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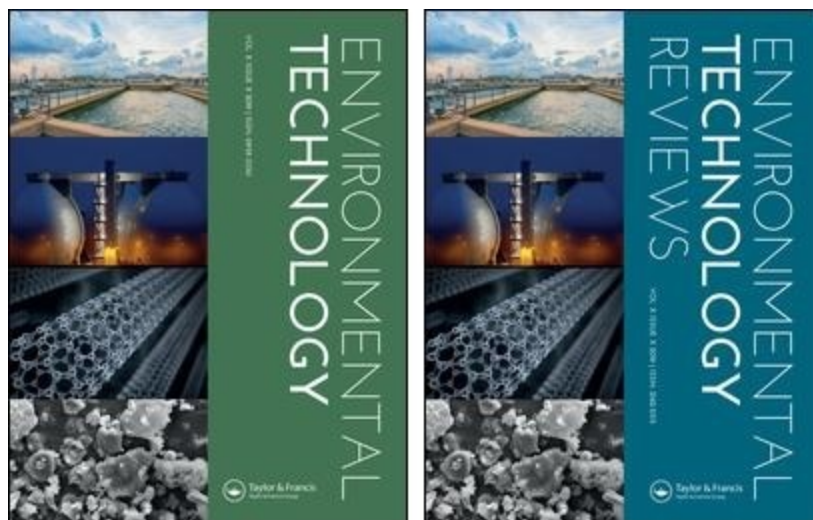
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Developing the OECD 106 fate testing protocol for active pharmaceuticals in soil

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Abstract

The ability to determine accurately the fate of APIs in soil is essential for rigorous risk assessment associated with wastewater reuse or biosolid recycling to land, particularly in lower income countries where water and fertiliser is scarce. Four APIs (naproxen, ofloxacin, propranolol and nevirapine) with wide ranging functionality were used as examples in the development of the OECD 106 soil partitioning and/or degradation study, with naproxen used to illustrate applying the full methodology. The data showed key methodological criteria require careful consideration and testing to generate accurate and consistent results. Only glass fibre membranes were suitable for all APIs, without unduly adsorbing APIs to their surface, thus effectively restricting the minimum practical pore size to 0.7 μm . Polypropylene plastic centrifuge tubes were shown to be suitable, with careful determination of recoveries. Direct injection liquid chromatography-mass spectrometry could reliably resolve all 4 APIs down to less than $\mu\text{g L}^{-1}$ in soil solutions, although allowance for matrix effects via standard additions was required in some cases. Greatest analytical challenges were found for the highest molecular weight API with the greatest affinity for sorption to surfaces (ofloxacin). Key variables that can impact on partitioning such as solution pH and dissolved organic carbon concentrations were shown to vary within tests over time and should be accounted for.

Key words: soil; naproxen; method development; pharmaceuticals; liquid chromatography-mass spectrometry; OECD106

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1 Introduction

There is substantial expansion of the use of active pharmaceutical ingredients (APIs) in low and lower-middle income countries (LLMIC) of Asia, Africa, Latin America and Australasia over the past two decades. In 2000, these countries accounted for 25 % of global pharmaceutical sales; in 2014 this figure was 32 %, with sales accounting for at least 25 % of the income of the largest pharmaceutical companies [1].

Many LLMICs are experiencing physical or economic water scarcity; to counter-act shortages many LLMICs use wastewater for irrigation. Globally, about 36 million ha of agricultural land is irrigated with wastewater [2]. The World Health Organisation has published guidelines on the safe use of wastewater in order to provide a management framework for safeguarding human health, while maximising the benefits of wastewater to agriculture [3-5]. Loadings of APIs in wastewater have been considered [2,3], but environmental risk assessments (ERA) for API inputs to soils and waters remain elusive in LLMICs [6-8]. However, even in high income countries it is only since 1998 and 2006, respectively, that the USA and European Union (EU) have required ERAs for all new marketing authorisation applications involving human medicinal products [9]. An assumption for ERAs in the EU is that wastewater is universally treated in wastewater treatment plants (WWTP) [10], an expectation that does not hold for LLMIC. Furthermore, the EU ERA is focussed only on exposure to APIs following application of sewage biosolids to soil [11], and does not include additional scenarios, such as irrigation of soils with wastewater and other contaminated water sources. In 2015, the UNEP International Conference on Chemicals Management highlighted the persistence and antimicrobial resistance of APIs as priority issues, and called for increased global knowledge of pharmaceuticals in all environmental compartments [12]. Understanding the mobility and fate of APIs in soil is therefore critical for any ERA in LLMICs.

The concentrations and distributions of APIs in soils are controlled by a number of processes, with sorption to soil solids [13], biodegradation and dilution/percolation through the soil profile particularly important [14,15]. The Organisation for Economic Co-operation and Development (OECD) 106 guideline was developed to assess the importance of the sorption (adsorption-desorption) of chemicals to soils using a batch-equilibrium method [16]. It is now a standard method, in both industrial and research contexts, for examining the sorption of APIs in soil. It has been endorsed for use by the EU [10, 17]. Notably, the method embraces the variety of soil types (with respect to organic carbon content, clay context, soil texture and pH) found in regions that rely on wastewater for irrigation [16,18].

It is important to note that OECD 106 is a guideline and not a Standard Operating Procedure. For this reason, Tier 1 Preliminary studies are outlined in OECD 106, to be undertaken prior to the Tier 2 Screening test and the Tier 3 Determination of Freundlich adsorption isotherms. Tier 1 studies include examining the adsorption of the test chemical on the surface of the test vessel, and the stability of the test substance. While not explicitly part of the Tier 1 assessment, it is made clear in the OECD 106 guideline that the development of an appropriate analytical method, with appropriate accuracy (bias, precision), reproducibility, recovery and detection limits, is also key to the successful performance of the test.

The study has developed a method, for testing APIs under the OECD 106 guideline, for examining the sorption of APIs with contrasting physico-chemical characteristics to two different soil types, taking account of all aspects of the process including laboratory ware, filter media and analytical optimisation. To demonstrate the effectiveness of the method development, 4 APIs of differing physico-chemical characteristics ofloxacin, propranolol, naproxen and nevirapine were used. Furthermore, the potential variation in pH and dissolved organic carbon (DOC), both of which could influence the sorption characteristic of the APIs of interest was monitored over the 120 hour test period.

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83 **2 Methods**

84 **2.1 API selection**

85 The APIs studied represented different therapeutic classes and also had contrasting

86 physico-chemical properties particularly the ionic charge at environmental pH (typically 5 –

87 9). Ofloxacin is a broad spectrum fluoroquinolone antibiotic, widely used in India and China

88 [19], while propranolol is an extensively used beta-blocker, with reported poor removal in

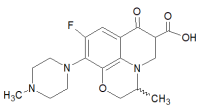
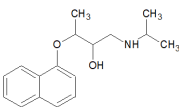
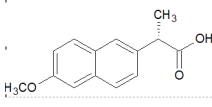
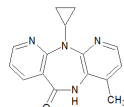
89 LLMIC WWTPs (Table 1) [20,21]. Naproxen is a non-steroidal anti-inflammatory drug

90 routinely prescribed in Bangladesh, Pakistan, the Philippines and Thailand [22]. Finally,

91 nevirapine (Table 1) is one of the most commonly prescribed antiretroviral drugs for preventing

92 HIV-1 transmission in resource-poor countries [23].

Table 1. Test API physico-chemical characteristics. Log K_{oc} = log organic carbon:water partition coefficient. Log K_{ow} = log octanol:water partition coefficient. * These values are not consistent with the low K_{ow} and may reflect partitioning into other soil components (e.g. clay) in addition to, or instead of, organic carbon.

	Ofloxacin	Propranolol	Naproxen	Nevirapine
Structure of neutral ion				
Ionic charge	Zwitterion	cation	anion	neutral
Formula	$C_{18}H_{20}FN_3O_4$	$C_{16}H_{21}NO_2$	$C_{14}H_{14}O_3$	$C_{15}H_{14}N_4O$
CAS No.	82419-36-1	318-98-9	22204-53-1	129618-40-2
MW (g mol ⁻¹)	361.4 ¹	260.2 ¹	231.1 ¹	267.1 ¹
Water sol. (mg L ⁻¹)	25,000 ¹	6,000 ¹	<3,000 ¹	0.7 ¹¹
pK _a	5.97, 8.28 ²	9.53 ⁴	4.15 ⁴	2.8 ⁹
Log K_{oc}	4.64-5.70* ^{2,3}	3.21-4.69 ²	1.98-2.72 ^{5,6,7}	n/a
Log K_{ow}	-0.39-0.65 ⁴	2.58-3.48 ⁴	2.99-3.18 ⁴	1.75-2.5 ⁸

¹[24-27]; ²[17]; ³[28]; ⁴[29]; ⁵[30]; ⁶[31]; ⁷[32]; ⁸[33]; ⁹[34].

2.2 Soil selection

Soils used for testing should be representative of the environments where the substances of interest will be applied, as well as being free from anthropogenic influences to reduce the risk of soils being contaminated with test substance. In the current study, two soils were chosen according to the OECD 106 guideline [16]. These soils were purchased from Lufa Speyer (Germany) and had a ≥ 5 year history of no pesticide, biocidal fertiliser, or organic manure application, resulting in a soil that should be as 'clean' from contaminants as possible. Sample handling by Lufa Speyer followed ISO 10381-6 (1993) and Good Laboratory Practice. Soil samples were provided air - dried and sieved to < 2 mm.

Table 2 shows the properties of the two soils, how they match with the OECD 106 Guideline and which LLMICs have similar soils. Both soils matched the requirements specified by OECD 106, except for the clay content of the loam soil, which was 0.8 % above the maximum.

Table 2. Properties of the two soils used in this study in comparison with the OECD 106 guideline (mean values of different batch analyses \pm s.d.)

	Sandy loam	Loam	OECD loamy sand	OECD silt loam
pH (0.01 M CaCl₂)	5.7 \pm 0.6	7.3 \pm 0.1	< 4.0 - 6.0	5.5 - 7.0
TOC (%)	0.7 \pm 0.0	2.0 \pm 0.2	< 0.5 - 1.5	1.5 - 3.0
Clay content (%)	6.5 \pm 1.6	25.8 \pm 1.8	< 10 - 15	15 - 25
Cation exchange capacity (MEq/100 g)	7.5 \pm 0.9	33.0 \pm 4.5		
LLMIC examples with similar soil properties	SE China (Guangxi and Yunnan provinces). NW India (Odisha and West Bengal). NE Democratic Republic of Congo	Malaysia El Salvador		

¹ [35]; ² [16]; ³ [36].

2.3 Laboratory ware

All re-usable plastic and glass apparatus was cleaned (2 % v/v Decon® for \geq 24 h, rinsed with high purity water (HPW, 18.2 M Ω .cm resistivity), then soaked for \geq 24 h in 10 % v/v HCl, and finally rinsed with HPW). Apparatus was dried in a Class 100 laminar flow hood, as a final step, and stored in resealable plastic bags. Sterile polypropylene centrifuge tubes (50 mL) were used, as were sterile syringes constructed only of polypropylene and polyethylene (i.e. no latex, rubber silicone, styrene or DEHP). Filter holders were also polypropylene and clear glass autosampler vials with silicon septa were used to minimise API losses (all purchased from Fisher Scientific, UK). Glass fibre filter membranes (GF/F, 0.7 μ m nominal pore

diameter: Fisher Scientific, UK) were wrapped in aluminium foil and ashed at 450 °C for 6 h before use. Polycarbonate and cellulose nitrate filters were prepared by being washed with approximately 1 mL of sample before collection.

2.3.1 Laboratory ware selection

Polypropylene centrifuge tubes (50mL) were chosen as they could hold the correct amount of solution and soil with enough head space left for adequate mixing on the shaker, when placed horizontally. Although glass may be considered more appropriate for organics such as APIs, they would not be practical in this case. To assess sorption of the APIs to the walls of the centrifuge tubes, 3 x 30 mL 10 mM CaCl₂ (Fisher Scientific, UK) were shaken overnight in separate tubes, then spiked with a mixed API solution and then shaken for a further 4 h at room temperature.

Filter membranes were required that would remove suspended solids from the soil suspensions, but not at the same time sorb the test APIs. The pore size required was 0.7 µm or below for analysis by HPLC-HRAM-MS and fluorescence spectrophotometry. Comparisons were made between concentrations of APIs in a range of standards made up in HPW before and after filtration and recoveries calculated. Three filter types were tested for propranolol, naproxen and ofloxacin; glass fibre (0.7 µm nominal pore diameter), polycarbonate (0.2 µm) and cellulose nitrate (0.2 µm). Only glass fibre filters were assessed for nevirapine.

2.4 Sample storage

To ensure that the samples were stable during storage freshly made mixed stock solutions of the APIs were made in HPW and diluted into a calibration range (0.1, 1, 10, 50, 100 µg L⁻¹). These were analysed and compared against a calibration range that had been made from individual API standard stock solutions (in HPW) and stored for a minimum of 6 months at -20 °C.

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151 **2.5 Instrumentation - Liquid chromatography – mass spectrometry**

152 A high performance liquid chromatography-high resolution accurate mass-mass
153 spectrometer (HPLC-HRAM-MS) was used to detect and quantify APIs. This consisted of an
154 Ultimate U3000 UHPLC liquid chromatography system and a Q Exactive Focus Orbitrap mass
155 spectrometer (both Thermo Scientific, UK). Gaseous phase ions were generated from solution
156 by electrospray ionisation before detection by mass spectrometry, allowing for the sensitive
157 analysis of both ionised species and neutral compounds [37]. The HRAM detection simplified
158 sample preparation, allowing for lower waste solvent volumes and shorter analysis times, as
159 neither solid phase extraction nor pre-concentration were not required at experimental
160 concentrations. Peak detection was achieved using Xcalibur software (Figure S1 of the
161 electronic supporting information).

162 **2.6 Summary of the OECD 106 soil – solution test procedure requirements**

163 The OECD adsorption-desorption test procedure is described below, although API data are not
164 presented here. The method was run without the addition of APIs but including the sediment
165 and water in order to monitor any changes in pH and dissolved organic carbon concentrations
166 in the test solutions over the 120 hour test period as these are critical parameters controlling
167 the partitioning of organics.

168 **2.2.1 Adsorption**

169 The principle of this method is that a known concentration of a test substance is added
170 to a known weight of soil in a 10 mM CaCl₂ solution. The soil suspension is shaken for a set
171 period, and then centrifuged (and perhaps filtered) prior to analysis. It is an indirect method as
172 only the aqueous phase is analysed and the difference between the amount of API added and
173 recovered from the liquid phase is assumed to be the amount adsorbed to the soil.

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3 174 A 6 g aliquot of soil are added to 50 mL polypropylene centrifuge tubes along with 30
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5 175 mL of 10 mM CaCl_2 solution, in triplicate for each time point. These are put onto a reciprocal
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7 176 shaker (132 rpm), laid horizontally to allow the system to equilibrate overnight. Compounds
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9 177 of interest are added to the tubes, which were then returned to the shaker. Tubes were sacrificed
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11 178 at pre-selected times after API addition (0, 3, 6, 24, 48, 72, 96, 120 hours) centrifuged (4000
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13 179 RPM, 15 minutes) then filtered using 0.7 μm glass fibre filters (Fisher Scientific, UK). Samples
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15 180 are stored at -20°C until analysis. Experiment take place at room temperature ($20 - 25^\circ\text{C}$) and
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17 181 in the dark to ensure that photodegradation of the APIs did not occur.
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22 182 2.2.2 Desorption

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24 183 Desorption of APIs after sorption equilibrium has been reached provides information
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26 184 on how reversible the sorption of APIs is to the soils. This is important to know because it can
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28 185 provide ERAs with more data on the overall fate and mobility of an API in soils, especially
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30 186 those in wet climates or with repeated wastewater irrigation schemes. The OECD 106 method
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32 187 can take this into account (OECD, 2000). Soils are prepared as above and spiked with the same
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34 188 concentration of APIs after the system was shaken overnight. The tubes are then shaken in the
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36 189 dark at room temperature ($18-22^\circ\text{C}$) until adsorption equilibrium is reached and then all the
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38 190 tubes centrifuged (4000 RPM, 15 mins) and the supernatant removed, a subsample is taken and
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40 191 filtered to be analysed on HPLC-MS. The exact volume of 10 mM CaCl_2 was then replaced
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42 192 and the soil pellet is suspended again by 5 seconds on a minishaker. Samples are then sacrificed
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44 193 at 0, 3, 6, 24, 48, 72, 96 and 120 hours, centrifuged and filtered as before storing at -20°C until
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46 194 analysis along with matching calibration solutions.
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52 195 For this method development, to identify changes in the soil solutions after 10 mM
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54 196 CaCl_2 replacement, pH and DOC was measured from separate tubes handled at the same time
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56 197 but without APIs in them. pH and DOC were determined in the soil-water solutions at the pre-
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58 198 selected times using a Shimadzu TOC (total organic carbon) 5000A analyser for DOC and a
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199 calibrated HANNA HI 9025 microcomputer pH meter fitted with a Camlab epoxy tough single
200 junction combination pH electrode. Desorption is calculated as the percentage of the test
201 substance which is desorbed related to the quantity of substance previously adsorbed at
202 desorption equilibrium taking into account the incomplete removal of the supernatant after the
203 sorption experiment (approximately 3 mL) [16]. The apparent desorption coefficient (K_{des}) is
204 the ratio between the mass of the API on the soil and the mass concentration of the desorbed
205 API in the aqueous solution once desorption equilibrium has been reached [16].

206 2.2.3 Soil solution ratio

207 The OECD 106 guideline sets out several parameters when selecting the soil : solution
208 ratio including using at least 1 g of soil, achieving preferably >50 % sorption of the test
209 substance to the soil and the ratio can be as high as 1:1 (low K_d) or as low as 1:100 (high K_d)
210 depending on the estimated K_d of the API using molecular modelling software and an available
211 soil:solution ratio matrix (Figure S2) [16]. As an example a for the naproxen test a 1:5 ratio (6
212 g soil to 30 mL 10 mM $CaCl_2$) was chosen as the best balance between generating significant
213 sorption (predicted to be somewhere between 20% and 80% sorption) but still ensuring the
214 amount of naproxen left in solution was sufficiently high to accurately determine. The results
215 showed that the losses from solution after shaking for four hours would give quantifiable
216 results.

218 3 Results and Discussion

219 3.1 Liquid chromatography – mass spectrometry method development

220 A key aim was to develop an optimised LC-MS method for the simultaneous
221 quantification of the four APIs used in the study. This included assessing HPLC columns,

mobile phase composition, gradient elution, ionisation mode (positive or negative) and scanning method.

Three C18 HPLC columns were examined, including a Kinetex 2.6 μm EVO C18 100 Å (2.1 x 100 mm), a Waters XBridge BEH C18 2.5 μm (2.1 x 50 mm) Column XP and a Waters XSelect CSH C18 2.5 μm (2.1 x 50 mm); all columns had a pre-column filter fitted (HiChrom 0.5 μm). The Waters columns were selected as they had lower back pressures and produced better peak shapes, relative to the Kinetex column. The aqueous mobile phases considered were 0.1 % (v/v) ammonia and 0.1 % (v/v) formic acid, both in HPW. The use of 0.1 % (v/v) ammonia improved analyte signal response but carry over was detected, particularly for ofloxacin and propranolol. As a result, 0.1 % (v/v) formic acid was chosen as the aqueous phase (eluent C). Methanol (100 %) used as the organic mobile phase (eluent D). The high sensitivity of the mass spectrometer facilitated an injection volume of 5 μL . This reduction in injection volume helped reduce carryover, as did an injector needle wash before and after each injection using 1:1 (v/v) HPW : methanol. The gradient elution programme ran from 95:5 to 0:100 C:D over 6 minutes, then held for 2 minutes and returned to the starting ratio for 2 minutes. The retention times showed that the four APIs were clearly resolved from each other initially by testing individually and then finally as a mixture (Figure 1) allowing the APIs to be spiked into soil incubations as a mixture, and reducing both sample processing and analysis time.

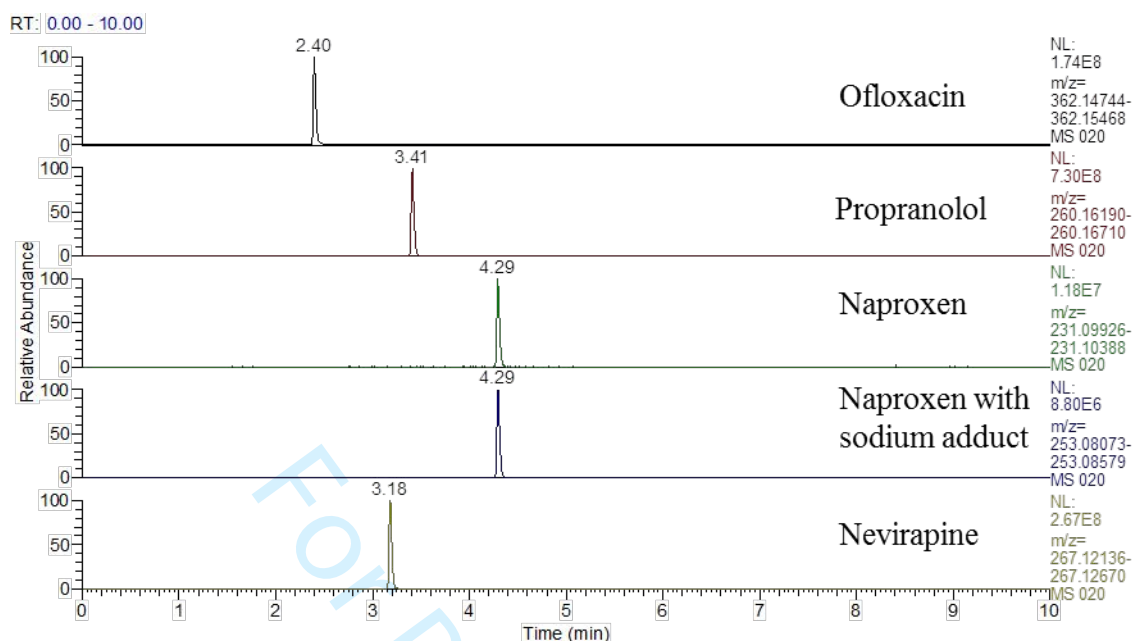


Figure 1. Example total ion chromatogram of the APIs studied (100 µg L⁻¹ in HPW), using a Waters XBridge BEH C18 2.5 µm 2.1x50 mm Column XP.

The eluent stream was diverted to waste from 0.1 – 1.5 minutes of the analytical run before entering the mass spectrometer in order to reduce contamination of the ionization source with salts from the sample matrix. All mass spectrometric analyses were performed in the positive ion mode. Parallel reaction monitoring (collision energy 30 eV) was used initially to target the specific ions and enable confident API identification with respect to background signals. Once it was apparent that the matrices of the soil filtrates did not interfere with the linearity of the calibration, full scan mode (m/z 100-1000) was used for all analyses.

Prior to all analysis, the MS response was assessed using a test mix (n-butylamine, caffeine, MRFA, and Ultramark 1621, Pierce LTQ Velos ESI, Thermo Fisher Scientific, UK). Methanol (HiPerSolv Chromanorm, VWR, UK) and HPW containing 0.1 % (v/v) formic acid (Optima Thermo Fisher Scientific, UK) were added and purged through the system before samples were analysed.

Due to problems with degradation or loss of ofloxacin to the glass autosampler vials, a batch of standards was made up in advance and frozen in 1 mL aliquots and defrosted alongside samples in extended periods of analysis longer than 48 hours.

3.1.1 Data analysis

All samples were analysed at least in duplicate. The raw data were analysed using Xcalibur software (Thermo Scientific, UK). Peaks were detected and smoothed using ICIS Peak Integration (Figure S1). The APIs were identified by retention time and m/z values measured to 5 decimal places. Sodium is a common contaminant in LC-MS, with main sources being glassware and mobile phases used. As naproxen forms a sodiated adduct, the peak areas of the parent ion and the sodiated adduct were combined. It is this level of scrutiny that is required to ensure the quality of data obtained via OECD tests on APIs.

Calibration standards included at least six separate concentrations and spanned at least two orders of magnitude (Table S1 in the electronic supporting information). Because of the wide range in the concentrations of the standards and with the loss of homoscedasticity, regression lines were weighted, using the Xcalibur software, to adjust the best fit line by a factor related to an inverse $1/x$ function of the concentration [38]. This served to reduce the relative error (variance) of each standard calibration measurement and made the relationship more relevant (reduced bias) at the lower end of the concentration range [39]. All API calibrations were linear and positive, with R^2 values > 0.99 (Table S2).

3.1.2 Analytical figures of merit

Precision

The precision of the analytical measurements was calculated to quantify random error over the length of an analytical run. To achieve this, the relative standard deviation (RSD) was

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calculated at three concentrations of API in HPW using at least 7 repeat measurements at each concentration (Table S3) [40].

With the exception of ofloxacin, the APIs showed good precision according to the EU decision 2002/657/EC as each RSD was < 20 % (Table S4) [40]. This decision specifies common criteria for the interpretation of analytical results to ensure that sample data are comparable between laboratories. Ofloxacin showed > 20 % RSD at the lower and middle concentrations. Ofloxacin is known to sorb to glass vials, and this is more pronounced at lower concentrations [41]. As the samples used for the precision experiment were in the glass vials of the autosampler for different lengths of time, loss of ofloxacin to glass vials would likely have varied.

Limits of detection

Instrument LOD was calculated using at least 5 low calibration concentrations to ensure that the data presented were both robust and analytically relevant (Table 3). LOD was derived using the standard deviation of the regression line intercept for each calibration, rather than the more usual standard deviation of replicate calibration blank measurements (Equation S1; [42].

Table 3 LODs for APIs in the different matrices used in this study ($\mu\text{g L}^{-1}$)

	HPW	Loam	Sandy loam
Ofloxacin	0.36	0.16	0.14
Propranolol	0.39	1.95	0.51
Naproxen	0.14	0.82	0.99
Nevirapine	0.15	0.28	0.25

As can be seen from Table 4, LODs below $\mu\text{g L}^{-1}$ were achieved, which is in agreement with reported values [43]. Instrument LODs for all APIs were adequate for this study.

3.1.3 Matrix effects

Initially, matrix effects on the measurement of APIs were quantified by spiking filtered soil suspensions in 10 mM CaCl_2 and comparing the results to HPW. The soil suspensions were prepared by shaking overnight, centrifuging at 4000 rpm for 15 min, followed by filtration through 0.7 μm GFF at different concentrations, and assessing the linearity and suppression of peak areas at each concentration compared to spiked HPW. A set of calibration solutions (in HPW) was prepared in parallel using the same API standard stock solutions, for comparison.

Matrix effects were observed but were not consistent for each API. Where an effect was identified, it was always quenching of the signal (Figure 2). Ofloxacin in sandy loam had the greatest matrix effect compared to HPW, with an 80 % decrease in the average peak area measured but linearity was not affected. These data indicate that calibration standard solutions should be matrix matched to the samples due to the signal quenching.

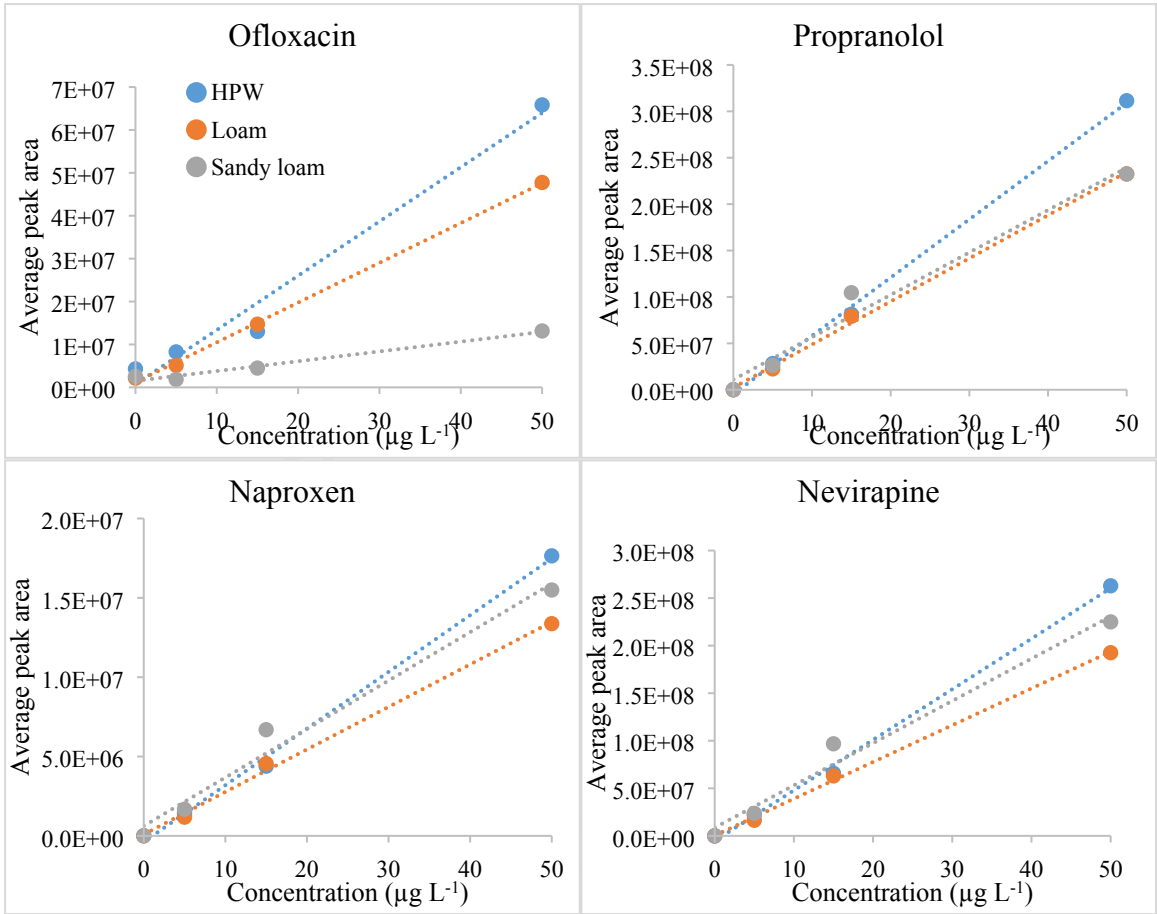


Figure 2 Soil filtrate matrix effects. Legend is shown in ofloxacin graph and consistent throughout this set of graphs. Error bars are present but not visible ($\bar{x} \pm \text{s.d.}$, $n=3$)

Samples were incubated with shaking for 120 h to check that changes in the soil matrix, such as desorption of organic carbon or other organic material, disaggregation of colloids etc, which might occur over the 120 h sorption experiment, affected API measurements. Separate soil suspensions (6 g soil 30 mL 10 mM CaCl₂) without added APIs were shaken for 24 h and 120 h (to match the longest incubation experiment) and these incubations finished at the same time. The shaken soil suspensions were centrifuged and filtered as before and APIs spiked into the filtrates in the concentration range 0.1 – 100 μg L⁻¹. The data showed that any temporal changes in matrix did not affect analyte response in the instrument (Figure 3), with the largest difference < 20 % (ofloxacin in loam soil).

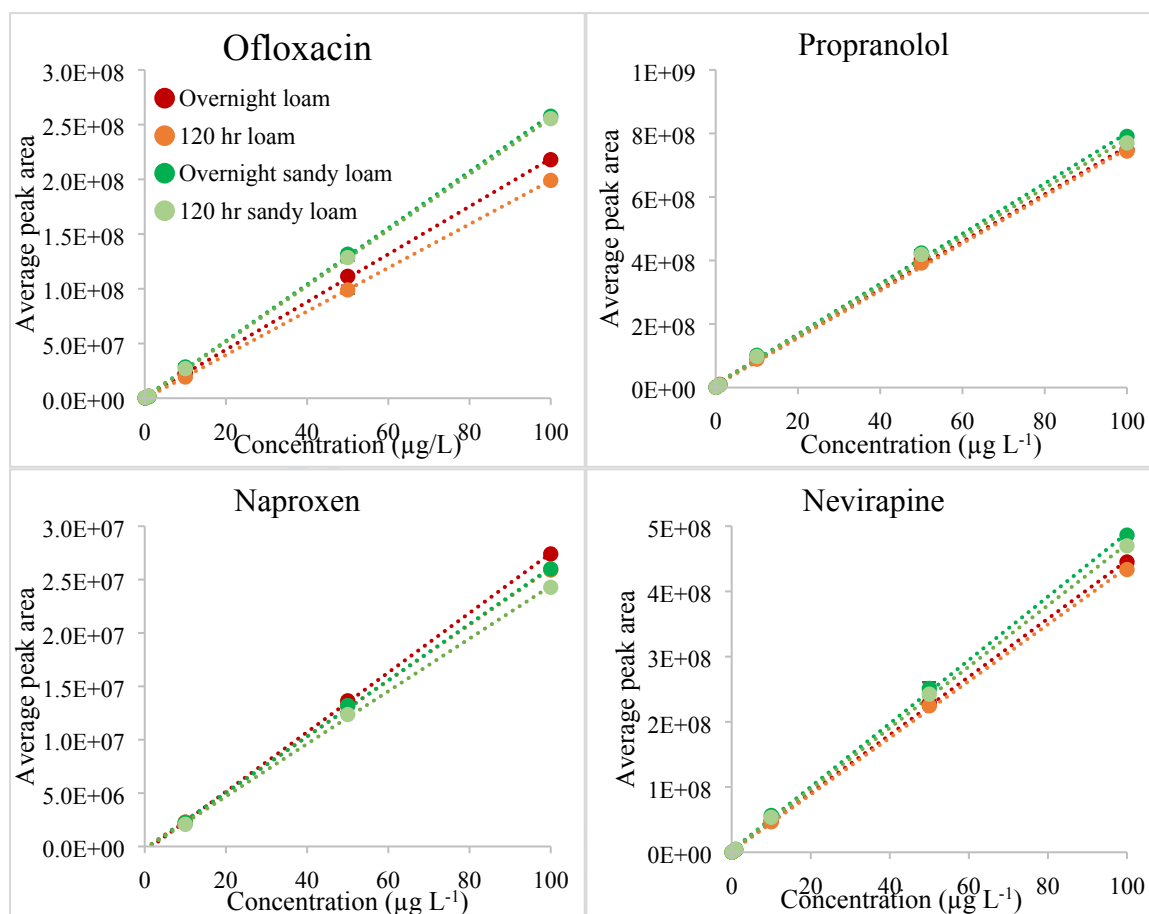


Figure 3 120 h soil matrix effects. Legend is shown in ofloxacin graph and is consistent throughout this set of graphs. Error bars are present but not visible ($\bar{x} \pm \text{s.d.}$ n=3)

For ease of analysis, all calibration standards were made up in soil filtrates representing the 24 h incubation. While there was some variation in specific APIs, the data were generally consistent across the time period and a 120 h calibration added little analytical value while increasing both workload and costs.

3.2 API loss to laboratory ware

3.2.1 Plastic ware

The results show that these APIs are unlikely to be lost to the centrifuge tubes as all recoveries were $> 90\%$ (Table 4). As a result, all sorption experiments were undertaken in 50 mL polypropylene centrifuge tubes.

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Table 4 Spike concentrations of APIs and % recoveries of APIs from 50 mL polypropylene centrifuge tubes ($\bar{x} \pm \text{RSD } n = 3$)

	Spike concentration ($\mu\text{g L}^{-1}$)	Recovery (%)
Ofloxacin	500	98.6 ± 4.2
Propranolol	240	101 ± 1
Naproxen	58	91.7 ± 17.1
Nevirapine	50	115 ± 4

3.2.2 Filter membranes

Glass fibre filters showed the lowest sorption of APIs and hence highest recovery for all three APIs, relative to the organic material-based membranes (Figure 4). Nevirapine also showed quantitative recovery and was only assessed for the glass fibre filters. The OCED 106 guideline requires filters with a pore size of 0.2 μm if centrifugation does not reach this level. These are not available in glass fibre membranes causing problems for compliance with the guideline but due to the better recoveries for these four APIs these filters were chosen for the experiments.

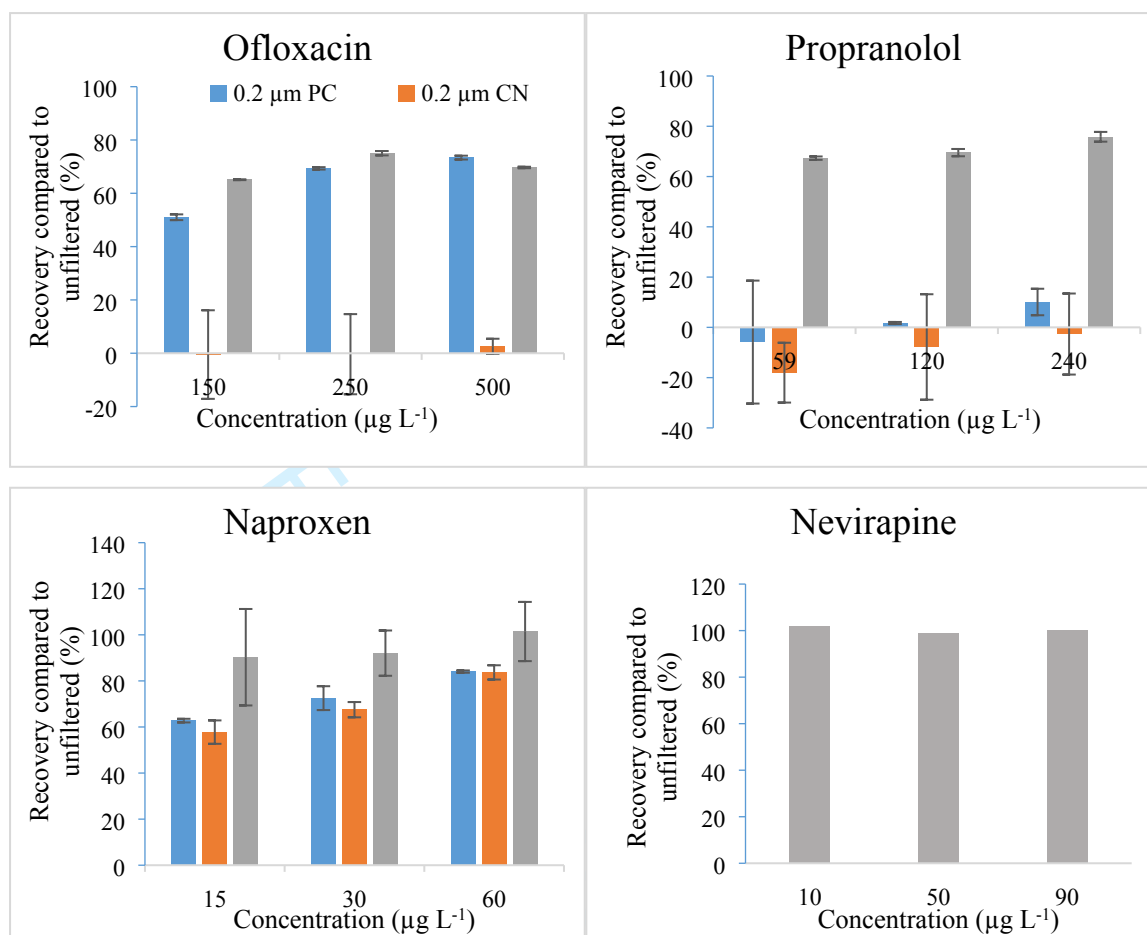


Figure 4 Recovery of APIs following filtration (PC – polycarbonate, CN – cellulose nitrate, GF – glass fibre) ($\bar{x} \pm \text{RSD}$, $n=3$; except nevirapine, $n=2$ no RSD). Legend is shown in ofloxacin graph.

3.3 API and sample storage

The aim of this experiment was to test that the samples could be successfully stored at $-20\text{ }^{\circ}\text{C}$ without effecting the concentration of API in the samples. This was necessary as the HPLC-HRAM-MS was not always immediately available so adequate sample preservation was required until it could be used.

Concentrations of ofloxacin were significantly lower in the calibration solutions that had been stored frozen (ANOVA, $p \leq 0.05$). The range was 15-34 %, with the largest observed difference at $10\text{ }\mu\text{g L}^{-1}$ and the lowest at $100\text{ }\mu\text{g L}^{-1}$ (Figure S3). The other APIs were less influenced by this storage method. If samples were stored at $-20\text{ }^{\circ}\text{C}$, a set of standard solutions was prepared alongside samples and stored in the same manner for consistency. Fresh API

calibration solutions were used to test the sensitivity and repeatability precision of the instrument.

3.4 Soil solution changes

The pH and DOC of the soil solutions are key variables in controlling the partitioning of organic chemicals and are rarely determined throughout the course of an equilibrium experiment. Data provided here shows that there was variation in the properties of the matrix after a period of shaking. pH of the loam soils started at 7.2 before reducing by 0.5 pH units to 6.7 over the 120 hours (Figure 5A). In contrast the sandy loam soil started at 6.1 then increased by 0.5 pH units to 6.6. The DOC results showed that the loam soil solution had a greater concentration of DOC throughout the experiments starting at 30 mg L⁻¹ before increasing over the shaking period to 44 mg L⁻¹ (Figure 5B). Sandy loam DOC started at 3.6 mg L⁻¹ before increasing to 9.4 mg L⁻¹. These are important considerations because such factors are rarely considered in soil testing experiments. Any increase in DOC in solution over the duration of the test is likely to lead to stabilisation of an API in solution, therefore reducing the calculated K_{oc} and may also influence the attainment of equilibrium. The pH of soil suspensions influences the net charge on ionisable APIs and they will be fully ionised (> 99 %) when the pH is at ± 2 pH units from their pK_a [18]. Variations in soil suspension pH over the length of the sorption experiment could therefore have an impact on the ionisation state of the APIs in question. Variation in pH could cause more significant changes to sorption behaviour in other compounds with a pK_a around 6 under these conditions. Soil pH will also influence the pH-dependent charge on the organic matter, clay minerals and metal sesquioxide components of the soil, which may influence API sorption [44].

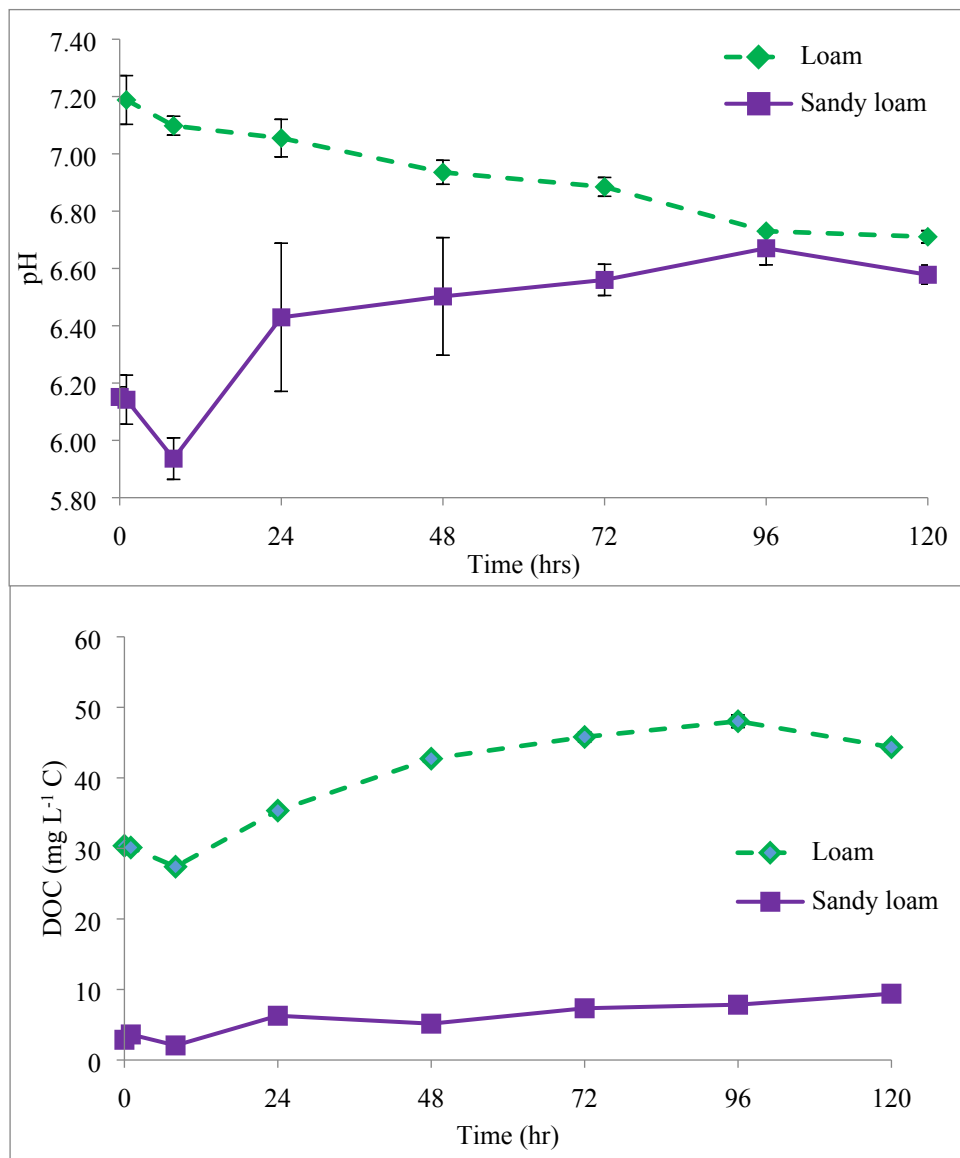


Figure 5 A - pH of soil solutions over 120 hours shaking ($\bar{x} \pm \text{S.D.}$, $n = 3$). B - DOC in soil solutions over 120 hours shaking ($\bar{x} \pm \text{S.D.}$, $n = 3$)

4. Conclusions

The ability to measure and understand the behaviour of APIs in soil is essential for any rigorous risk assessment associated with wastewater reuse or biosolid recycling to land, particularly in LLMIC where water and fertiliser availability are limited. An existing OECD testing methodology was fully evaluated for key variables which could impact on the quality of the data generated. Four APIs with wide ranging functionality, charge, molecular weight and sorption characteristics were used and data for naproxen provided as a case study to demonstrate the outcomes of applying such a technique to APIs. A full assessment of the sample preparation, sample stability, analytical method choice and performance and potential matrix interferences have been completed to enable practitioners and scientists to replicate the methodology for these and other APIs.

The data from the study shows that the choice of materials used for testing is crucial, with any filtration requiring glass fibre membranes, which restricts the minimum practical pore size to 0.7 μm . Adsorption experiments, such as the OECD 106 guideline, can be performed in polypropylene plastic centrifuge tubes with careful measurement of recoveries, which were 100% +/- 15% for the 4 APIs tested.

HPLC-HRAM-MS was shown to deliver LODs below 1 $\mu\text{g L}^{-1}$ for most matrices, with excellent precision and clear resolution of the eluting APIs. Ofloxacin was the most challenging API to measure, due to its high affinity for surface adsorption leading to instrument carry-over, and through loss to apparatus under certain conditions. This illustrated the importance of careful analytical quality control to ensure that useable data are obtained in the subsequent fate studies. Sample storage was shown to be viable and there were no long- term (5 day) impacts of matrix changes on analytical performance. Some matrix effects were observed for all 4 of the APIs tested, with quenching of the signal in all cases, particularly for ofloxacin,

consequently it is recommended that all analyses are undertaken alongside matrix-matched calibration.

Important aspects such as changes in solution pH and DOC over the course of the partitioning experiments should show how important it is to monitor these critical variables which impact on the partitioning and fate of chemicals in the soil/water environment.

Overall, the data presented in this study shows the importance of systematic method development prior to undertaking soil risk assessment studies for APIs.

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Developing the OECD 106 fate testing protocol for active pharmaceuticals in soil

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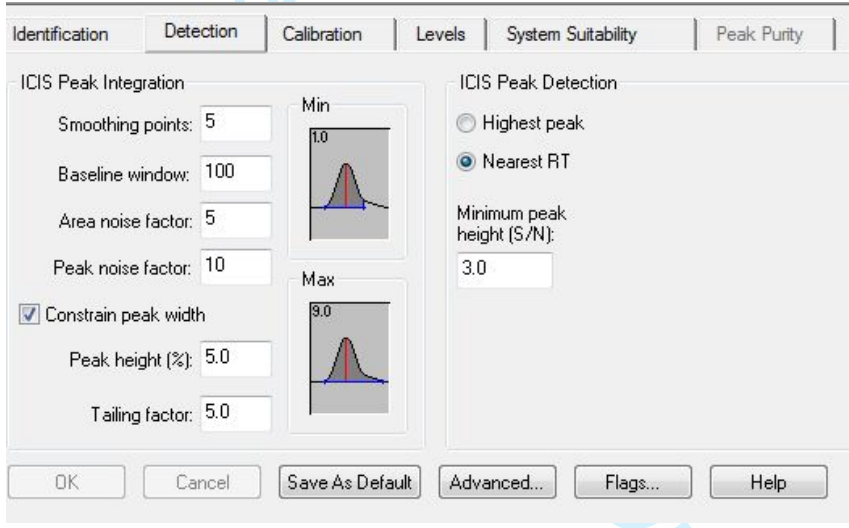


Figure S1. Peak detection and processing method from Thermo Scientific Xcalibur software

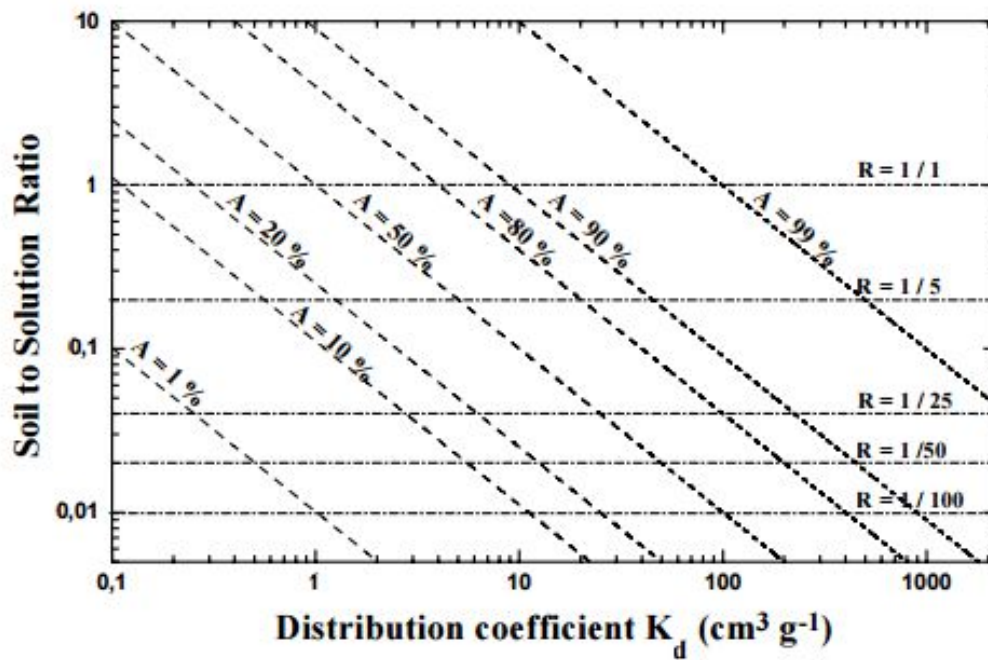


Figure S2 Relationship between soil : solution ratios and K_d at various percentages of adsorbed test substance; A = adsorption, R = soil : solution ratio [1].

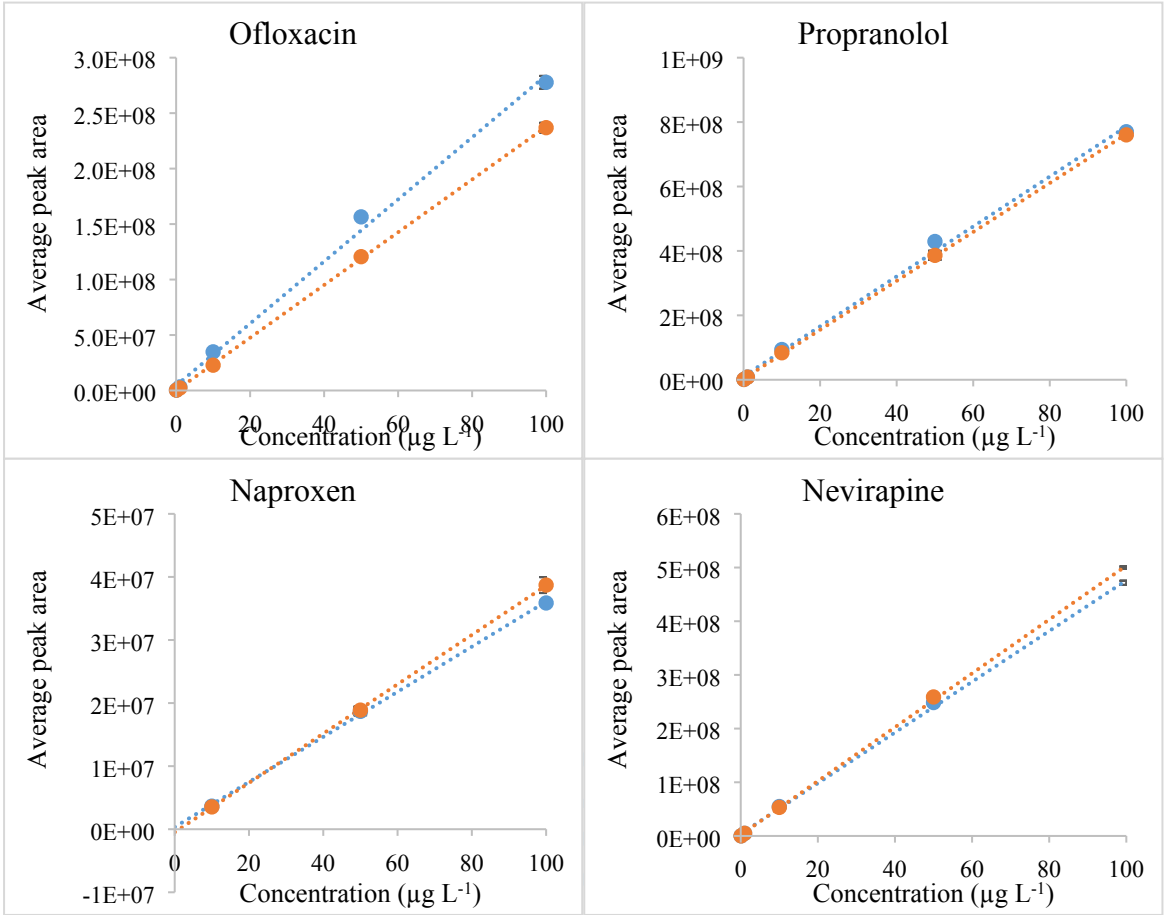


Figure S3. Effects of storage on API calibrations. ● = freshly prepared, ● = frozen. Error bars were calculated but are not visible in most cases ($\bar{x} \pm \text{s.d.}$ n=3)

Table S1. Calibration concentrations for each API ($\mu\text{g L}^{-1}$)

Cal label	Ofloxacin	Propranolol	Naproxen	Nevirapine
C1	0.1	0.8	0.3	0.4
C2	0.625	5	1.875	2.5
C3	1.25	10	3.75	5
C4	5	40	15	20
C5	10	80	30	40
C6	20	160	60	80
C7	25	200	75	100

Table S2. Calibration data in all matrices

API	Matrix	Equation	R ²
Ofloxacin	HPW	$y = 2.48 \times 10^6 \chi + 802007$	0.9945
	Loam	$y = 2.21 \times 10^6 \chi + 105035$	0.9925
	Sandy loam	$y = 2.75 \times 10^6 \chi + 18175$	0.9997
	SWW	$y = 0.93 \times 10^6 \chi + 930210$	0.9689
	SWW + loam	$y = 0.76 \times 10^6 \chi + 25681$	0.9917
Propranolol	HPW	$y = 7.87 \times 10^6 \chi + 3.66 \times 10^6$	0.9925
	Loam	$y = 7.51 \times 10^6 \chi + 4.87 \times 10^6$	0.9950
	Sandy loam	$y = 7.51 \times 10^6 \chi + 2.08 \times 10^7$	0.9934
	SWW	$y = 6.20 \times 10^6 \chi - 5552 \chi^2 + 1.27 \times 10^6$	0.9994
	SWW + loam	$y = 8.11 \times 10^6 \chi - 7194 \chi^2 + 2.37 \times 10^6$	0.9981
	SWW + sandy loam	$y = 6.34 \times 10^6 \chi - 6409 \chi^2 + 0.8 \times 10^6$	0.9995
Naproxen	HPW	$y = 0.29 \times 10^6 \chi - 464824$	0.9981
	Loam	$y = 0.27 \times 10^6 \chi - 438441$	0.9986
	Sandy loam	$y = 0.25 \times 10^6 \chi - 453599$	0.9984
	SWW	$y = 0.42 \times 10^6 \chi - 74745$	0.9994
	SWW + loam	$y = 0.46 \times 10^6 \chi - 51935$	0.9992
	SWW + sandy loam	$y = 0.41 \times 10^6 \chi - 74212$	0.9997
Nevirapine	HPW	$y = 5.13 \times 10^6 \chi - 199410$	0.9993
	Loam	$y = 0.26 \times 10^6 \chi - 438441$	0.9986
	Sandy loam	$y = 5.24 \times 10^6 \chi + 445596$	0.9986
	SWW	$y = 4.45 \times 10^6 \chi + 69675$	0.9995
	SWW + loam	$y = 6.82 \times 10^6 \chi + 1.93 \times 10^6$	0.9955
	SWW + sandy loam	$y = 4.84 \times 10^6 \chi + 345717$	0.9986

Table S3 Concentration of APIs used to calculate precision

	Low (µg L ⁻¹)	Middle (µg L ⁻¹)	High (µg L ⁻¹)
Ofloxacin	1.25	10	25
Propranolol	10	80	200
Naproxen	7.5	60	150
Nevirapine	7.5	60	150

Table S4. Relative standard deviation of HPLC-HRAM-MS method for APIs at low, middle and high concentrations

	Low (%)	Middle (%)	High (%)
Ofloxacin	94	23	17
Propranolol	1	3	6
Naproxen	4	4	4
Nevirapine	3	4	4

Equation S1 HPLC-HRAM-MS Limit of detection [2]

$$LOD = \frac{(3.3 \sigma)}{S}$$

Where σ = standard deviation of the y-intercepts of the regression line and S = slope of the calibration curve.

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