiley & Sons, Ltd.,
966	Publication.
967	Herman, A. W., Harvey, M. (2006). Application of normalized biomass size
968	spectra to laser optical plankton counter net intercomparisons of zooplankton
969	distributions. J. Geophys. Res. Oceans, 11, C05S05.
970	Hernandez-Agreda, A., Leggat, W., Bongaerts, P., Herrera, C., Ainsworth,
971	T.D. (2018). Rethinking the coral microbiome: Simplicity exists within a diverse
972	microbial biosphere. <i>mBio</i> , 9, e00812-18.
973	Hill, R., Brown, C.M., DeZeeuw, K., Campbell, D.A., Ralph, P.J. (2011).
974	Increased rate of D1 repair in coral symbionts during bleaching is insufficient to
975	counter accelerated photo-inactivation. Limnol. Oceanogr., 56, 139e146
976	Hölker, F., Wurzbacher, C., Weißenborn, C., Monaghan, M. T., Holzhauer, S. I.
977	J., Premke, K. (2015). Microbial diversity and community respiration in freshwater
978	sediments influenced by artificial light at night. Phil. Transac. Royal Soc. B., 370,
979	20140130.
980	Hughes, T. P., Barnes, M. L., Bellwood, D. R., Cinner, J. E., Cumming, G. S.,
981	Jackson, J. B., et al. (2017). Coral reefs in the Anthropocene. Nature, 546, 82-90.
982	Hughes, T.P., Anderson, K.D., Connolly, S.R., et al. (2018). Spatial and temporal
983	patterns of mass bleaching of corals in the Anthropocene. Science, 359, 80-83.
984	IDA (2021) https://www.darksky.org/our-work/lighting/lighting-principles/

985	Jaramillo, E., Contreras, H., Duarte, C., Avellanal, M.H. (2003). Locomotor
986	activity and zonation of upper shore arthropods in a sandy beach of north central Chile.
987	Estuar. Coast Shelf Sci., 58, 177-197.
988	Jaramillo, E., Dugan, J. E., Hubbard, D. M., Manzano, M., Duarte, C. (2021).
989	Ranking the ecological effects of coastal armoring on mobile invertebrates across
990	intertidal zones on sandy beaches. Sci. Total Environ. 755, 142573.
991	Jechow, A., Hölker, F. (2019). How dark is a river? Artificial light at night in
992	aquatic systems and the need for comprehensive night-time light measurements. Wiley
993	Interdisciplinary Reviews: Water, 6, e1388.
994	Jechow, A., Kyba, C., Hölker, F. (2019). Beyond all-sky: assessing ecological
995	light pollution using multi-spectral full-sphere fisheye lens imaging. Journal of
996	Imaging, 5, 46.
997	Jenkins, S.R., Hartnoll, R.G. (2001). Food supply, grazing activity and growth
998	rate in the limpet Patella vulgata L.: a comparison between exposed and sheltered
999	shores. J. Exp. Mar. Biol. Ecol., 258, 123-139.
1000	Kaartvedt, S., Røstad, A., Opdal, A. F., Aksnes, D. L. (2019). Herding
1001	mesopelagic fish by light. Mar. Ecol. Progr. Ser., 625, 225-231.
1002	Kamrowski, R. L., Limpus, C., Jones, R., Anderson, S., Hamann, M. (2014).
1003	Temporal changes in artificial light exposure of marine turtle nesting areas. Glob.
1004	Change Biol., 20, 2437-2449.
1005	Kamrowski, R. L., Limpus, C., Moloney, J., Hamann, M. (2012). Coastal light
1006	pollution and marine turtles: assessing the magnitude of the problem. <i>Endang. Species</i>
1007	<i>Res.</i> , <i>19</i> , 85-98.
1008	Kamrowski, R. L., Limpus, C., Pendoley, K., Hamann, M. (2015). Influence of
1009	industrial light pollution on the sea-finding behavior of flatback turtle hatchlings.
1010	Wildlife Res., 41, 421-434.
1011	Katie, E., Hillyer, D.J., Beale, J.S.S. (2021). Artificial light at night interacts with
1012	predatory threat to alter reef fish metabolite profiles. Sci. Total Environ., 769, 144482.
1013	Kelly, I., Leon, J. X., Gilby, B. L., Olds, A. D., Schlacher, T. A. (2017). Marine
1014	turtles are not fussy nesters: a novel test of small-scale nest site selection using
1015	structure from motion beach terrain information. PeerJ, 5, e2770.
1016	Kennedy, F., Naylor, E., Jaramillo, E. (2000). Ontogenetic differences in the
1017	circadian locomotor activity rhythm of the talitrid amphipod crustacean
1018	Orchestoidea tuberculata. Mar. Biol. 137, 511-517.

1019 1020	Keough, M.J., Raimondi, P.T. (1995). Responses of settling invertebrate larvae to bioorganic films: Effects of different types of films. <i>J. Exp. Mar. Biol. Ecol.</i> 185, 235–
1021	253.
1022	King, P., Symes, D. (2004). Potential Loss of GNP and GSP from a Failure to
1023	Maintain California's Beaches. Shore and Beach. 72, 3-8.
1024	Krafft, B.A., Krag, L.A. (2021). Antarctic krill (Euphausia superba) exhibit
1025	positive phototaxis to white LED light. Polar Biol., 44, 483–489.
1026	Krediet, C.J., Ritchie, K.B., Paul, V.J., Teplitski, M. (2013). Coral-associated
1027 1028	micro-organisms and their roles in promoting coral health and thwarting diseases. <i>Proc. R. Soc. B Biol. Sci.</i> , 280, 20122328.
1029	LaJeunesse, T.C., Parkinson, J.E., Gabrielson, P.W., Jeong, H.J., Reimer, J.D.,
1030	Voolstra, C.R., Santos, S.R. (2018). Systematic revision of Symbiodiniaceae
1031 1032	highlights the antiquity and diversity of coral endosymbionts. Curr. Biol., 28, 2570e2580.
1002	237002300.
1033	Last, K. S., Hobbs, L., Berge, J., Brierley, A. S., Cottier, F. (2016). Moonlight
1034 1035	drives ocean-scale mass vertical migration of zooplankton during the Arctic winter. <i>Curr. Biol. 26</i> , 244–251.
1033	Curr. Biol. 20, 244–231.
1036	Leopold, M.F., Philippart, C.J.M., Yorio, P. (2010). Nocturnal feeding under
1037 1038	artificial light conditions by Brown-hooded Gull (<i>Larus maculipennis</i>) in Puerto Madryn harbour (Chubut Province, Argentina). <i>Hornero</i> , 25, 55–60.
1039	Levy, O. et al. (2007). Light-responsive cryptochromes from a simple
1040	multicellular animal, the coral Acropora millepora. Science, 318, 467–470.
1041	Levy, O., Marangoni, L.F.B., Benichou, J. I., Rottier, C., Béraud, E., Grover, R.,
1042	et al. (2020). Artificial light at night (ALAN) alters the physiology and biochemistry
1043	of symbiotic reef building corals. Environ. Poll., 266, 114987.
1044	Limpus, C., Kamrowski, R. L. (2013). Ocean-finding in marine turtles: the
1045	importance of low horizon elevation as an orientation cue. Behaviour, 15, 863-893.
1046	Lin, C-H, Takahashi, S., Mulla, A. J., Nozawa, Y. (2021). Moonrise timing is key
1047	for synchronized spawning in coral Dipsastraea speciosa. PNAS, 118, e2101985118.
1048	Lohmann, K. J., Witherington, B. E., Lohmann, C. M., Salmon, M. (2017).
1049	Orientation, navigation, and natal beach homing in sea turtles. In The biology of sea
1050	turtles. CRC Press. pp. 108-135.
1051	Longcore, T., Rich, C. (2004). Ecological light pollution. Front. Ecol. Environ.,
1052	2, 191–198.

1053 1054 1055 1056	Longcore, T., Duriscoe, D., Aubé, M., Jechow, A., Kyba, C., Pendoley, K. L. (2020). Commentary: Brightness of the night sky affects loggerhead (Caretta caretta) sea turtle hatchling misorientation but not nest site selection. <i>Frontiers in Marine Science</i> , 7.
1057 1058 1059	Longcore, T., Rodríguez, A., Witherington, B., Penniman, J.F., Herf, L., et al. (2018). Rapid assessment of lamp spectrum to quantify ecological effects of light at night. <i>J. Exp. Zool. Part A Ecol. Integr. Physiol.</i> , <i>329</i> , 511–521.
1060 1061 1062	Luarte, T., Bonta, C.C., Silva-Rodriguez, E.A., Quijón, P.A., Miranda, C., Farias, A.A., Duarte, C. (2016). Light pollution reduces activity, food consumption and growth rates in a sandy beach invertebrate. <i>Environ. Pollut., 218</i> , 1147-1153.
1063 1064 1065	Lucas, M., Salmon, M., Fritz, E., Wyneken, J. (1992). Sea finding by hatchling sea turtles: role of brightness, silhouette and beach slope as orientation cues. <i>Behaviour, 122</i> , 56-77.
1066 1067 1068	Ludvigsen, M., Berge, J., Geoffroy, M. et al. (2018). Use of an autonomous surface vehicle reveals small-scale diel vertical migrations of zooplankton and susceptibility to light pollution under low solar irradiance. <i>Sci. Advanc., 4</i> , eaap9887.
1069 1070 1071	Lynn, K.D, Flynn, P.T., Manríquez, K., Manríquez, P.H., Pulgar, J., Duarte, C., Quijon, P.A. (2021a). Artificial light at night alters the settlement of acorn barnacles on a man-made habitat in Atlantic Canada. <i>Mar. Poll. Bull.</i> , <i>163</i> , 111928.
1072 1073 1074 1075	Lynn, K.D., Quintanilla-Ahumada, D., Anguita, C., Widdicombe, S., Pulgar, J., Manríquez, P.H., Quijón, P.A., Duarte, C. (2021b). Disruption and recovery: Artificial Light at Night (ALAN) alter the activity and feeding behavior of sandy beach Amphipods from Atlantic Canada. <i>Sci. Total Environ.</i> , <i>780</i> , 146568.
1076 1077	Maggi, E., Benedetti-Cecchi, L. (2018). Trophic compensation stabilizes marine primary producers exposed to artificial light at night. <i>Mar. Ecol. Prog. Ser.</i> , 606, 1–5.
1078 1079 1080	Maggi, E., Bongiorni, L., Fontanini, D., Capocchi, A., Dal Bello, M., Giacomelli, A., et al. (2020a). Artificial light at night erases positive interactions across trophic levels. <i>Funct. Ecol.</i> , <i>34</i> , 694–706.
1081 1082 1083	Maggi, E., Bertocci, I., Benedetti-Cecchi, L. (2020b). Light pollution enhances temporal variability of photosynthetic activity in mature and developing biofilm. <i>Hydrobiol.</i> 847, 1793–1802.
1084 1085 1086	Mavraki N, Georgiadis M, Koutsikopoulos C, Tzanatos E. (2016). Unravelling the nocturnal appearance of bogue Boops boops shoals in the anthropogenically modified shallow littoral. <i>J Fish Biol.</i> 88, 2060-6.

Global Change Biology

1087 1088 1089	McLachlan, A., Defeo, O., Jaramillo, E., Short, A.D. (2013) Sandy beach conservation and recreation: guidelines for optimizing management strategies for multi-purpose use. <i>Ocean Coast Manag.</i> 71, 256–268.
1090 1091 1092 1093	Manríquez, K., Quijon, P.A., Manríquez, P.H., Miranda, C., Pulgar, J., Quintanilla- Ahumada, D., Duarte, C. (2021). Artificial Light at Night (ALAN) negatively affects the settlement success of two prominent intertidal barnacles. <i>Mar. Poll. Bull.</i> , <i>168</i> , 112416.
1094 1095 1096 1097	Manríquez, P.H., Jara M.J., Gonzalez G.P., Seguel M, Quijon, P.A., Widdicombe S., Pulgar J.M., Quintanilla-Ahumada D., Anguita C., Duarte C. (2021) Effects of artificial light at night and predator cues on foraging and predator avoidance in the keystone inshore mollusc <i>Concholepas concholepas</i> . <i>Environ. Poll.</i> , <i>280</i> , 116895.
1098 1099 1100 1101	Manríquez, P.H., Jara, M.E., Díaz, M.I., Quijon, P.A., Widdicombe, S., Manríquez, K., Quintanilla-Ahumada, D., Duarte, C. (2019). Artificial light pollution influences behavioral and physiological traits in a keystone predator species, <i>Concholepas concholepas. Sci. Total Environ,</i> . 661, 543–552.
1102 1103 1104 1105	Manríquez, P.H., Jara, M.E., Mardones, M.L., Torres, R., Navarro, J.M., Lardies, M.A., Vargas, C.A., Duarte, C., Lagos, N.A. (2014). Ocean acidification affects predator avoidance behaviour but not prey detection in the early ontogeny of a keystone species. <i>Mar. Ecol. Prog. Ser., 502</i> , 157–167.
1106 1107 1108	Manríquez, P.H., Lagos, N.A., Jara, M.E., Castilla, J. (2009). Adaptive shell color plasticity during the early ontogeny of an intertidal keystone predator. <i>Proc. Natl. Acad. Sci. U. S. A., 106</i> , 16298–16303.
1109 1110 1111	Marchesan, M., Spoto, M., Verginella, L., Ferreiro, E., A. (2005). Behavioral effects of artificial light on fish species of commercial interest. <i>Fish. Res.</i> , 73, 171-185.
1112 1113	Martin, G.R. (2017). <i>The Sensory Ecology of Birds</i> . Great Clarendon Street, Oxford, OX2 6DP, United Kingdom, Oxford University Press.
1114 1115 1116	Martin, C.W., Reynolds, L.K., Scheffel, W.A. et al. (2021). Diel Variability and Influence of Artificial Light on Fish and Macroinvertebrate Communities in Gulf of Mexico Seagrass Beds. <i>Estuaries and Coasts</i> . <i>44</i> , 431–441.
1117 1118 1119	Menge, B. A., Branch, G. M. (2001). Rocky intertidal communities. In: <i>Marine Community Ecology</i> (M. D. Bertness, S. D. Gaines, M. E. Hay, eds.). Sunderland, MA: Sinauer Associates. pp. 221–251.
1120 1121	Merkel, F.R., Johansen, K.L. (2011). Light-induced bird strikes on vessels in Southwest Greenland. <i>Mar. Pollut. Bull.</i> , <i>62</i> , 2330–2336.

1122	Meschini, E., Gagliardo, A., Papi, F. (2008). Lunar orientation in sandhoppers is
1123	affected by shifting both the moon phase and the daily clock. Anim. Behav. 76, 25-35.
1124	Mundy, C.N., Babcock, R.C. (1998). Role of light intensity and spectral quality
1125	in coral settlement: implications for depth-dependent settlement? J. Exp. Mar. Biol.
1126	<i>Ecol.</i> , 223, 235–255.
1127 1128	Muscatine, L., Grossman, D., Doino, J. (1981). Release of symbiotic algae by tropical sea anemones and corals after cold shock. <i>Mar. Ecol. Prog. Ser.</i> , 77, 233e243.
1129	Nagarkar, S., Williams, G. S., Subramanian, G., Saha, S. K. (2004).
1130	Cyanobacteria-dominated biofilms: A high quality food resource for intertidal
1131	grazers. Hydrobiologia, 519, 89–94.
1132	Navarro-Barranco, C., Hughes, L. E. (2015). Effects of light pollution on the
1133	emergent fauna of shallow marine ecosystems: Amphipods as a case study. Mar.
1134	<i>Poll. Bull.</i> , <i>94</i> , 235-240.
1135	Newell, G.E. (1965). The eye in Littorina littorea. Proc. Zool. Soc. Lond., 144,
1136	75-86.
1137	Nguyen, T. T., Le, MH., Doan, N. X., Pham, H. Q., Vu, M. T. T., Dinh, K. V.
1138	(2020). Artificial light pollution increases the sensitivity of tropical zooplankton to
1139	extreme warming. Environ. Technol. Innov., 20, 101179.
1140	NSW (2016). NSW Planning and Environment. Dark Sky Planning Guideline
1141	Protecting the observing conditions at Siding Spring
1142	https://www.planning.nsw.gov.au/-/media/Files/DPE/Guidelines/dark-sky-planning-
1143	guideline-2016-06.pdf
1144	O'Connor, J.J., Fobert, E.K., Besson, M. Jacob, H., Lecchini, D. (2019). Live
1145	fast, die young: Behavioural and physiological impacts of light pollution on a marine
1146	fish during larval recruitment. Mar. Poll. Bull., 146, 908-914.
1147	Olson, N., Ainsworth, T., Gates, R., Takabayashi, M. (2009). Diazotrophic
1148	bacteria associated with Hawaiian Montipora corals: Diversity and abundance in
1149	correlation with symbiotic dinoflagellates. J. Exp. Mar. Biol. Ecol., 371, 140-146.
1150	Oro, D., de León, A., Minguez, E., Furness, R.W. (2005). Estimating predation
1151	on breeding European storm-petrels (Hydrobates pelagicus) by yellow-legged gulls
1152	(Larus michahellis). J. Zool., 265, 421–429.
1153	Paine, R. T. (1966). Food Web Complexity and Species Diversity. American
1154	Naturalist, 100, 65-&.

1155 1156 1157	Patrício, A. R., Varela, M. R., Barbosa, C., Broderick, A. C., Airaud, M. B. F., Godley, B. J., Catry, P. (2018). Nest site selection repeatability of green turtles, Chelonia mydas, and consequences for offspring. <i>Animal Behaviour</i> , <i>139</i> , 91-102.
1158 1159 1160	Peña, M., Cabrera-Gámez, J., Domínguez-Brito, A. C. (2020). Multi-frequency and light-avoiding characteristics of deep acoustic layers in the North Atlantic. <i>Mar. Environ. Res.</i> , <i>154</i> , 104842.
1161 1162	Pendleton, L., Kildow, J., Rote, J.W. (2006). The non-market value of beach recreation in California. <i>Shore and Beach</i> , <i>74</i> , 34-37.
1163 1164 1165	Pendoley, K., Kamrowski, R. L. (2015). Influence of horizon elevation on the sea- finding behaviour of hatchling flatback turtles exposed to artificial light glow. <i>Mar.</i> <i>Ecol. Prog. Series.</i> , 529, 279-288.
1166 1167 1168	Picheral, M. et al. (2010). The Underwater Vision Profiler 5: An advanced instrument for high spatial resolution studies of particle size spectra and zooplankton. <i>Limnol. Oceanogr. Meth.</i> , <i>8</i> , 462–547.
1169 1170 1171	Pugh, A.R., Pawson, S.M. (2016). Artificial light at night potentially alters feeding behaviour of the native southern black-backed gull (<i>Larus dominicanus</i>). <i>Notornis</i> , <i>63</i> , 37–39.
1172 1173 1174 1175	Pulgar, J., Zeballos, D., Vargas, J., Aldana, M., Manriquez, P.H., Manriquez, K., Quijón, P.A., Widdicombe, S., Anguita, C., Quintanilla, D., Duarte, C. (2019). Endogenous cycles, activity patterns and energy expenditure of an intertidal fish is modified by artificial light pollution at night (ALAN). <i>Environ. Pollut.</i> , 244, 361-366.
1176 1177 1178	Qian, P.Y., Lau, S.C.K., Dahms, H.U., Dobretsov, S., Harder, T. (2007). Marine biofilms as mediators of colonization by marine macroorganisms: Implications for antifouling and aquaculture. <i>Mar. Biotechnol.</i> , <i>9</i> , 399–410.
1179 1180 1181 1182	Quintanilla-Ahumada, D., Quijón, P. A., Pulgar, J., Manríquez, P. H., García-Huidobro, M. R., Duarte, C. (2021). Exposure to artificial light at night (ALAN) alters RNA: DNA ratios in a sandy beach coleopteran insect. <i>Mar. Pollut. Bull.</i> , <i>165</i> , 112132.
1183 1184 1185 1186	Quintanilla-Ahumada, D., Quijón, P.A., Manríquez, P.H., Pulgar, J., García-Huidobro, M.R., Miranda, C., Molina, A., Zuloaga, R., Duarte, C. (2022). Artificial light at night (ALAN) causes variable dose-responses in a sandy beach isopod. <i>Env. Sci. Pollut. Res.</i> DOI:10.1007/s11356-021-17344-2.
1187 1188 1189 1190	Rebke, M., Dierschke, V., Weiner, C.N., Aumüller, R., Hill, K., et al. (2019). Attraction of nocturnally migrating birds to artificial light: The influence of color, intensity and blinking mode under different cloud cover conditions. <i>Biol. Conserv.</i> , <i>233</i> , 220–227.

1191 1192 1193	Regular, P.M., Hedd, A., Montevecchi, W.A. (2011). Fishing in the Dark: A Pursuit-Diving Seabird Modifies Foraging Behaviour in Response to Nocturnal Light Levels. <i>PLoS One</i> , 6, e26763.
1194 1195	Ringelberg, J. (2010). Diel Vertical Migration of Zooplankton in Lakes and Oceans: Causal explanations and adaptations. Springer Science, Dordrecht.
1196 1197 1198	Riou, S., Hamer, K.C. (2008). Predation risk and reproductive effort: impacts of moonlight on food provisioning and chick growth in Manx shearwaters. <i>Anim. Behav.</i> , <i>76</i> , 1743–1748.
1199 1200 1201	Rivas, M.L., Santidrián Tomillo, P., Uribeondo, J.D., Marco, A. (2015). Leatherback hatchling sea-finding in response to artificial lighting: interaction between wavelength and moonlight. <i>J. Exp. Mar. Biol. Ecol.</i> , <i>463</i> , 143-149.
1202 1203 1204	Rodríguez, A., Arcos, J.M., Bretagnolle, V., Dias, M.P., Holmes, N.D., et al. (2019). Future Directions in Conservation Research on Petrels and Shearwaters. <i>Front. Mar. Sci.</i> , <i>6</i> , 94.
1205 1206 1207	Rodríguez, A., Burgan, G., Dann, P., Jessop, R., Negro, J.J., et al. (2014). Fatal attraction of Short-Tailed Shearwaters to artificial lights Schierwater, B. (ed.). <i>PLoS One</i> , <i>9</i> , e110114.
1208 1209 1210	Rodríguez, A., Chiaradia, A., Wasiak, P., Renwick, L., &Dann, P. (2016). Waddling on the Dark Side: Ambient Light Affects Attendance Behavior of Little Penguins. <i>J. Biol. Rhythms.</i> , <i>31</i> , 194–204.
1211 1212 1213	Rodríguez, A., Dann, P., Chiaradia, A. (2017a). Reducing light-induced mortality of seabirds: High pressure sodium lights decrease the fatal attraction of shearwaters. <i>J. Nat. Conserv.</i> , <i>39</i> , 68–72.
1214 1215 1216	Rodríguez, A., Holmberg, R., Dann, P. & Chiaradia, A. (2018). Penguin colony attendance under artificial lights for ecotourism. <i>J. Exp. Zool. Part A Ecol. Integr. Physiol.</i> , 329, 457–464.
1217 1218 1219	Rodríguez, A., Holmes, N.D., Ryan, P.G., Wilson, KJ., Faulquier, L., et al. (2017b). Seabird mortality induced by land-based artificial lights. <i>Conserv. Biol.</i> , <i>31</i> , 986–1001.
1220 1221	Rodríguez, A., Rodríguez, B., & Negro, J.J. (2015). GPS tracking for mapping seabird mortality induced by light pollution. <i>Sci. Rep.</i> , <i>5</i> , 10670.
1222 1223 1224	Rodríguez, A., Rodríguez, B., Acosta, Y., Negro, J.J. (2022). Tracking Flights to Investigate Seabird Mortality Induced by Artificial Lights. <i>Front Ecol Evol.</i> , <i>9</i> , 786557.

1225 1226 1227	Ronconi, R.A., Allard, K.A., Taylor, P.D. (2015). Bird interactions with offshore oil and gas platforms: Review of impacts and monitoring techniques. <i>J. Environ. Manage.</i> , <i>147</i> , 34–45.
1228 1229 1230	Rooper, C. N., Williams, K., De Robertis, A., Tuttle, V. (2015). Effect of underwater lighting on observations of density and behavior of rockfish during camera surveys. <i>Fish. Res.</i> , <i>172</i> , 157-167.
1231 1232 1233	Rosenberg, Y., Doniger, T., & Levy, O. (2019). Sustainability of coral reefs are affected by ecological light pollution in the Gulf of Aqaba/Eilat. <i>Commun. Biol.</i> , <i>2</i> , 289.
1234 1235 1236 1237	Rubolini, D., Maggini, I., Ambrosini, R., Imperio, S., Paiva, V.H., et al. (2015). The Effect of Moonlight on Scopoli's Shearwater <i>Calonectris diomedea</i> Colony Attendance Patterns and Nocturnal Foraging: A Test of the Foraging Efficiency Hypothesis. <i>Ethology</i> , <i>121</i> , 284–299.
1238 1239	Ryan, P. G., Ryan, E. M., Glass, J. P. (2021). Dazzled by the light: the impact of light pollution from ships on seabirds at Tristan da Cunha. <i>Ostrich.</i> , <i>92</i> , 218–224.
1240 1241	Ryer, C. H., Stoner, A. W., Iseri, P. J., Spencer, M. L. (2009). Effects of simulated underwater vehicle lighting on fish behavior. <i>Mar. Ecol. Progr. Ser.</i> , <i>391</i> , 97-106.
1242 1243 1244	Sainmont, J. et al. (2014). Inter- and intra-specific diurnal habitat selection of zooplankton during the spring bloom observed by Video Plankton Recorder. <i>Mar. Biol.</i> , <i>161</i> , 1931–1941.
1245 1246 1247	Salmon, M., Wyneken, J. (1987). Orientation and swimming behavior of hatchling loggerhead turtles <i>Caretta caretta</i> L. during their offshore migration. <i>J. Exp. Mar. Biol. Ecol.</i> , <i>109</i> , 137-153.
1248 1249	Salmon, M. (2003). Artificial night lighting and sea turtles. <i>Biologist</i> , 50, 163-168.
1250 1251 1252 1253	Salmon, M., Tolbert, M. G., Painter, D. P., Goff, M., Reiners, R. (1995). Behavior of loggerhead sea turtles on an urban beach. II. Hatchling orientation. <i>J. Herpet.</i> , <i>29</i> , 568-576. Sanders, D., Frago, E., Kehoe, R., Patterson, C., & Gaston, K. J. (2021). A
1254 1255	meta-analysis of biological impacts of artificial light at night. <i>Nat. Ecol. Evol., 5</i> , 74-81.
1256 1257 1258 1259 1260	Schlacher, T.A., Lucrezi, S., Connolly, R.M., Peterson, C.H., Gilby, B.L., Maslo, B., Olds, A.D., Walker, S.J., Leon, J.X., Huijbers, C.M., Weston, M.A., Turra, A., Hyndes, G.A., Holt, R.A., & Schoeman, D.S. (2016). Human threats to sandy beaches: A meta-analysis of ghost crabs illustrates global anthropogenic impacts. <i>Estuar. Coast. Shelf Sci.</i> , <i>169</i> , 56–73.

1261	Schligler, J., Cortese, D., Beldade, R., Swearer, S.E., Mills, S.C. (2021). Long-
1262	term exposure to artificial light at night in the wild decreases survival and growth of a
1263	coral reef fish. Proc. R. Soc. B., 288, 20210454.
1264	Schmid, M. S., Aubry, C., Grigor, J. & Fortier, L. (2016). The LOKI underwater
1265	imaging system and an automatic identification model for the detection of zooplankton
1266	taxa in the Arctic Ocean. Meth. Oceanogr., 15-16, 129-160.
1267	Schulz, J. et al. (2010). Imaging of plankton specimens with the light frame on-
1268	sight key species investigation (LOKI) system. J. Eur. Opt. Soc. 5, 10017s.
1269	Shimada, T., Meekan, M. G., Baldwin, R., Al-Suwailem, A. M., Clarke, C.,
1270	Santillan, A. S., Duarte, C. M. (2021). Distribution and temporal trends in the
1271	abundance of nesting sea turtles in the Red Sea. Biol. Conserv., 261, 109235.
1272	Smyth, T. J., Wright, A. E., McKee, D., Tidau, S., Tamir, R., Dubinsky, Z.,
1273	. Davies, T. W. (2021). A global atlas of artificial light at night under the sea.
1274	Elementa: Science of the Anthropocene, 9, 00049. DOI:
1275	https://doi.org/10.1525/elementa.2021.00049
1276	Stickney, A. P. (1969). Factors influencing the attraction of Atlantic Herring.
1277	Fish. Bull., 68, 73-85.
1278	Stoner, A. W., Ryer, C. H., Parker, S. J., Auster, P. J., & Wakefield, W. W. (2008).
1279	Evaluating the role of fish behavior in surveys conducted with underwater vehicles.
1280	Can. J. Fish. Aquat., 65, 1230-1243.
1281	Syposz, M., Padget, O., Willis, J., Van Doren, B. M., Gillies, N., Fayet, A. L., et al.
1282	(2021). Avoidance of different durations, colors, and intensities of artificial light by adult
1283	seabirds. Sci. Rep., 11, 18941.
1284	Szekeres, P., Wilson, A.D.M., Haak, C.R., Danylchuk, A.J., Brownscombe, J.W.,
1285	Elvidge, C.K., Shultz, A.D., Birnie-Gauvin, Kim, Cooke, S.J. (2017). Does coastal
1286	light pollution alter the nocturnal behavior and blood physiology of juvenile bonefish
1287	(Albula vulpes)? Bull. Mar. Sci., 93, 491-505.
1288	Tamir, R., Eyal, G., Cohen, I., & Loya, Y. (2020). Effects of light pollution on
1289	the early life stages of the most abundant Northern Red Sea Coral. Microorganisms,
1290	8, 193.
1291	Tamir, R., Lerner, A., Haspel, C., Dubinsky, Z., & Iluz, D. (2017). The spectral
1292	and spatial distribution of light pollution in the waters of the northern Gulf of Aqaba
1293	(Eilat). Sci. Rep., 7, 42329.
1294	Thompson, J.R., Rivera, H.E., Closek, C.J., & Medina, M. (2015). Microbes in
1295	the coral holobiont: Partners through evolution, development, and ecological
1296	interactions. Front. Cell. Infect. Microbiol., 4, 176.

1297 1298	Thorson, G. (1964). Light as an ecological factor in the dispersal and settlement of larvae of marine bottom invertebrates. <i>Ophelia</i> , <i>1</i> , 167–208.
1299 1300	Torres, D., Tidau, S., Jenkins, S., & Davies, T. (2020). Artificial skyglow disrupts celestial migration at night. <i>Curr. Biol.</i> , <i>30</i> , R696-R697.
1301 1302 1303	Trembley, P., Grover, R., Maguer, JF., Legendre, L., & Ferrier-Page, C. (2012). Autotrophic carbon budget in coral tissue: a new ¹³ C-based model of photosynthates translocation. <i>J. Exp. Biol.</i> , <i>215</i> , 1384e1393.
1304 1305 1306	Trenkel, V. M., Lorance, P. & Mah.vas, S. (2004). Do visual transects provide true population density estimates for deepwater fish?. ICES <i>J. Mar. Sci., 61</i> , 1050–1056.
1307 1308 1309	Truscott, Z., Booth, D. T., & Limpus, C. J. (2017). The effect of on-shore light pollution on sea-turtle hatchlings commencing their off-shore swim. <i>Wildlife Res.</i> , <i>44</i> , 127-134.
1310 1311 1312	Ugolini, A., Fantini, T., & Innocenti, R. (2003). Orientation at night: an innate moon compass in sandhoppers (Amphipoda: Talitridae). <i>Proc. R. Soc. Lond.</i> 270, 279-281.
1313 1314	Underwood, C. N., Davies, T. W., & Queirós, A. M. (2017). Artificial light at night alters trophic interactions of intertidal invertebrates. <i>J. Anim. Ecol.</i> 86, 781–789.
1315 1316	Underwood, M. J., Utne Palm, A. C., Øvredal, J. T., & Bjordal, Å. (2020). The response of mesopelagic organisms to artificial lights. <i>Aquac. Fish.</i> , <i>6</i> , 519-529.
1317 1318 1319	Utne-Palm, A. C., Breen, M., Løkkeborg, S., & Humborstad, O. B. (2018). Behavioral responses of krill and cod to artificial light in laboratory experiments. <i>PLoS ONE</i> , <i>13</i> , e0190918.
1320 1321 1322	Utne-Palm, A. C., Breen, M., Løkkeborg, S., & Humborstad, O. B. (2018). Behavioural responses of krill and cod to artificial light in laboratory experiments. PLoS ONE, <i>13</i> .
1323 1324	Voigt, C.C., Currie, S.E., Fritze, M., Roeleke, M., Lindecke, O. (2018). Conservation strategies for bats flying at high altitudes. <i>BioScience</i> , <i>68</i> , 427-435.
1325 1326	Wells, R. (1980). Activity pattern as a mechanism of predator avoidance in two species of acmaeid limpet. <i>J. Exp. Mar. Biol. Ecol.</i> , 48, 151–158.
1327 1328 1329	Widder, E. A., Robison, B. H., Reisenbichler, K. R. & Haddock, S. H. D. (2005). Using red light for in situ observations of deep-sea fishes. <i>Deep-Sea Res. Part I, 52</i> , 2077–2085.

1330	Wijgerde, T. et al. (2014). Red light represses the photophysiology of the
1331	scleractinian coral Stylophora pistillata. PLOS ONE, 9, e92781.
1332	Williams, K., Rooper, C. N. & Towler, R. (2010). Use of stereo camera systems
1333	for assessment of rockfish abundance in untrawlable areas and for recording pollock
1334	behaviour during midwater trawls. Fish. Bull., 108, 352-362.
1335	Willmott, N.K., Henneken, J., Selleck, C.J. & Jones, T.M. (2018). Artificial light
1336	at night alters life history in a nocturnal orb-web spider. Peer J, 6, e5599.
1337	Wilson, P., Thums, M., Pattiaratchi, C., Meekan, M., Pendoley, K., Fisher, R., &
1338	Whiting, S. (2018). Artificial light disrupts the nearshore dispersal of neonate flatback
1339	turtles Natator depressus. Mar. Ecol. Progr. Series., 600, 179-192.
1340	Wilson, P., Thums, M., Pattiaratchi, C., Whiting, S., Meekan, M., & Pendoley, K.
1341	(2021). Nearshore wave characteristics as cues for swimming orientation in flatback
1342	turtle hatchlings. J. Exp. Mar. Biol. Ecol., 535, 151475.
1343	Windle, A. E., Hooley, D. S., & Johnston, D. W. (2018). Robotic vehicles enable
1344	high-resolution light pollution sampling of sea turtle nesting beaches. Front. Mar. Sci.,
1345	5, 493.
1346	Witherington, B. E. & Bjorndal, K. A. (1991a). Influences of artificial lighting on
1347	the seaward orientation of hatchling loggerhead turtles Caretta caretta. Biol. Conserv.,
1348	55, 139-149.
1349	Witherington, B. E. & Bjorndal, K. A. (1991b). Influences of wavelength and
1350	intensity on hatchling sea turtle phototaxis: implications for sea-finding behavior.
1351	<i>Copeia</i> , <i>1991</i> , 1060-1069.
1352	Wood, D. W. & Bjorndal, K. A. (2000). Relation of temperature, moisture,
1353	salinity, and slope to nest site selection in loggerhead sea turtles. Copeia, 2000, 119.
1354	Wyneken, J. & Salmon, M. (1992). Frenzy and post renzy swimming activity in
1355	loggerhead, green, and leatherback hatchling sea turtles. Copeia, 2, 478-484.

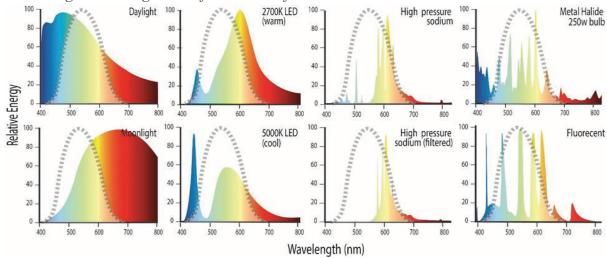
Box 1. How research activities can interfere and create bias due to artificial light

Despite a growing body of literature reporting behavioral disturbance of marine organisms exposed to artificial light, external light sources remain widely used in oceanography and marine ecology studies. Advances in optical technology, combined with the increased desire to use non-lethal observation approaches, have driven the development of new sensors and instruments to document marine ecosystems (Bicknell et al. 2016), but these instruments generally require an external light source. For example, Optical probes such as the Underwater Vision Profiler (Picheral et al. 2010), the Laser-Optical Plankton Counter (Herman & Harvey 2006, Basedow et al 2013), the Video Plankton Recorder (Sainmont et al. 2014), and the Light frame On-sight Key species Investigation system (Schulz et al. 2010, Schmid et al. 2016) all use light sources and optical sensors to assess the vertical distribution and abundance of zooplankton. Researchers and the industry alike increasingly use High Definition (HD) video cameras or stereo-cameras mounted on trawls to document the catchability of different species or size classes of fish (Underwood et al. 2020, Williams et al. 2010, Boldt et al. 2018). Such camera systems, when used in dim environments, rely on external light sources to distinguish, and identify marine animals at depth. Although previous studies have raised concerns about the impact of artificial visible light on measurements from optical instruments (Boldt et al. 2018, Trenkel et al. 2004, Widder et al. 2005, Benoit-Bird et al. 2010, Doya et al. 2014), these biases have rarely been quantified (Bicknell et al. 2016). Nonetheless, artificial lighting is assumed to be the main source of biases in fish surveys using cameras and underwater vehicles (Ryer et al. 2009, Stoner et al. 2008, Rooper et al. 2015). The use of red light has been suggested for marine surveys requiring external light sources because it is assumed that most species do not react as much to red light as to shorter wavelengths, such as blue or green (Boldt et al. 2018, Widder et al. 2005, Rooper et al. 2015). In support of this hypothesis, Pena et al. (2020) and Underwood et al. (2020) deployed oceanographic probes equipped with different light colors and showed that mesopelagic (200–1000 m) fish avoid white, blue and green, but not red light. This, however, was recently challenged by Geoffroy et al. (2021) who, using hull-mounted echosounders above an acoustic probe or a baited video camera, each equipped with light sources of different colors (white, blue, and red), demonstrated that pelagic organisms in Arctic and temperate regions strongly avoid artificial light, including visible red light (575– 700 nm), from instruments lowered in the water column. Light levels varied within the range of 11-18 µW cm⁻² for the colors tested (see Geoffroy et al., 2021 for details). The density of organisms decreased by up to 99% when exposed to artificial light and the distance of avoidance varied from 23 to 94 m from the light source, depending on colors, irradiance levels and, possibly, species communities (Geoffroy et al. 2021).

Box 2: Challenges in measuring artificial light in biological studies

Measuring light for ecological studies is still undefined and is poorly understood by biologists who have used a wide range of techniques and instruments to measure light at scales ranging from a few centimeters to tens of kilometers. The wide range of instruments employed typically use different measurement systems and units which means study results cannot be confidently compared. There is currently no globally recognized standard method, or agreed unit of measurement, for monitoring biologically active light (Barentine, 2019).

Visible light can be described by 2 physical parameters: wavelength and intensity. The relative distribution and weighting of different wavelengths in emitted light determines its color; however, how this light is perceived by the observer is also a function of the physiology of the receiving detector (e.g., eye). The intensity of light reaching a detector is a function of distance from the light source as light waves spread out from the light source the number of waves falling on a defined area decreases proportional to the distance travelled. Light is measured either radiometrically or photometrically. Radiometry is the measurement of wavelengths across the entire electromagnetic spectrum. In biological applications this is typically restricted to the ultraviolet, visible, and infrared region of the spectrum between 350 nm and 800 nm and is measured in watts per meter squared. Most commercial light measurement equipment records photometric light which is visible light wavelengths weighted specifically to the sensitivity of the human eye (CIE, 2004) and is reported in units of lumens per meter squared or Lux. Photometric detectors have reduced sensitivity to wavelengths below 450nm (blue light) or above 650 nm (red light). Consequently, photometric instruments commonly used to quantify light in biological studies, such as lux meters, do not account for the blue light that is most visible to biological receptors. The figure below demonstrates this. Commercial lux meters only quantify light that is within the CIE curve (area under grey dashed line) and exclude light wavelengths that fall outside of the CIE curve.



Credit: Kellie Pendoley.

Lux meters are further limited in that they were designed for use in measuring light in buildings, have poor sensitivity to low light levels and cannot detect sky glow or dim light in a field setting. Finally, they do not provide any spectral information, i.e., the relative distribution of light wavelengths, in the light source.

The limitations and challenges around measuring ecological light on scales ranging from landscape to small scale is discussed in more detail in Commonwealth (2020); Longcore et al (2020), Hanel et al (2018), Jechow and Holker (2019) and Jechow et al. (2019).