

2019-07-01

Jurassic shift from abiotic to biotic control on marine ecological success

Eichenseer, K

<http://hdl.handle.net/10026.1/14472>

10.1038/s41561-019-0392-9

Nature Geoscience

Nature Research

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Please cite only the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author.

S1. ASI uncertainty and components

Mg/Ca data. A variety of proxies have been developed to reconstruct the original Mg/Ca ratio of past seawater. These include abiotic proxies, such as the composition of fluid inclusions in halite⁷²⁻⁷⁵ or the Mg-concentrations in carbonate veins from Mid-Ocean ridge flanks⁷⁶, and biotic proxies based on the calcareous skeletons of echinoderms⁷⁷⁻⁷⁹, rudists⁸⁰, and foraminifera^{81,82}. While together these proxies reflect the general pattern of aragonite-calcite sea oscillations first described by Sandberg (1983)¹⁰, there is substantial spread between different proxies and even between different measurements of the same proxy (e.g. for fluid inclusion data; see Fig. S1a). As each proxy has different underlying assumptions, it is difficult to directly compare different proxies. Furthermore, the time-coverage of existing proxy data is very scattered with a noticeable paucity of data in the lower-mid Palaeozoic (Fig. S1a).

Among existing proxies, echinoderm skeletal remains have perhaps the highest potential to eventually provide a densely sampled continuous Phanerozoic record of a single proxy. However, the suitability of echinoderms skeletal remains for reconstructing seawater Mg/Ca ratios has been questioned as magnesium concentrations significantly vary within individual skeletons⁸³, are dependent on the food source⁸⁴, and do not always mirror expected secular Mg/Ca trends even in very well-preserved specimens⁸⁵.

In addition to proxy records, various modelling approaches have been developed to reconstruct the Mg/Ca ratio of past seawater^{16,22,86-88}, (see Fig. S1a). All models are to some degree informed by proxy data, with some using in-model proxy calibrations as part of their approach (e.g. Farkaš et al. 2007; Arvidson et al. 2013). In the latter approach parts of the time series lacking proxy data are strongly biased by the closest proxy data, which is a particular problem for the Palaeozoic. To calculate the CaSO₄ flux in the model by Arvidson et al. (2013), for example, the fluid inclusion data by Lowenstein et al. (2003) are used. With only five Palaeozoic data points (one each for the Cambrian, Silurian, and Devonian, and two for the Permian). The resulting Phanerozoic Mg/Ca curve has a smooth sinusoidal shape that does not track stage-scale oscillations.

The Farkaš et al. (2007) model uses a combination of fluid inclusion data^{72,74}, echinoderms⁷⁹, and rudists⁸⁰. Given the above mentioned uncertainties about echinoderms as Mg/Ca proxy, and the overall challenge of directly comparing different proxies, we did not consider this model any further.

The models of Berner 2004 and of Demicco et al. 2005 do not rely on proxy data as an internal model component, but compare model outputs with proxy data (in the Berner (2004) model, starting conditions were adjusted based on how well the model output fitted the Lowenstein et al. (2001) and Horita et al. (2002) fluid inclusion data). Both are similar in their relative timing of changes of the Mg/Ca ratio as they both rely on the same seafloor production curve⁸⁹. However, Berner's (2004) forward modelling underestimates modern seawater Mg/Ca ratio. By contrast, Demicco et al. (2005) used backwards modelling, starting with the major ion composition of modern seawater and incorporates reactions along the cooler mid-ocean ridge flanks.

We thus use the Mg/Ca ratio obtained from model A in Demicco *et al.* (2005) for our estimation of ASI. For comparison we show below how similar the ASI is when based on the Demicco et al. (2005) and the Berner (2004) models (Fig. S1b), that that linear regression and CCM analysis of SCOR_{ara} and ASI based on both these models comes to very similar results (Fig. S2, S4b).

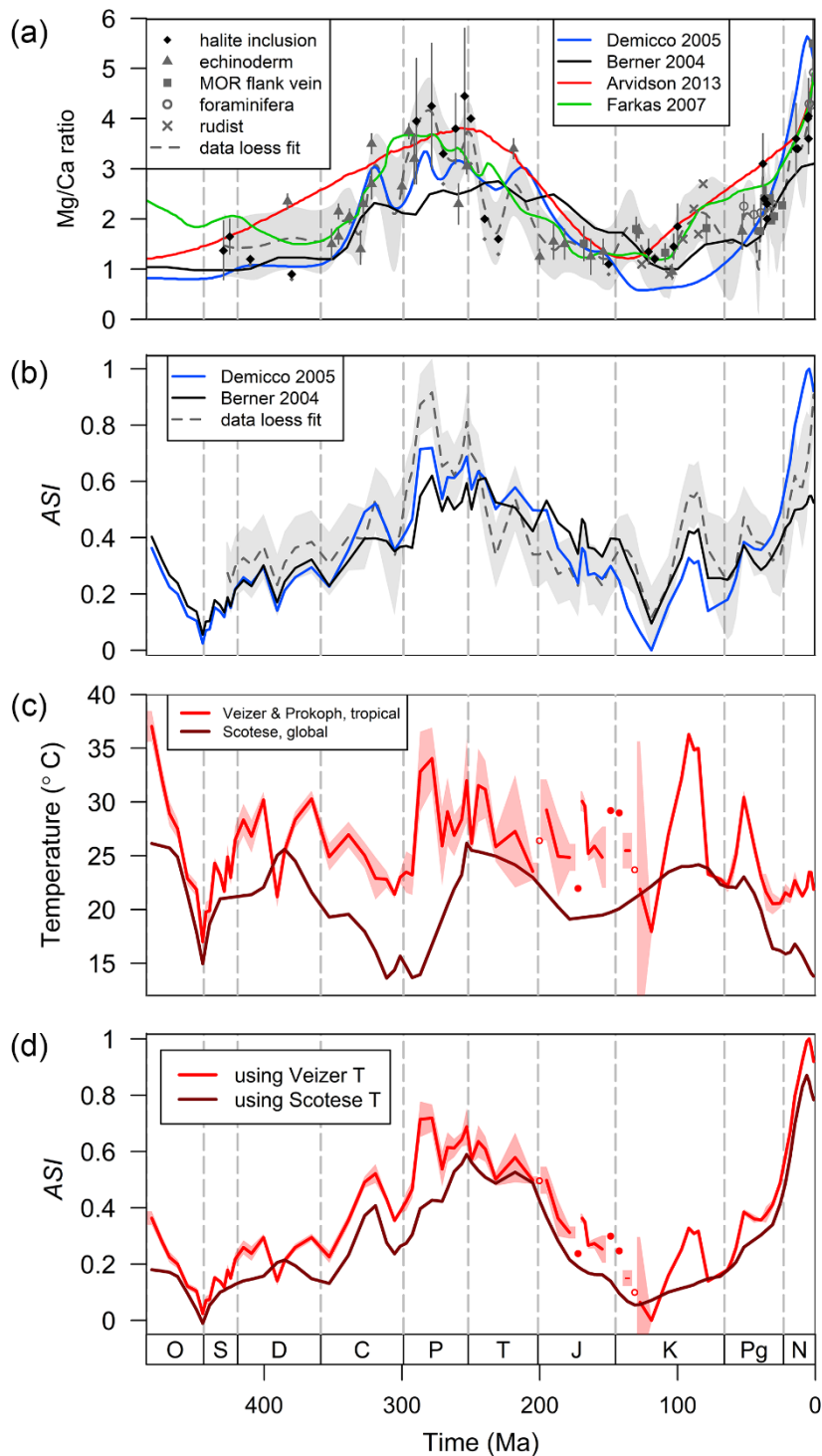


Figure S1: Alternative temperatures and Mg/Ca ratios

(a) Mg/Ca ratios from models of Demicco *et al.* (blue), Berner (black), Arvidson *et al.* (red) and Farkas *et al.* (green) and proxy data from halite inclusions (diamond)^{72-75,90-92}, echinoderm skeletons^{77,78} (triangle), mid ocean ridge flank veins⁷⁶ (square), foraminifera tests^{81,82} (circle), and rudist shells⁸⁰ (cross). Error bars denote uncertainties where given, the tiny dots are fluid inclusion Mg/Ca ratios with an alternative calcium and sulphate ion product assumption. The dashed line denotes a locally estimated scatterplot smoothing (LOESS) regression with span = 0.1, based on all proxy data, with one standard error around the regression estimate (grey envelope).

(b) ASI calculated using the temperature data of Veizer and Prokoph and the Mg/Ca ratios from models of Demicco *et al.* (blue) and Berner (black). ASI based on the Mg/Ca LOESS regression in (b) is shown as dotted grey line, with an error envelope based on one standard error around the Mg/Ca estimate.

(c) Mean tropical shallow water temperatures from $\delta^{18}\text{O}$ measurements from a compilation of Veizer and Prokoph (2015) and a few additional data^{23,93} (red) with 2 standard error envelopes (light red), and global average temperatures from Scotese (2016, brown)⁹⁴.

Stages with only one measurement are drawn as solid dots, stages without observations have been averaged from the neighbouring stages and are shown as circles.

(d) ASI calculated using the Mg/Ca model of Demicco *et al.* and the temperatures of Scotese (brown) and Veizer and Prokoph (blue), respectively. The light blue area is a 2 standard error envelope based on the temperature data of Veizer and Prokoph (c). Stages with only one measurement are drawn as solid dots. Stages without observations (circles) have been averaged from the neighbouring stages.

Temperature data. The tropical sea surface temperatures we are using are based on over 15,000 $\delta^{18}\text{O}$ measurements compiled by Veizer and Prokoph (2015)²³ and a few more measurements from a more recent publication⁹³, resulting in an almost continuous, stage-level temperature record (Fig. S1d). To test how sensitive our analysis is to potential temperature errors, we compare this data set with an alternative, global temperature curve generated by Scotese (2016)⁹⁴. Although some of the long-term trends, e.g. a temperature drop corresponding to the Late Palaeozoic ice age feature in both curves, the Scotese (2016) curve shows a lot less short term variation, which results in a smoother ASI curve (Fig. S1c).

ASI sensitivity test. To test how robust the relationship between ASI and SCOR_{ara} is to differences in ASI resulting from different models or data, we calculated an ASI from each of the two temperature curves from Fig. S1a and two of the Mg/Ca models from Fig. S1b. We conducted linear regressions in expanding windows of SCOR_{ara} against the four resulting ASI combinations. The regression patterns with the alternative ASI combinations (Fig. S2b-d) are encouragingly similar to the patterns produced by our initially proposed ASI (Fig. S2a). The Scotese (2016) temperatures barely change the correlation in the Palaeozoic, but yield a stronger correlation in the early Mesozoic. The Berner model results in a weaker correlation in the early Mesozoic when paired with the Veizer and Prokoph (2015) temperatures, but the loss of positive correlation after the early Mesozoic can still be seen. The negative correlation in the later Mesozoic and early Cenozoic with this specific model is likely spurious.

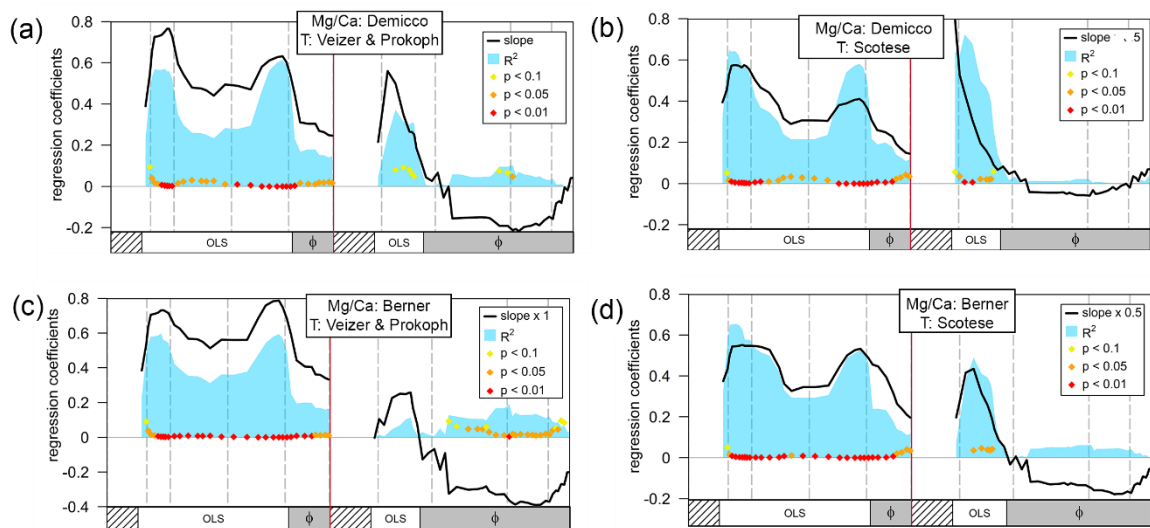


Figure S2: Regression in expanding windows with alternative ASI

Linear models of SCOR_{ara} against ASI in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately. Slope (black) and R^2 (blue) of each model are drawn at the last included stage of the respective window. p -values are only shown when they are < 0.1 . All Palaeozoic models start at the first Ordovician stage (Tremadocian), and all Mesozoic – Cenozoic models start with the Early Triassic (Induan). The box at the bottom shows whether the models were generated using ordinary least squares (OLS), or generalised least squares with autoregressive errors (ϕ). No models were fitted for intervals with five or less stages (hatched pattern).

(a) ASI was calculated using the Mg/Ca ratio of Demicco *et al.* and the temperatures from Veizer and Prokoph as in Fig. 2b.

(b) ASI was calculated using the Mg/Ca ratio of Demicco *et al.* and the temperatures from Scotese.

(c) ASI was calculated using the Mg/Ca ratio of Berner and the temperatures from Veizer and Prokoph.

(d) ASI was calculated using the Mg/Ca ratio of Berner and the temperatures from Scotese.

Mg/Ca and Temperature separately

To evaluate the relative importance of the two components of the ASI model, Mg/Ca ratio and temperature, we assess their independent effect on $SCOR_{ara}$. The Mg/Ca model shows little variation in the Ordovician – Devonian, and linear regressions in this time window behave erratic as a consequence (Fig. 3b). With increasing Mg/Ca variability in the late Palaeozoic and the early Mesozoic, the correlation patterns inferred with ASI extending to these intervals are similar to the patterns with just Mg/Ca ratio. Temperature alone drives the linear correlation in the earlier part of the record, but becomes dominated by Mg/Ca ratio from the Carboniferous onwards. CCM results are not limited to linear dynamics and thus may give a better idea of which model component exerts greater control on $SCOR_{ara}$. We find that Mg/Ca ratio shows stronger dynamical coupling with $SCOR_{ara}$ than temperature does (Fig. S4b), although the analysis is only significant at the 95th confidence level when temperature and Mg/Ca ratio are combined to the ASI forcing model which best reflects current understanding of abiotic influences on calcification.

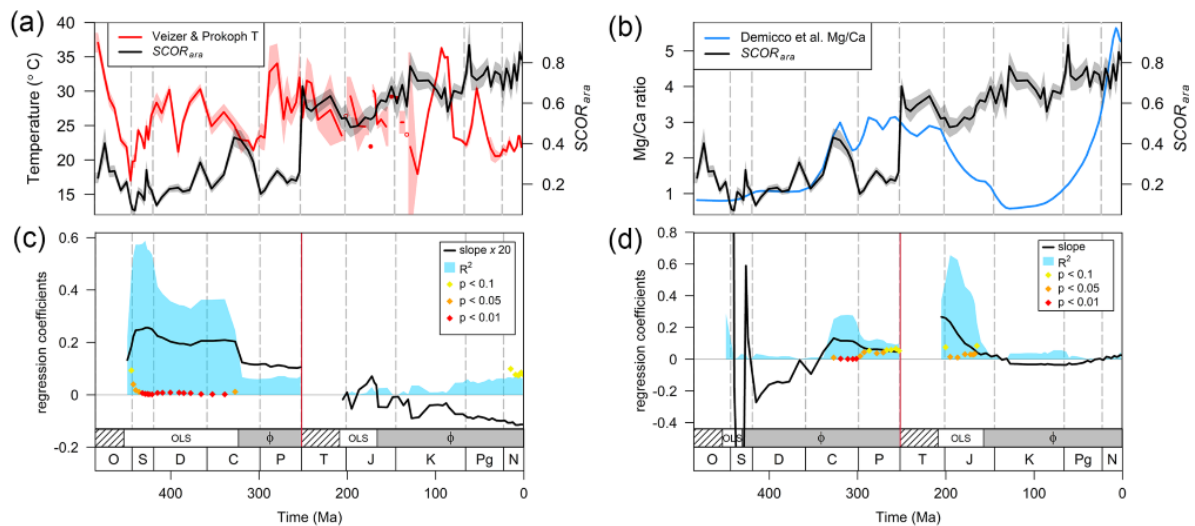


Figure S3: Temperature and Mg/Ca separately with $SCOR_{ara}$

- (a) Mean tropical shallow water temperatures from Veizer and Prokoph as in Fig. S1c (red) and $SCOR_{ara}$ (black).
- (b) Modelled Mg/Ca ratio from Demicco *et al.* as in Fig. S1a (blue) and $SCOR_{ara}$ (black).
- (c) Linear models of $SCOR_{ara}$ against temperature (a) in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately.
- (d) Linear models of $SCOR_{ara}$ against Mg/Ca ratio (b) in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately.

S2. Convergent cross mapping (CCM)

In the following, we discuss the application of CCM in the context of our study, starting with a brief outline of the algorithm (see Sugihara *et al.*, 2012 for details), then give a heuristic theoretical basis for the expanding window analysis.

CCM algorithm

The CCM algorithm works as follows. First, reconstruct the state space for the driver system by a time-delay embedding of the driver time series. Then, pick a point (one point = one sample) on the reconstructed Mg/Ca orbit and keep track of its time index in terms of the original time series. Say the point P_i with time index t_i is picked, with the corresponding scalar point in the driver time series being $D(t_i)$. Next, locate the $dim + 1$ nearest neighbours of P_i in the reconstructed phase space (forming a dim -

dimensional simplex enclosing a volume around the point), say their time indices are $t_{k_1}, t_{k_2}, \dots, t_{k_{(\text{dim}+1)}}$. Then, make a prediction of $D(t_i)$ by computing a weighted average of $D(t_{k_1}), D(t_{k_2}), \dots, D(t_{k_{(\text{dim}+1)}})$, where the weights are controlled by the distances between P_i and $P(t_{k_1}), P(t_{k_2}), \dots, P(t_{k_{(\text{dim}+1)}})$. The main idea of CCM is that if the underlying variables are dynamically coupled, close neighbours in the response system should map to close neighbours in the driver system.

Convergence criterion

CCM uses convergence of prediction skill as the number of points increases to distinguish causation from correlation²⁷. For each time series pair, the library used for the analysis, i.e. those data points from the response time series which are used to predict the driver time series, is gradually increased up to the total length of the time series. To determine a minimum time window for which cross mapping is applicable, we consider analyses convergent if the median CCM prediction skill over 500 samples is higher at large library sizes than at lower library sizes. Convergence of an analysis is assessed using a Wilcoxon rank-sum test: if the test for a given time window fails to reject the null at the 0.01 level, we consider the analysis non-convergent. For the shortest time windows, no robust test could be performed, and these analyses are considered non-convergent.

Robustness to data uncertainty

To explore the robustness of the results when incorporating the uncertainty in the data, we use the uncertainty handling machinery in the UncertainData.jl (<https://github.com/kahaaga/UncertainData.jl>) Julia package, in combination with the high-performance CrossMappings.jl package (<https://github.com/kahaaga/CrossMappings.jl>).

Theoretical basis for expanding window analysis

Due to, for example, secular changes in orbital configuration and time-varying mid-ocean ridge forcing, the Earth system is inherently nonergodic. To interpret the CCM results in a dynamical systems framework, we thus adapt a pullback attractor interpretation of the system, which we heuristically summarise in the following (we refer to Chekroun et al., 2011 for an in-depth introduction to this topic⁹⁵).

Namely, consider the Earth system as a nonautonomous dynamical system, i.e. $\mathbf{x}' = (t, \mathbf{x})$, where the governing equations f depend explicitly on time. Let (s, t) be its solution at time t , where \mathbf{x} is the initial state at time s (with $s \leq t$). In the autonomous case the system is time-translation invariant, i.e. $(s, t) = (t - s)\mathbf{x}$. However, in the nonautonomous case, when fixing a time t ("now"), the initial time s at which the system started becomes important, and the asymptotic behaviour occurs when $s \rightarrow -\infty$. Solutions obtained when fixing s and letting $t \rightarrow \infty$, and when fixing t and letting $s \rightarrow \infty$, may thus differ. In forced systems, this gives rise to the notion of a pullback attractor, which is a collection $\cup_{t \in \mathbb{R}} (t)$ of dynamically invariant objects (t) that each attracts some initial points starting in the asymptotic past. Provided the time window $t - s$ is sufficiently larger than the typical convergence time to the attractor, this defines a family of attractors defined by the "freezing time" t . In this interpretation, expanding the time window corresponds to fixing t in (s, t) at increasing t , yielding a set of attractors from which to cross map. Hence, if cross mapping convergence occurs for a certain time series length in the expanding window analysis, we may adopt the pullback attractor view for any time series length beyond that.

If time-dependent dynamical coupling existed between ASI and SCOR, this would be reflected in the pullback attractors. Thus, we would expect a change in the ability for SCOR to predict the putative ASI driver as the time window expands. This is precisely what we observe in our CCM analyses: after convergence is achieved for a sufficiently long time series, including more points does not alter convergence but changes the magnitude of the prediction skill (Fig. S4a).

Expanding window analysis on Phanerozoic ASI and SCOR_{ara}

In Figure 2c we only show the results of analyses for which convergence could be demonstrated, i.e. the windows spanning the Ordovician – late Middle Triassic (Ladinian) until the Ordovician – Late Pliocene (Piacenzian). No convergence was achieved in windows from the Ordovician – Silurian until the Ordovician – early Middle Triassic (Anisian), which we attribute to the low number of data points in these analyses. The lack of convergence in the longest window, the Ordovician – Pleistocene may be due to the weakening influence of ASI on SCOR_{ara} and is preceded by a decrease in CCM skill throughout the Cenozoic. Despite the decline of CCM skill in the Jurassic – Early Cretaceous, intervals extending to the mid-Cretaceous – Palaeogene show a significant coupling of ASI and SCOR_{ara} (Fig. 2c). The analysis in these time windows may be dominated by the stronger coupling in older time intervals, or signify a genuine, albeit weakened influence of ASI extending through to the Palaeogene. The overall declining cross mapping skill is found to be robust against the uncertainty in the stage-level SCOR_{ara} and ASI data (Fig. S4b), both when using the Demicco or Berner Mg/Ca model. Significant cross mapping in any time window only occurs between SCOR_{ara} and the combined ASI record, and not for SCOR_{ara} and Mg/Ca alone, which is expected if ASI is a better measure of the abiotic forcing of SCOR_{ara} than Mg/Ca alone.

Lastly, we note that although using the stage level data for CCM analyses pushes the limits of the applicability of the method, we do not apply interpolation to a regular grid and introduce more data points, because that would introduce spurious dynamical information which we have no way of verifying. Rather, we assume that the sequence of stage level SCOR_{ara} and ASI contains sufficient variability to assess the dynamical relationship between the variables on the time scales considered here. Despite the coarse-grained data, we find that the relationship between SCOR_{ara} and ASI is surprisingly robust to noise and the choice of Mg/Ca models and T records (Fig. S4b). The congruence between the linear correlation and cross mapping results supports that the covariation of SCOR_{ara} and ASI is not merely a statistical coincidence.

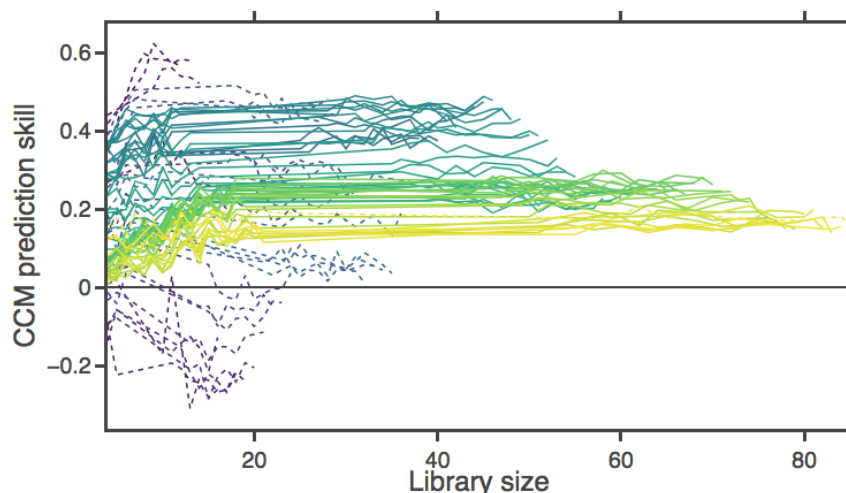


Figure S4a: CCM convergence plot for time windows of increasing length (gradient from blue to yellow), ranging from Ordovician–Silurian (blue) until the Ordovician–Pleistocene (yellow). The colour coding of the lines obeys the library size. For a

given time window, the analysis is convergent (solid lines) if the prediction skills are higher at large library sizes than at lower library sizes, and non-convergent if not (dashed lines).

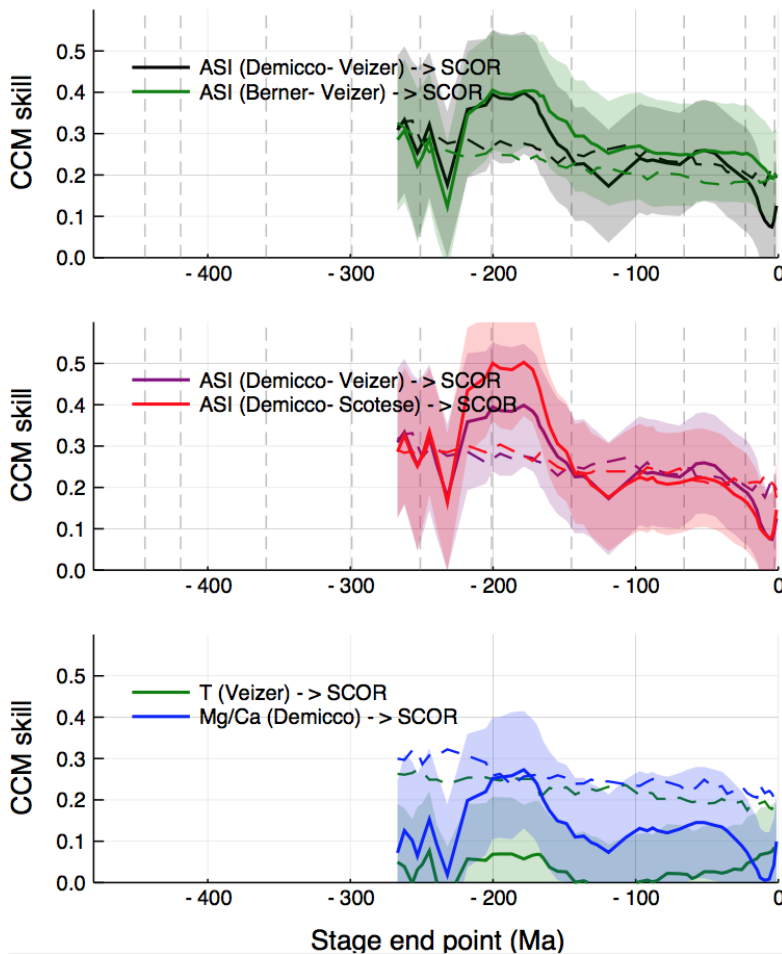


Figure S4b: Cross mapping sensitivity to uncertainty in SCOR and ASI records, and to the choice of Mg/Ca model. Error envelopes are the standard deviation of the mean CCM skill over an ensemble of 200 random realizations of the ASI and SCOR data. Each ensemble member is constructed by drawing a random realization within the 5-95% quantile range (for SCOR) and 33-67 % quantile range (for ASI, which has much larger uncertainties) of the data uncertainties. Dashed lines are the 95th percentiles of the median CCM skill over 300 surrogate realizations that are drawn in the same manner as the original data, but where the putative response time series is randomly shuffled. Causal interpretations (opposite of cross mapping direction) are indicated.

S3. Aragonite bias

Aragonitic shells are less likely to be preserved in the fossil record than calcitic taxa^{51,96,97}. This would jeopardize the veracity of our results about the relative success of calcitic and aragonitic taxa, if the relative preservation potential of aragonite and calcite were to differ through time. To avoid temporal differences in aragonite preservation, we excluded occurrences from unlithified sediments, as those provide a different preservation potential than lithified rocks⁹⁸; we also excluded all collections with aragonite preservation. Both of those preservational exceptions are predominantly found in the Cenozoic, and their inclusion might overestimate SCOR_{ara} in younger stages. Preservation bias against aragonitic taxa seems to be prominent in carbonates, but less significant in siliciclastic sediments²⁶. Calculating SCOR separately in both environments thus allows us to estimate the impact of aragonite loss on our analysis: SCOR_{ara} is lower in carbonate environments than in siliciclastic settings in 53 out

of 85 stages (see Figure S5a). The mean ratio of carbonate : siliciclastic $SCOR_{ara}$ is 0.97. The ratio varies through time, but is not systematically higher in older time bins (Fig. S5b): A linear model, $\log(SCOR_{ara, carbonate} / SCOR_{ara, siliciclastic})$ against time, is not significant ($p = 0.16$). If older stages were more affected by aragonite loss, we would expect higher $SCOR_{ara}$ in siliciclastic settings than in carbonate settings in those stages, and a decrease of the discrepancy towards the present – i.e., a positive slope in the regression. We conclude that while an aragonite bias might affect the data *within* stages, it acts in a temporally consistent way, and relative changes through time are biologically meaningful.

As a second test to compare the completeness of the fossil record of aragonitic and calcitic taxa, we calculate the proportion of gaps among them⁹⁹. A gap describes the absence of a genus in a stage in which it is known to have lived due to occurrences of that genus in older and younger stages. The gap proportion (gp) of a group of genera in stage s is the number of gaps (g_s), divided by g_s plus the number of genera actually encountered in s (h_s):

$$(1) \quad gp = \frac{g_s}{g_s + h_s} .$$

The gap proportion throughout the Ordovician – Pleistocene is similar for aragonitic and calcitic taxa (Fig S6a). The median gp of aragonitic taxa is 0.41, slightly higher than the median gp of calcitic taxa, 0.38. A Wilcoxon signed-rank test shows that difference in ranks of aragonitic and calcitic gp is not significant ($p = 0.28$).

A narrower definition of a gap is the “part-timer”¹⁴. A part-timer is a genus that is not encountered in stage s , but is found in the stage immediately before and in the stage immediately after s . We calculate the part-timer proportion (pp) in stage s by dividing the number of part-timers (pt) by pt plus the number of three-timers ($3t$), i.e. the genera that were found in s and in the stage immediately before and in the stage immediately after s :

$$(2) \quad p(pt) = \frac{pt_s}{pt_s + 3t_s} .$$

pp of aragonitic and calcitic taxa, too, are similar (Fig S6b). The median pp is 0.22 for aragonitic taxa and 0.19 for calcitic taxa, and the difference is not significant when assessed with a Wilcoxon signed-rank test ($p = 0.52$).

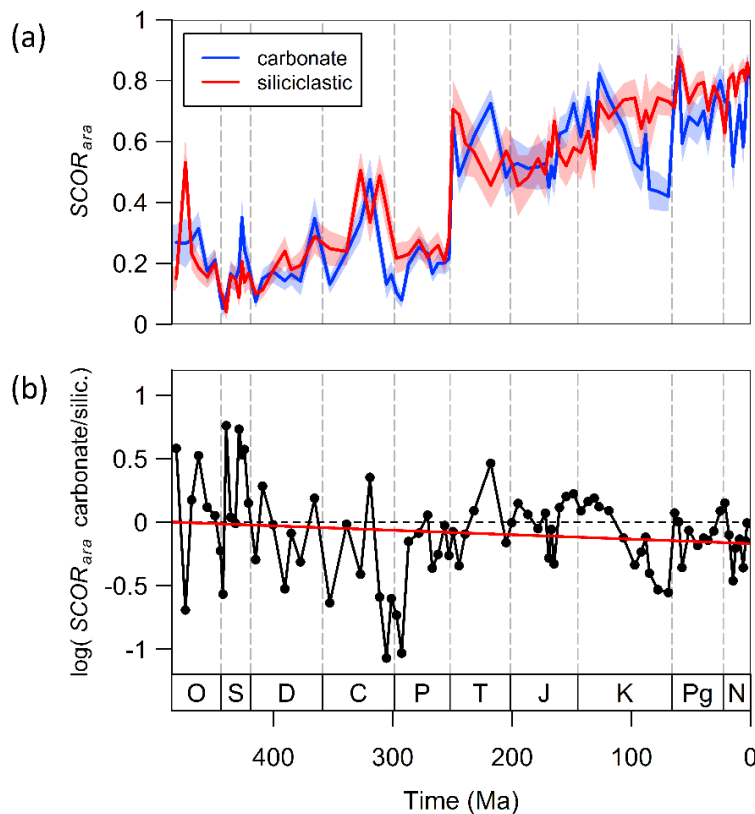


Figure S5: Carbonate vs siliciclastic settings.

(a) The relative success of aragonitic calcifiers, $SCOR_{ara}$, in carbonates (blue) and siliciclastic settings (red) in 85 Ordovician – Cenozoic stages. Shaded envelopes denote 95% confidence intervals.

(b) Natural logarithms of the ratio of the $SCOR_{ara}$ in carbonates and in siliciclastic settings, $\log(SCOR_{ara, carbonate} / SCOR_{ara, siliciclastic})$, in 85 Ordovician – Cenozoic stages. The red line shows a linear model of $\log(SCOR_{ara, carbonate} / SCOR_{ara, siliciclastic})$ against time. The model is not significant ($R^2 = 0.02$, $p = 0.16$, intercept = -0.17, slope = -0.00035).

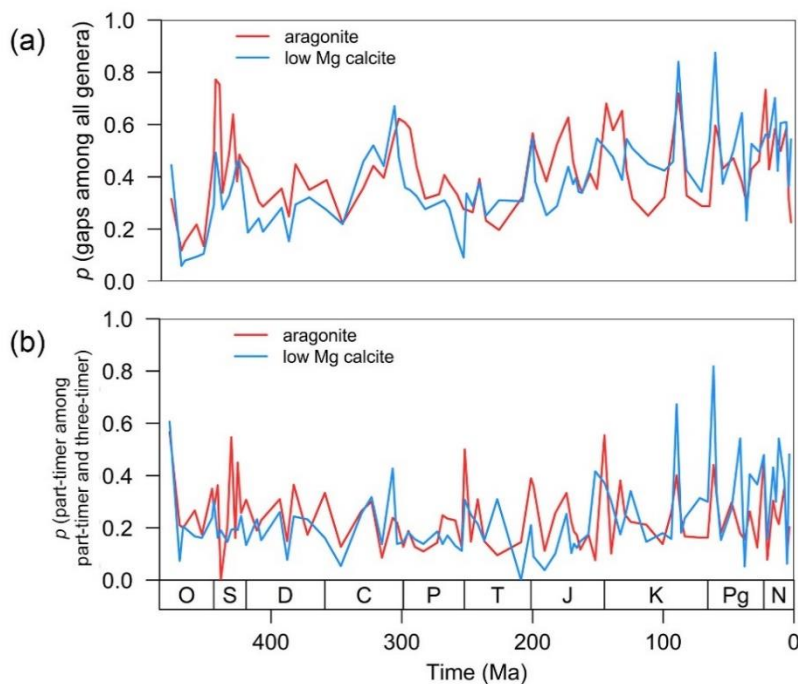


Figure S6: Sampling completeness of aragonitic and calcitic genera.

(a) Gap proportions of aragonitic and calcitic genera (sensu Paul 1982)⁹⁹. For every genus, the span between their first and their last occurrence is calculated. The number of gaps in a stage s is the number of genera which don't occur in s , but which occur in any stage before and after s . This gap number is divided by the sum of the number of genera that occur in stage s and the gap number, to get the gap proportions.

(b) Part-timer proportions of aragonitic and calcitic genera (sensu Alroy 2014)¹⁴. Part-timers are genera that don't occur within stage s , but do occur in the stages immediately before and after s . The number of part-timers is divided by the number of three-timers, i.e. genera that occur in and immediately before and after stage s , plus the number of part-timers, to get the part-timer proportion.

S4. Unbuffered taxa

Taxonomic groups that are thought to be more vulnerable to changes in the physiochemical environment are commonly referred to as “unbuffered”¹. We expect the subset of unbuffered taxa to react to changes in ASI consistent with the patterns in the full data set, but perhaps even more strongly. We therefore calculated $SCOR_{ara}$ with only unbuffered taxa, following the classification of ref. 25, except we included cephalopods following ref. 2 (Table S1). The patterns seen in the entire data set are essentially confirmed in the unbuffered subset (Fig. S7), although the R^2 of linear models in expanding windows are lower in most of the Palaeozoic windows, partly because models with autoregressive errors were selected from the Devonian onwards. In the Mesozoic – Cenozoic, a relationship between $SCOR_{ara}$ and ASI can be inferred up to the Middle Jurassic (Fig. S7b).

Table S1: Unbuffered taxa. Selection of physiologically unbuffered taxa is based on ref. 25. We added Cephalopods following ref. 2.

Higher taxon	remark
Cephalopods	excluding Nautilida
Epifaunal Bivalves	including semi-infaunal bivalves
Stenolaemate bryozoans	
Cheilostome bryozoans	
Corals	
Ostracods	
Calcifying sponges	
Foraminifera	
Calcareous algae	
Rhynchonelliform brachiopods	
Echinoderms	

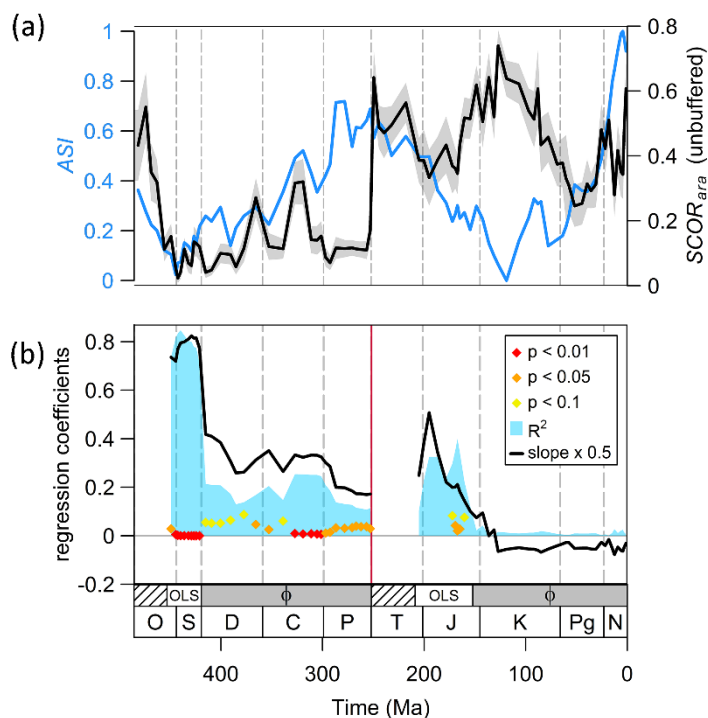


Figure S7: SCOR_{ara} of unbuffered taxa and aragonite sea intensity (ASI).

(a) ASI (blue) and the relative success of aragonitic calcifiers, SCOR_{ara}, in the subset of unbuffered taxa (black) in 85 Ordovician – Pleistocene stages. The grey envelope indicates the 95% confidence interval of SCOR_{ara}.

(b) Linear models of SCOR_{ara} in the subset of unbuffered taxa against ASI in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately. Slope (black) and R² (blue) of each model are drawn at the last included stage of the respective window. *p*-values are only shown when they are < 0.1. All Palaeozoic models start at the first Ordovician stage (Tremadocian), and all Mesozoic – Cenozoic models start with the Early Triassic (Induan). The box at the bottom shows whether the models were generated using ordinary least squares (OLS), or generalised least squares with autoregressive errors of the first order (ϕ). No models were calculated for intervals with five or less stages (hatched pattern).

S5. Major taxonomic groups

Bivalves, brachiopods, gastropods, cephalopods and corals are the most common calcifying clades in the data set. For every stage, we evaluated clade success through time by dividing the SCOR of the taxonomic group of interest by the SCOR of all calcifiers. Changes in the relative SCOR of taxonomic groups are thus always influenced by simultaneous changes in the success of other calcifying groups. For individual groups, a smaller amount of data is available than for the entirety of aragonitic calcifiers, which leads to more volatile patterns of group success through time. This makes estimating the extent of ASI influence on the SCOR of individual taxonomic groups less reliable than the analysis with SCOR_{ara}, but several interesting patterns emerge nonetheless.

Brachiopods. Calcifying brachiopods are calcitic, with the exception of the trimellerids, which are aragonitic⁵² and are not included in the brachiopod subset. Brachiopod SCOR is calculated among the subset of benthic taxa. Brachiopods were the most successful calcifying clade from the late Ordovician to the End-Permian mass extinction. They retained a moderate SCOR until the Middle Jurassic, and almost continuously declined thereafter (Fig. S8a). Brachiopod SCOR and ASI have a significant negative linear relationship over much of the Ordovician – Carboniferous (Figure S8a). The relationship disappears with the addition of the Permian, presumably due to the high CaCO₃ supersaturation of the ocean at that time. After the Jurassic, brachiopods were excluded from most shallow water,

benthic communities despite calcite sea conditions. Ecological interactions with competitors such as bivalves, rather than environmental changes, may have been the cause of this decline¹⁰⁰.

Bivalves. Bivalves build either purely aragonitic shells, or shells comprised of aragonitic and calcitic layers¹⁰¹. If the proportion of calcite in the shells exceeds 90 %, we treat them as calcitic (see ref. 25 for a detailed description of how mineralogies were assigned). Bivalve shell mineralogy is largely determined by phylogeny: Palaeoheterodonta, Protobranchia and taxa of unknown affinity are exclusively aragonitic, as are most non-rudist Heterodonta (Fig. S9). Most Pteriomorphia are bimineralic, with e.g. oysters having many dominantly calcitic taxa¹⁰¹. We calculated separately the success of aragonitic, calcitic and bimineralic bivalves relative to all calcifying taxa, only including benthic occurrences (Fig. S8e-g). A positive linear relationship between the SCOR of aragonitic bivalves and ASI is evident in the Palaeozoic (Fig. S8e). The negative relationship of aragonitic bivalves and ASI inferred for the Triassic – Early Cretaceous is probably spurious and disappears in the linear models in which autocorrelation is considered.

Patterns in the SCOR of bimineralic bivalves are difficult to interpret. In the Devonian – Carboniferous, and less robustly also in the Mesozoic, their SCOR is positively correlated with ASI. We did not include the Ordovician – Silurian, as bivalves with calcitic shell layers are rare or absent during much of the Ordovician and Silurian. In contrast, an earlier study reported a Phanerozoic correspondence between calcite seas and diversity increases of bivalves with calcitic shell layers²⁰. The majority of these partly calcitic bivalves are included in our bimineralic subset. Considering ASI rather than the binary aragonite sea – calcite sea state, we cannot confirm a positive relationship between bimineralic bivalve SCOR and calcite sea conditions.

The success of dominantly calcitic bivalves could only be evaluated in the Mesozoic – Cenozoic, as they were rare in the Palaeozoic. Calcitic bivalves were most successful during the Jurassic – Cretaceous, and the Middle Jurassic – Cretaceous is an interval of relatively low ASI (Fig. S8f). A negative linear relationship of calcitic bivalve SCOR and ASI is significant in intervals extending up to the Early Cretaceous and disappears thereafter.

There is some evidence that pteriomorphs exert more control over the secretion of calcite prisms than heterodont bivalves¹⁰², and the only time when bimineralic heterodonts were successful was during the Cretaceous calcite sea (Fig. S9d). Calcitic pteriomorphs, too, were most successful during the Jurassic and Cretaceous, but calcitic pteriomorph SCOR rose in the Triassic and Early Jurassic aragonite sea (Fig. S9c). The overall relative SCOR of heterodonts and pteriomorphs among all bivalves seems to be unrelated to aragonite-calcite sea conditions (Fig. S9a).

The quantitative evidence for a relationship of bivalve SCOR with ASI remains ambiguous. The dominant pattern in the history of bivalves is a long-lasting increase in success, especially following the end-Permian extinction.

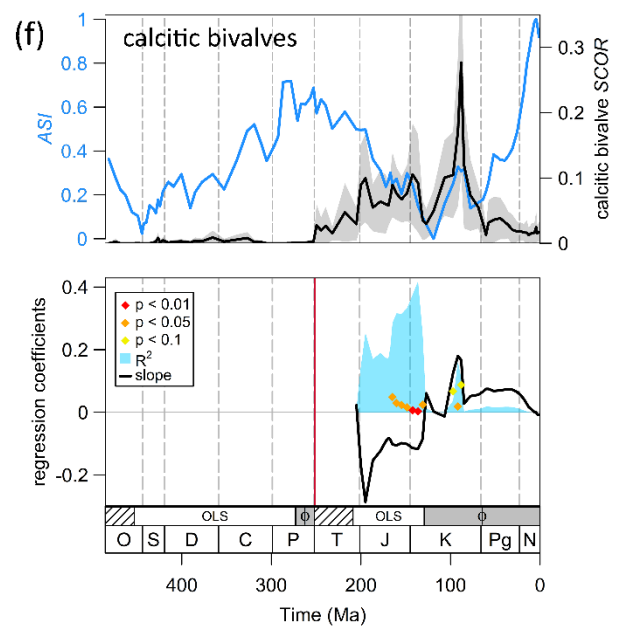
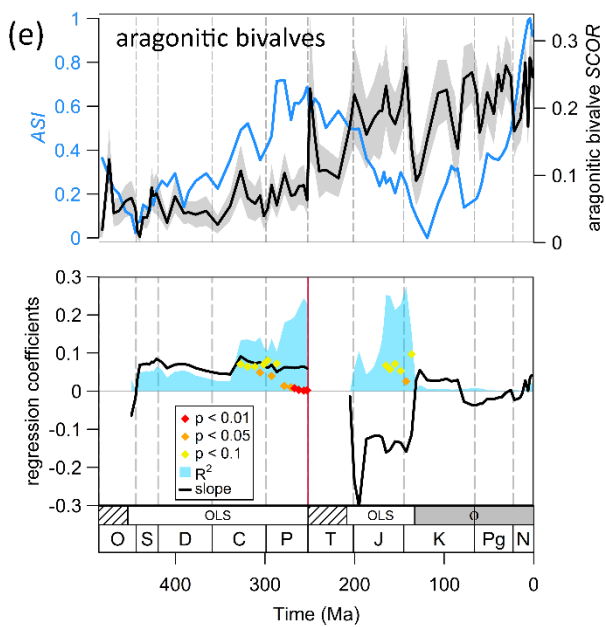
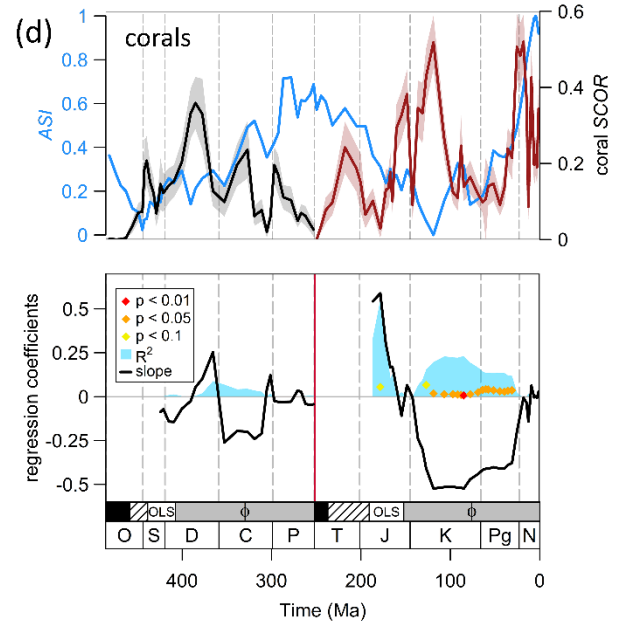
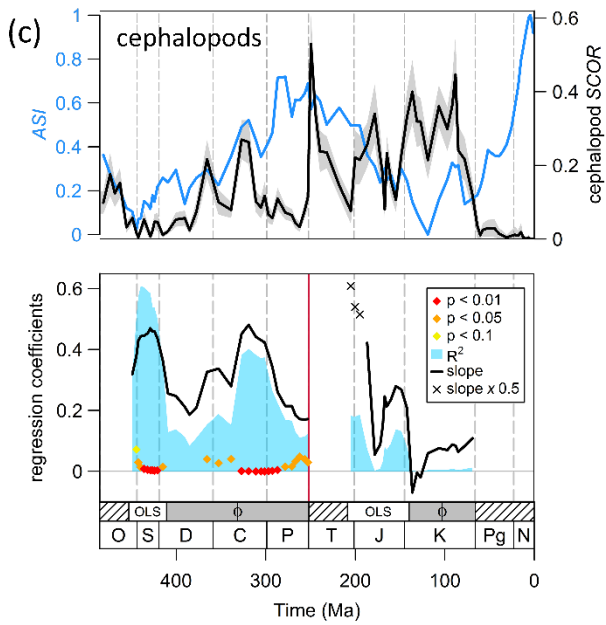
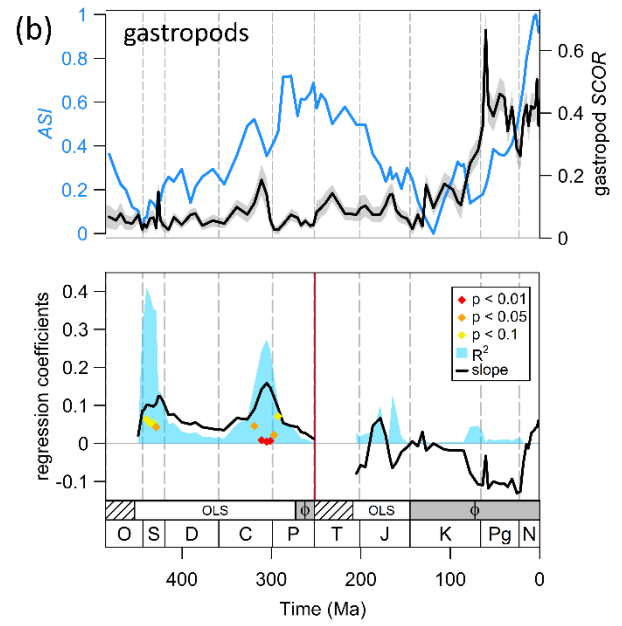
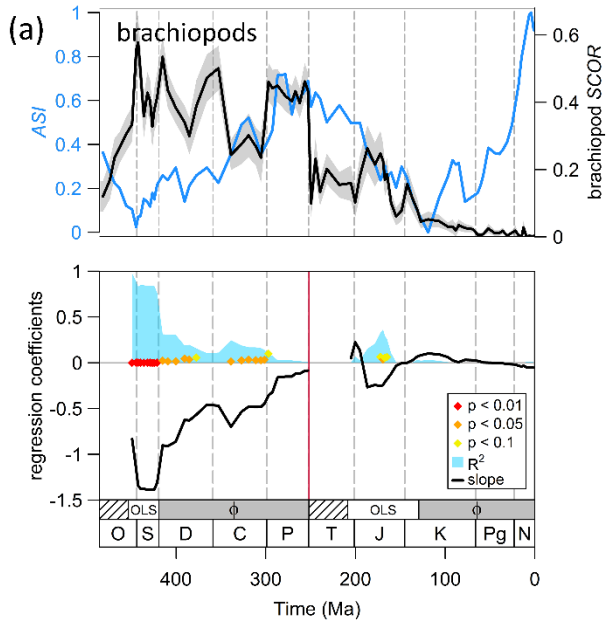
Gastropods. Gastropods are mostly aragonitic, and we excluded all taxa with calcitic shell layers comprising > 10 % of the total shell for the calculation of gastropod SCOR. Analysis was carried out only among benthic taxa. A positive linear relationship of gastropod SCOR and aragonite sea intensity is significant in some intervals in the Ordovician – Carboniferous, weakens in the Permian, and no relationship can be inferred in the Mesozoic – Cenozoic (Fig. S8b).

Cephalopods. Cephalopod SCOR was calculated without the calcitic belemnites. Due to the extinction of most shelled cephalopods at the Cretaceous - Palaeogene boundary, Cenozoic stages were not included in the linear models. The linear relationship between the SCOR of cephalopods relative to all calcifiers and ASI is strong in the Palaeozoic, but not significant in the Mesozoic (Fig. S8c). The decreased response of Mesozoic cephalopods to aragonite-calcite sea conditions may have to do with the extinction of most Palaeozoic cephalopods at the end-Permian mass extinction: Ammonoids were

the dominant clade of Mesozoic cephalopods, and almost all of their Mesozoic members were descendants of a few survivors¹⁰³. It is possible that the surviving ammonoids were less susceptible to physiochemical changes than Palaeozoic cephalopods on average, as the end-Permian extinction selectively favoured survival of physiologically buffered taxa²⁸.

Corals. Corals were analysed separately in the Palaeozoic and the Mesozoic – Cenozoic. Palaeozoic corals, Tabulata and Rugosa, were calcitic, whereas scleractinian corals are aragonitic. Corals occur preferentially in carbonate settings. To avoid distortion of temporal patterns of coral SCOR by changing abundances of occurrences from carbonate and siliciclastic settings, respectively, we calculated coral SCOR only in carbonate settings and only included benthic occurrences in the analysis as in ref. 25. Corals were rare in the Early – Middle Ordovician and modern corals only appeared in the Middle Triassic; hence we excluded the Early – Middle Ordovician and Early – Middle Triassic when estimating models of coral SCOR against ASI.

Contrary to expectations, of the main taxonomic groups, corals show the least response to changing ASI (Fig. S8d). The linear models of coral SCOR against ASI are not significant in the Palaeozoic. In the Triassic – Cretaceous until the Triassic – Palaeogene, significant relationships with a negative slope persist. However, if aragonite-calcite seas influenced the success of aragonitic corals, ASI should be positively correlated with coral SCOR. Calcification of modern corals is aided by photosymbiosis, and is biologically controlled to a high degree¹⁰⁴, and some Palaeozoic corals may have harboured photosymbionts as well¹⁰⁵. However, experiments showed a strong decrease of scleractinian coral growth under calcite sea conditions¹⁰⁶. Scleractinian skeletal mineralogy can be influenced by Mg/Ca ratio and temperature, although mostly higher aragonite proportions than predicted by the inorganic CaCO₃ system were retained in experiments¹⁷. The lack of a long-term influence of aragonite-calcite sea conditions on the SCOR of corals may stem from the affinity of corals to reef settings. Reefs are hotspots of evolution, diversity and biotic interactions¹⁰⁷, and they may well be a hotspot for biotically, rather than environmentally controlled evolution. The Phanerozoic fossil record of reefs exhibits a “boom and bust pattern” that cannot be explained by secular environmental changes¹⁰⁸.



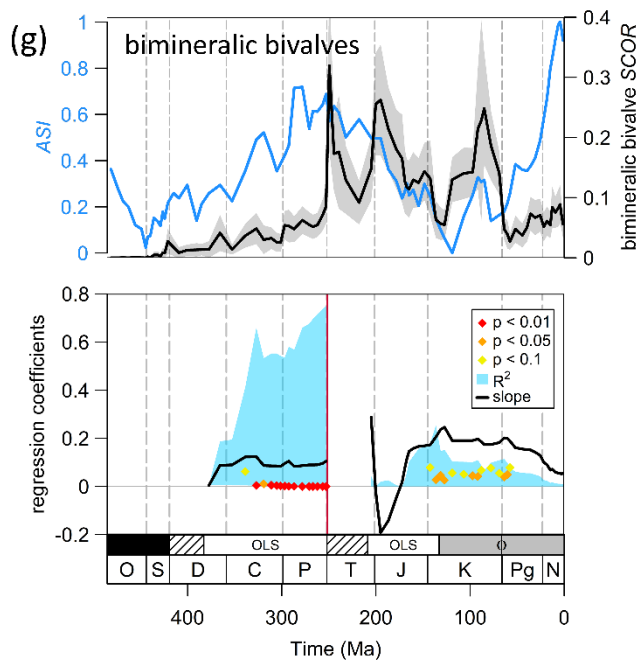


Figure S8a-g: Relative SCOR of major taxonomic groups (brachiopods, gastropods, cephalopods, corals, bivalves), and linear models with ASI.

Upper figures: Aragonite sea intensity (blue) and the relative success of a major taxonomic group, SCOR (black) in 85 Ordovician – Cenozoic stages. The grey or red envelope indicates $1.96 \times$ standard error of SCOR.

Bottom figures: Linear models of the SCOR of taxonomic groups against ASI in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately. Slope (black) and R^2 (blue) of each model are drawn at the last included stage of the respective window. p -values are only shown when they are < 0.1 . All Palaeozoic models start at the first Ordovician stage (Tremadocian), and all Mesozoic – Cenozoic models start with the Early Triassic (Induan). The box at the bottom shows whether the models were generated using ordinary least squares (OLS), or generalised least squares with autoregressive errors of the first order (ϕ). No models were calculated for intervals with five or less stages (hatched pattern).

(a) Brachiopod SCOR includes only calcitic brachiopods. The analysis was carried out with only benthic taxa.

(b) Gastropod SCOR includes only aragonitic gastropods. The analysis was carried out with only benthic taxa.

(c) Cephalopod SCOR includes only aragonitic cephalopods (i.e., belemnites were excluded).

(d) Corals. Palaeozoic coral SCOR includes only calcitic corals, and the Lower – Middle Ordovician are excluded from the linear model. Mesozoic – Cenozoic coral SCOR includes only aragonitic corals, and the Lower – Middle Triassic are excluded from the linear models. The analyses were carried out with occurrences only from carbonate environments, and only with benthic taxa.

(e) Aragonitic bivalve SCOR includes bivalves with $> 90\%$ aragonite in their skeleton. The analysis was carried out with only benthic taxa.

(f) Calcitic bivalve SCOR includes bivalves with $> 90\%$ calcite in their skeleton. Only the Mesozoic – Cenozoic model is estimated, due to the rarity of dominantly calcitic bivalves in the Palaeozoic. The analysis was carried out with only benthic taxa.

(g) Bimineralic bivalve SCOR includes bivalves with $> 10\%$ aragonite and $> 10\%$ calcite in their skeleton. The Ordovician – Silurian are excluded from the linear model. The analysis was carried out with only benthic taxa.

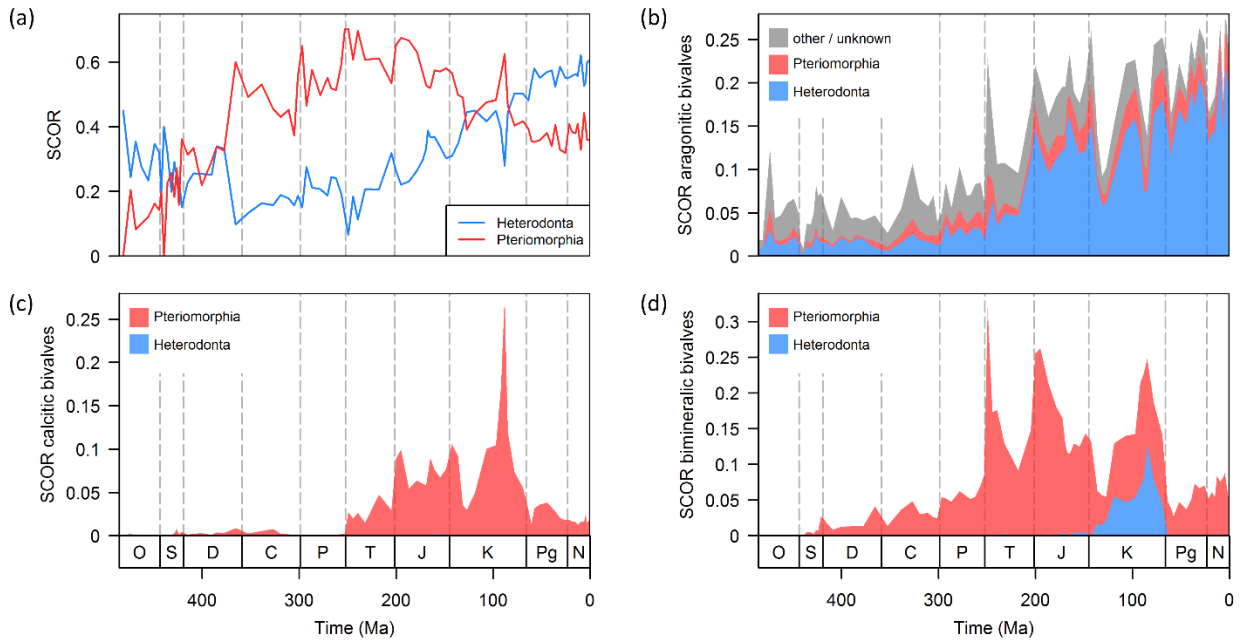


Figure S9: Bivalve subclasses.

(a) Relative SCOR of the two dominant bivalve subclasses among all bivalves: Heterodonta (blue), and Pteriomorphia (red).

(b) SCOR of aragonitic bivalves as in Fig. S8e, broken down into bivalves subclasses. The contribution to SCOR of Heterodonta (blue), Pteriomorphia (red), and other or unknown bivalve subclasses (grey) are shown.

(c) SCOR of calcitic bivalves as in Fig. S8f, broken down into bivalves subclasses. Only Pteriomorphia (red) contribute to SCOR, except for a minor contribution of Heterodonta (blue) in the Upper Jurassic.

(d) SCOR of bimineralic bivalves as in Fig. S8g, broken down into bivalves subclasses. Only Pteriomorphia (red) and Heterodonta (blue) contribute to SCOR.

S6. *De novo* acquisition of skeletons and changes in skeletal mineralogy

1. *De novo* acquisition

The skeletal mineralogy of clades of marine calcifiers is in most cases determined by the seawater composition of the time at which the clade first evolved a skeleton^{21,109,110}. These *de novo* acquisitions of skeletons have the potential to shift SCOR_{ara} without necessarily changing the real ecological success of involved taxa, as the newly calcifying taxa might have been successful even before secreting mineralised hard parts.

Novel acquisitions of mineralised hard parts are rare – Porter (2010) lists 40 acquisition events of calcium carbonate skeletons ranging from the Ediacaran – Palaeogene, most of them concentrated in the early Palaeozoic²¹. Of the major clades composing our data set, molluscs, calcifying brachiopods, trilobites and echinoderms acquired their skeleton already in the Cambrian, and those together make up 79 % of the total occurrences in our data (Fig. S10, Table S2). The *de novo* acquisitions of their skeleton thus cannot drive the patterns seen in SCOR_{ara}. Of the remaining occurrences in our data, 13 % belong to taxa that acquired their calcareous skeletons during the Ordovician, which minimises their potential impact on the observed Jurassic decoupling of ASI and SCOR_{ara}. A further 4.9 % of the occurrences in the data set are from taxa evolving a skeleton in the Triassic – Palaeogene, the large majority of those occurrences are scleractinian corals (4.3 %) which originated in the Middle Triassic. Demosponges and calcifying red and green algae make up most of the remaining 2.6 % of occurrences, and those cannot be assigned to a single timing of skeletal origin¹¹¹⁻¹¹³.

To test whether the Mesozoic patterns in the relationship of ASI and SCOR_{ara} are driven by skeletal acquisitions, we removed all groups that acquired a skeleton *de novo* in the Mesozoic or Cenozoic²¹ and calculated SCOR_{ara} with only the remaining taxa (Fig. S11a). The taxa excluded for this analysis are scleractinian corals, cheilostomate bryozoans and several other, rare taxonomic groups. The resulting correlation (Fig. S11b) is very similar to results obtained for the total dataset (Fig. 2b). The timing of *de novo* skeletal acquisitions therefore cannot explain the early Mesozoic correlation and later decoupling of SCOR_{ara} and ASI.

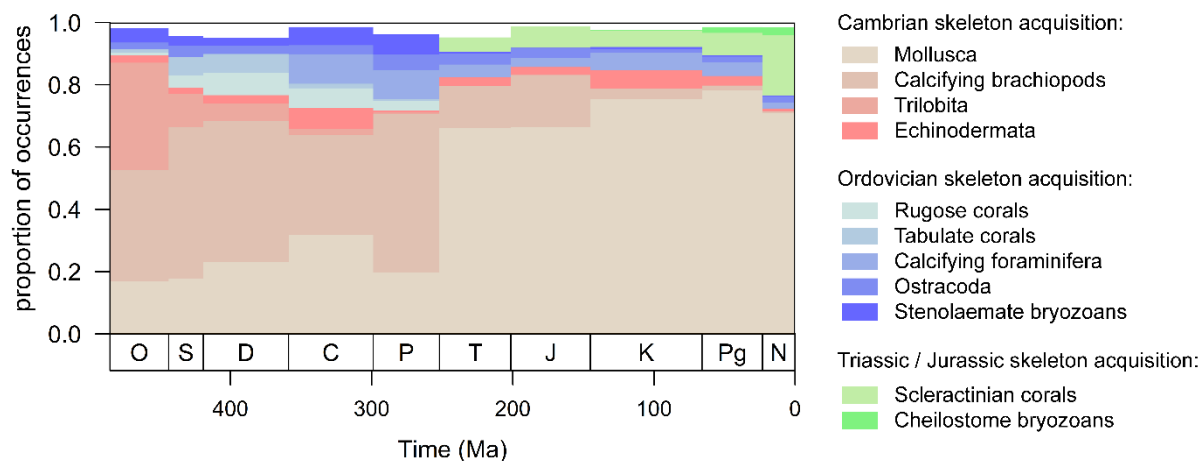


Figure S10: Frequency of major calcifying clades

Proportions of occurrences of major calcifying groups in the PBDB data set in geological periods. The group proportions are displayed in order of their evolutionary acquisition of a calcium carbonate skeleton^{21,114}: Clades with a Cambrian origin of their skeleton are shown in beige – red, an Ordovician origin is denoted with blue colours, and clades with a Triassic (scleractinian corals) or Jurassic (cheilostome bryozoans) skeletal acquisition are shown in green.

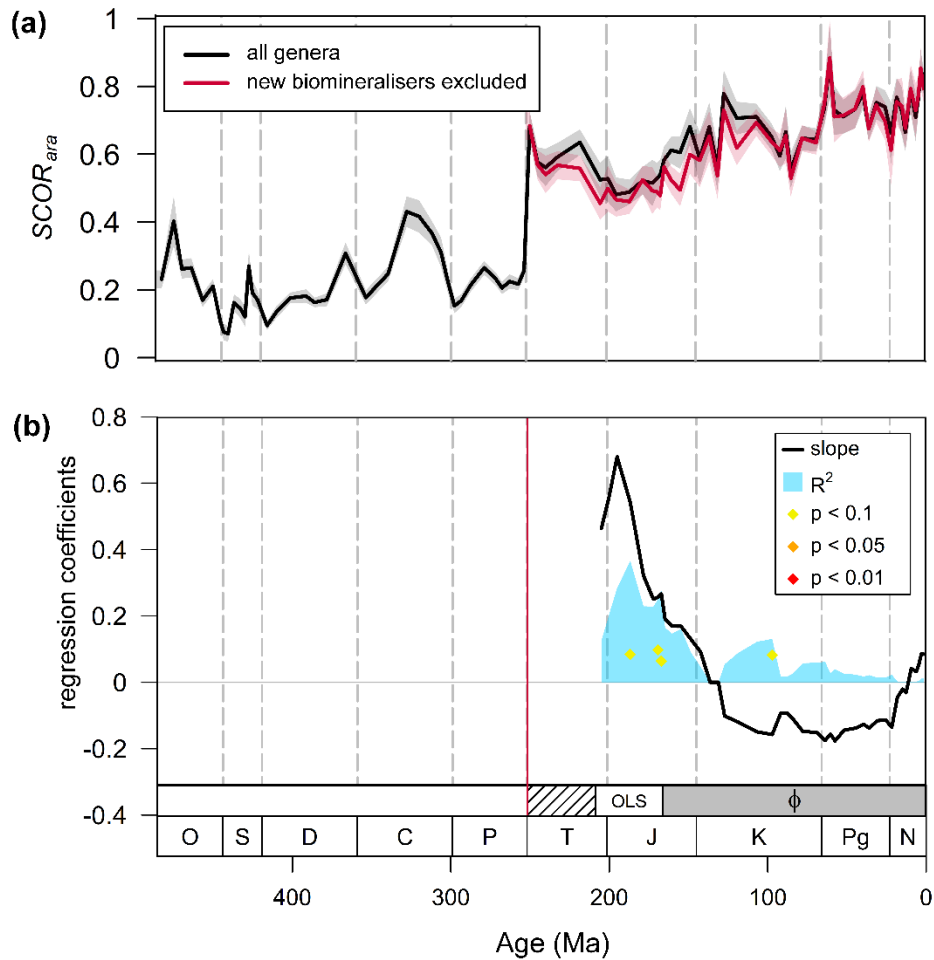


Figure S11a: $SCOR_{ara}$ without originating genera and aragonite sea intensity (ASI).

(a) Relative success of aragonitic calcifiers, $SCOR_{ara}$, of all genera (black) and of genera excluding taxa that acquired their skeletons in the Mesozoic or Cenozoic (red), respectively, in 85 Ordovician – Pleistocene stages. The excluded taxa comprise Scleractinia, cheilostomate bryozoans, serpulid worms and several other anthozoan orders. The envelopes indicate 95% confidence intervals.

(b) Linear models of $SCOR_{ara}$ excluding Mesozoic and Cenozoic skeletal acquisitions as in (a), against ASI in windows of increasing length in the Mesozoic – Cenozoic. Slope (black) and R^2 (blue) of each model are drawn at the last included stage of the respective window. p -values are only shown when they are < 0.1 . All models start with the Early Triassic (Induan). The box at the bottom shows whether the models were generated using ordinary least squares (OLS), or generalised least squares with autoregressive errors of the first order (ϕ). No models were calculated for intervals with five or less stages (hatched pattern). The pattern is very similar to that in Fig. 2b with the $SCOR_{ara}$ of all genera.

Table S2: Timing of *de novo* acquisition and contribution to our dataset

Most taxa in our data set evolved their skeleton in the Cambrian, Ordovician or in the Triassic – Jurassic. These taxa and the percentage they constitute in our entire Ordovician – Pleistocene data set are listed below. The CaCO₃ skeletons of calcifying algae and demosponges have multiple evolutionary origins¹¹¹⁻¹¹³, and a further 0.97 % of remaining occurrences has origins in the middle – late Palaeozoic, or an origin could not be assigned.

Taxon	Skeletal acquisition	Occurrences %
Mollusca	Cambrian	48.1
Calcifying brachiopods	Cambrian	25.0
Trilobita	Cambrian	3.28
Echinodermata	Cambrian	2.90
Calcarea	Cambrian	0.09
Hyalolitha	Cambrian	0.04
total	Cambrian	79.32
Calcifying foraminifera	Ordovician	4.55
Ostracoda	Ordovician	2.93
Stenolaemate bryozoans	Ordovician	2.53
Rugose corals	Ordovician	2.03
Tabulate corals	Ordovician	1.11
Machaeridia	Ordovician	0.01
total	Ordovician	13.16
Scleractinian corals	Triassic	4.31
Cheilostome bryozoans	Jurassic	0.39
other	Mesozoic-Cenozoic	0.18
total	Mesozoic - Cenozoic	4.88
Demospongia		1.00
Chlorophyta		0.36
Rhodophyta		0.29
other		0.97
total	various	2.62

2a. Changes in skeletal composition and SCOR_{ara}

Changes in skeletal mineralogy after a clade had acquired a specific skeletal composition are a second possibility of how SCOR_{ara} could change without an actual change in the ecological success of the constituent taxa. With the exception of some bivalve and cheilostome genera, skeletal mineralogy tends to be consistent within families, and more so within genera (see section 2b below.). A SCOR_{ara} time series with minimal contribution from changing skeletal mineralogies can thus be generated by excluding newly evolved genera from the data set (Fig. S12a). The resulting curve is very similar to the SCOR_{ara} of the full data set, and although the correlation with ASI is somewhat weaker in the Palaeozoic, it shows the same temporal pattern of a decoupling between SCOR_{ara} and ASI (Fig. S12b). Thus neither *de novo* acquisition of calcareous skeletons, nor changes of mineral composition within calcifying clades can account for the overall patterns of correlation between ecological success and aragonite sea intensity.

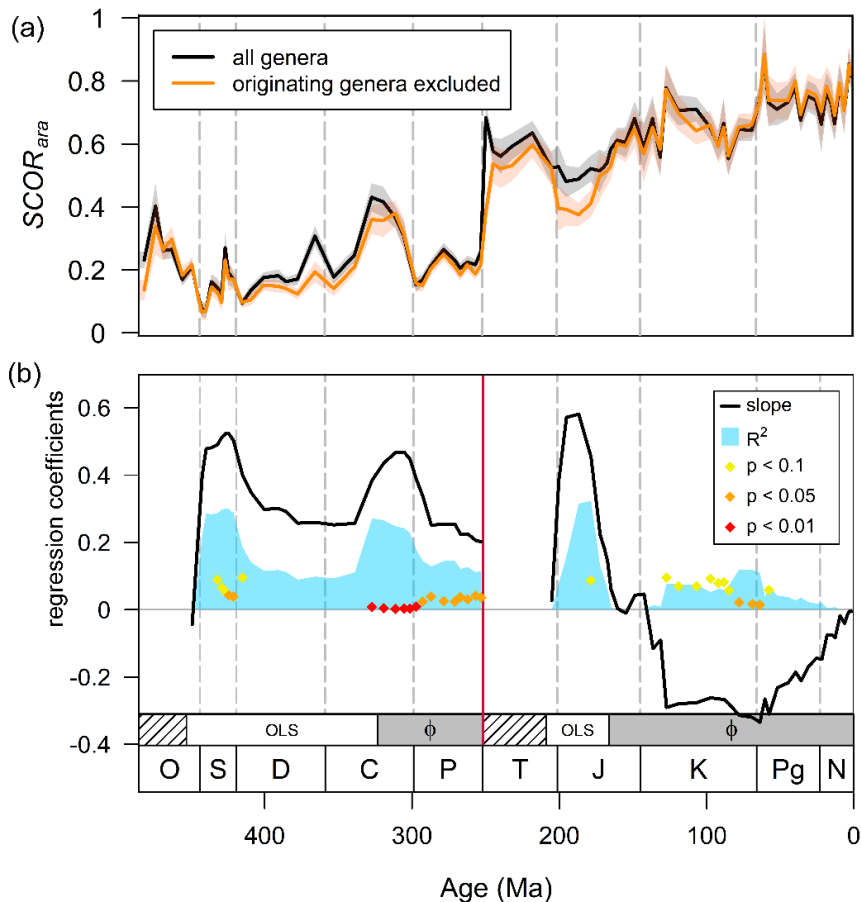


Figure S12: $SCOR_{ara}$ without originating genera and aragonite sea intensity (ASI).

(a) Relative success of aragonitic calcifiers, $SCOR_{ara}$, of all genera (black) and genera excluding first appearances (originating, orange), respectively, in 85 Ordovician – Pleistocene stages. The envelopes indicate 95% confidence intervals.

(b) Linear models of $SCOR_{ara}$ without originating genera against ASI in windows of increasing length in the Palaeozoic and in the Mesozoic – Cenozoic, separately. Slope (black) and R^2 (blue) of each model are drawn at the last included stage of the respective window. p -values are only shown when they are < 0.1 . All Palaeozoic models start at the first Ordovician stage (Tremadocian), and all Mesozoic – Cenozoic models start with the Early Triassic (Induan). The box at the bottom shows whether the models were generated using ordinary least squares (OLS), or generalised least squares with autoregressive errors of the first order (ϕ). No models were calculated for intervals with five or less stages (hatched pattern). The pattern is very similar to that in Fig. 2b, which includes originating data.

2b. Mineralogy changes in major clades

In cheilostome bryozoans, foraminifers, gastropods and especially in bivalves, multiple taxa have changed their skeletal mineralogy, commonly by the addition of a shell layer with a different mineralogy^{20,114-117}. In gastropods, significant skeletal calcite content is concentrated within a limited range of families and genera⁶⁹. The skeletal mineralogy of foraminifers can be resolved well along higher taxonomic divisions^{114,118}. In bivalves, the addition of calcitic shell layers to an originally aragonitic skeleton has evolved several times. Skeletal mineralogies are consistent within the majority of bivalve families¹¹⁹, although e.g. the mineralogy of bivalves in the genus *Mytilus* differs between species and individuals in different habitats¹²⁰. Similarly, some cheilostome bryozoans have significant mineralogical differences within genera¹¹⁵. In most major clades, however, changes in mineralogy are rare: trilobites, echinoderms, ostracods are exclusively calcitic, and brachiopods and stenolaemate bryozoans are generally calcitic, with the exception of trimerellid brachiopods⁵² and a few genera of Stenolaemata with minor skeletal aragonite content¹¹⁵. Ammonoids, nautiloids and scleractinian corals are, with rare exceptions in the latter two groups¹²¹⁻¹²³, aragonitic.

S7. Data cleaning and data summary

To resolve incomplete higher taxonomies of individual occurrences in the Paleobiology Database, we used taxonomic information from other PBDB occurrences and, when available, from the World Register of Marine Species (WoRMS)¹²⁴ using the taxize package¹²⁵.

Hierarchical assignments as implemented in the PBDB were used to fill in missing skeletal mineralogies, and the skeletal composition of several higher taxa was corrected following ref. 25 and references therein. Occurrences were excluded if the skeletal composition could not be resolved or if the occurrences could not be assigned to a genus. All filters that were applied to clean the data are listed in Table S3. The final data set comprises 398,199 occurrences. The number of occurrences and genera in every stage are given in Table S4.

Table S3: Data cleaning. Criteria based on which we excluded PBDB occurrences, and links to relevant information regarding these criteria in the PBDB.

Criterion	Category and reference to definitions
Marine	Palaeoenvironment (http://paleodb.org/public/tips/environtips.html)
Identified to genus level	Taxonomy (http://paleodb.org/trac/log/html/public/tips/taxontips.html?rev=748)
Is an invertebrate animal, or Foraminifera, Chlorophyta, Rhodophyta, or calcifying "Problematica"	Taxonomy
Is not plankton	Taxonomy, life habits (http://paleodb.org/public/tips/ecology_tips.html)
Assignable to an Ordovician – Pleistocene stage	Stratigraphy (http://paleodb.org/public/tips/tips.html)
No aragonite preservation recorded	Preservation (http://paleodb.org/public/tips/tips.html)
Lithology not "unlithified"	Lithology (http://paleodb.org/public/tips/tips.html)
Palaeocoordinates available	
Is calcifying	Skeletal mineralogy (ref. 25)

Table S4: Data summary. Number of genera, occurrences, and occurrences per million year for every stage of the final PBDB data set. The Induan and Olenekian are treated as one stage, as well as the four Pleistocene stages. Ages of the stage boundaries are taken from Gradstein *et al.* 2012; some ages have been updated following Cohen *et al.* 2013^{126,127}.

period	stage	lower stage boundary (myr)	duration	genera	occurrences	occurrences/myr
Ordovician	Tremadocian	485.4	7.7	590	3556	462
	Floian	477.7	7.7	258	935	121
	Dapingian	470	2.7	621	3195	1183
	Darriwilian	467.3	8.9	728	3464	389
	Sandbian	458.4	5.4	779	4768	883
	Katian	453	7.8	840	5626	721
	Hirnantian	445.2	1.4	423	2361	1686
Silurian	Rhuddanian	443.8	3.0	257	1561	520
	Aeronian	440.8	2.3	332	1360	591
	Telychian	438.5	5.1	549	2345	460
	Sheinwoodian	433.4	2.9	455	1947	671
	Homerian	430.5	3.1	411	1236	399
	Gorstian	427.4	1.8	419	1224	680
	Ludfordian	425.6	2.6	412	1638	630
	Pridoli	423	3.8	488	1740	458
Devonian	Lochkovian	419.2	8.4	780	4485	534
	Pragian	410.8	3.2	881	3484	1089
	Emsian	407.6	14.3	1193	5740	401
	Eifelian	393.3	5.6	879	4425	790
	Givetian	387.7	5.0	1107	15750	3150
	Frasnian	382.7	10.5	622	7149	681
	Famennian	372.2	13.3	654	3767	283
Carboniferous	Tournaisian	358.9	12.2	703	3899	320
	Visean	346.7	15.8	905	8955	567
	Serpukhovian	330.9	7.7	573	3238	421
	Bashkirian	323.2	8.0	495	2584	323
	Moscovian	315.2	8.2	626	3869	472
	Kasimovian	307	3.3	401	1930	585
	Gzhelian	303.7	4.8	509	3988	831
Permian	Asselian	298.9	3.9	677	4498	1153
	Sakmarian	295	4.9	806	6469	1320
	Artinskian	290.1	6.6	1000	6902	1046
	Kungurian	283.5	10.6	1125	11438	1084
	Roadian	272.95	4.2	1078	8769	2113
	Wordian	268.8	3.7	1009	8070	2181
	Capitanian	265.1	6.0	1017	8683	1447
	Wuchiapingian	259.1	5.0	1019	11473	2313
	Changhsingian	254.14	2.2	925	10353	4626
Triassic	Induan + Olenekian	251.902	4.7	608	9733	2070
	Anisian	247.2	5.2	664	6879	1323
	Ladinian	242	5.0	521	2369	474
	Carnian	237	10.0	824	4120	412
	Norian	227	18.2	902	5022	276
	Rhaetian	208.8	7.5	540	3607	481

period	stage	lower stage boundary (myr)	duration	genera	occurrences	occurrences/myr
Jurassic	Hettangian	201.3	2.0	296	1744	872
	Sinemurian	199.3	8.5	401	3167	373
	Pliensbachian	190.8	8.1	589	11608	1433
	Toarcian	182.7	8.6	472	9064	1054
	Aalenian	174.1	3.8	334	2438	642
	Bajocian	170.3	2.0	584	3577	1788
	Bathonian	168.3	2.2	624	4534	2061
	Callovian	166.1	2.6	757	6969	2680
	Oxfordian	163.5	6.2	785	6292	1015
	Kimmeridgian	157.3	5.2	637	4002	770
	Tithonian	152.1	7.1	668	4069	573
Cretaceous	Berriasian	145	5.2	348	1405	270
	Valanginian	139.8	6.9	442	2309	335
	Hauterivian	132.9	3.5	421	2349	671
	Barremian	129.4	4.4	600	2132	485
	Aptian	125	12.0	782	3513	293
	Albian	113	12.5	1006	6690	535
	Cenomanian	100.5	6.6	936	7521	1140
	Turonian	93.9	4.1	621	4276	1043
	Coniacian	89.8	3.5	308	1344	384
	Santonian	86.3	2.7	597	2857	1058
	Campanian	83.6	11.5	980	7304	635
	Maastrichtian	72.1	6.1	1139	14114	2314
	Palaeogene	Danian	66	4.4	907	3826
Selandian		61.6	2.4	391	1014	423
Thanetian		59.2	3.2	544	1876	586
Ypresian		56	8.2	892	4167	508
Lutetian		47.8	6.6	803	2689	407
Bartonian		41.2	3.4	972	5523	1624
Priabonian		37.8	3.9	1324	6334	1624
Rupelian		33.9	5.8	906	4670	805
Chattian		28.1	5.1	880	3816	753
Neogene	Aquitainian	23.03	2.6	434	1017	393
	Burdigalian	20.44	4.5	861	2418	541
	Langhian	15.97	2.2	635	1559	725
	Serravallian	13.82	2.2	764	2295	1048
	Tortonian	11.63	4.4	716	3058	698
	Messinian	7.246	1.9	630	2914	1523
	Zanclean	5.333	1.7	1051	4510	2602
	Piacenzian	3.6	1.0	1045	4700	4608
	Quaternary	Pleistocene	2.58	2.6	1347	9931

Supplementary References

- 2 Bambach, R. K., Knoll, A. H. & Sepkoski, J. J. Anatomical and ecological constraints on Phanerozoic animal diversity in the marine realm. *Proceedings of the National Academy of Sciences* **99**, 6854-6859 (2002).
- 10 Sandberg, P. A. An oscillating trend in Phanerozoic non-skeletal carbonate mineralogy. *Nature* **305**, 19-22 (1983).
- 14 Alroy, J. Accurate and precise estimates of origination and extinction rates. *Paleobiology* **40**, 374-397 (2014).
- 16 Hardie, L. A. Secular variation in seawater chemistry: An explanation for the coupled secular variation in the mineralogies of marine limestones and potash evaporites over the past 600 my. *Geology* **24**, 279-283 (1996).
- 17 Higuchi, T., Shirai, K., Mezaki, T. & Yuyama, I. Temperature dependence of aragonite and calcite skeleton formation by a scleractinian coral in low mMg/Ca seawater. *Geology* **45**, 1087-1090 (2017).
- 20 Harper, E. M., Palmer, T. J. & Alphey, J. Evolutionary response by bivalves to changing Phanerozoic sea-water chemistry. *Geological Magazine* **134**, 403-407 (1997).
- 21 Porter, S. Calcite and aragonite seas and the *de novo* acquisition of carbonate skeletons. *Geobiology* **8**, 256-277 (2010).
- 22 Demicco, R. V., Lowenstein, T. K., Hardie, L. A. & Spencer, R. J. Model of seawater composition for the Phanerozoic. *Geology* **33**, 877-880 (2005).
- 23 Veizer, J. & Prokoph, A. Temperatures and oxygen isotopic composition of Phanerozoic oceans. *Earth-Science Reviews* **146**, 92-104 (2015).
- 25 Kiessling, W., Aberhan, M. & Villier, L. Phanerozoic trends in skeletal mineralogy driven by mass extinctions. *Nature Geoscience* **1**, 527-530 (2008).
- 26 Foote, M., Crampton, J. S., Beu, A. G. & Nelson, C. S. Aragonite bias, and lack of bias, in the fossil record: lithological, environmental, and ecological controls. *Paleobiology* **41**, 245-265 (2015).
- 27 Sugihara, G. *et al.* Detecting causality in complex ecosystems. *science*, 1227079 (2012).
- 28 Knoll, A. H., Bambach, R. K., Payne, J. L., Pruss, S. & Fischer, W. W. Paleophysiology and end-Permian mass extinction. *Earth and Planetary Science Letters* **256**, 295-313 (2007).
- 51 Cherns, L. & Wright, V. P. Quantifying the impacts of early diagenetic aragonite dissolution on the fossil record. *Palaaios* **24**, 756-771 (2009).
- 52 Balthasar, U. *et al.* Relic aragonite from Ordovician–Silurian brachiopods: Implications for the evolution of calcification. *Geology* **39**, 967-970 (2011).
- 69 Alroy, J. *et al.* Phanerozoic trends in the global diversity of marine invertebrates. *Science* **321**, 97-100 (2008).
- 72 Lowenstein, T. K., Timofeeff, M. N., Brennan, S. T., Hardie, L. A. & Demicco, R. V. Oscillations in Phanerozoic seawater chemistry: Evidence from fluid inclusions. *Science* **294**, 1086-1088 (2001).
- 73 Lowenstein, T. K., Timofeeff, M. N., Kovalevych, V. M. & Horita, J. The major-ion composition of Permian seawater. *Geochimica et Cosmochimica Acta* **69**, 1701-1719 (2005).
- 74 Horita, J., Zimmermann, H. & Holland, H. D. Chemical evolution of seawater during the Phanerozoic: Implications from the record of marine evaporites. *Geochimica et Cosmochimica Acta* **66**, 3733-3756 (2002).
- 75 Brennan, S. T., Lowenstein, T. K. & Cendón, D. I. The major-ion composition of Cenozoic seawater: The past 36 million years from fluid inclusions in marine halite. *American Journal of Science* **313**, 713-775 (2013).
- 76 Coggon, R. M., Teagle, D. A., Smith-Duque, C. E., Alt, J. C. & Cooper, M. J. Reconstructing past seawater Mg/Ca and Sr/Ca from mid-ocean ridge flank calcium carbonate veins. *Science* **327**, 1114-1117 (2010).
- 77 Dickson, J. Echinoderm skeletal preservation: calcite-aragonite seas and the Mg/Ca ratio of Phanerozoic oceans. *Journal of Sedimentary Research* **74**, 355-365 (2004).
- 78 Dickson, J. Fossil echinoderms as monitor of the Mg/Ca ratio of Phanerozoic oceans. *Science* **298**, 1222-1224 (2002).
- 79 Ries, J. B. Effect of ambient Mg/Ca ratio on Mg fractionation in calcareous marine invertebrates: A record of the oceanic Mg/Ca ratio over the Phanerozoic. *Geology* **32**, 981-984 (2004).
- 80 Steuber, T. & Rauch, M. Evolution of the Mg/Ca ratio of Cretaceous seawater: implications from the composition of biological low-Mg calcite. *Marine Geology* **217**, 199-213 (2005).

- 81 Evans, D., Brierley, C., Raymo, M. E., Erez, J. & Müller, W. Planktic foraminifera shell chemistry response to seawater chemistry: Pliocene–Pleistocene seawater Mg/Ca, temperature and sea level change. *Earth and Planetary Science Letters* **438**, 139-148 (2016).
- 82 Evans, D. & Müller, W. Deep time foraminifera Mg/Ca paleothermometry: Nonlinear correction for secular change in seawater Mg/Ca. *Paleoceanography* **27** (2012).
- 83 Gorzelak, P., Stolarski, J., Mazur, M. & Meibom, A. Micro-to nanostructure and geochemistry of extant crinoidal echinoderm skeletons. *Geobiology* **11**, 29-43 (2013).
- 84 Kołbuk, D., Dubois, P., Stolarski, J. & Gorzelak, P. Effects of seawater chemistry (Mg²⁺/Ca²⁺ ratio) and diet on the skeletal Mg/Ca ratio in the common sea urchin *Paracentrotus lividus*. *Marine Environmental Research* (2019).
- 85 Gorzelak, P., Krzykowski, T. & Stolarski, J. Diagenesis of echinoderm skeletons: constraints on paleoseawater Mg/Ca reconstructions. *Global and Planetary Change* **144**, 142-157 (2016).
- 86 Berner, R. A. A model for calcium, magnesium and sulfate in seawater over Phanerozoic time. *American Journal of Science* **304**, 438-453 (2004).
- 87 Farkaš, J. *et al.* Calcium isotope record of Phanerozoic oceans: Implications for chemical evolution of seawater and its causative mechanisms. *Geochimica et Cosmochimica Acta* **71**, 5117-5134 (2007).
- 88 Arvidson, R. S., Mackenzie, F. T. & Guidry, M. W. Geologic history of seawater: A MAGic approach to carbon chemistry and ocean ventilation. *Chemical Geology* **362**, 287-304 (2013).
- 89 Gaffin, S. Ridge volume dependence on seafloor generation rate and inversion using long term sealevel change. *American Journal of Science* **287**, 596-611 (1987).
- 90 Brennan, S. T., Lowenstein, T. K. & Horita, J. Seawater chemistry and the advent of biocalcification. *Geology* **32**, 473-476 (2004).
- 91 Brennan, S. T. & Lowenstein, T. K. The major-ion composition of Silurian seawater. *Geochimica et Cosmochimica Acta* **66**, 2683-2700 (2002).
- 92 Timofeeff, M. N., Lowenstein, T. K., Da Silva, M. A. M. & Harris, N. B. Secular variation in the major-ion chemistry of seawater: Evidence from fluid inclusions in Cretaceous halites. *Geochimica et Cosmochimica Acta* **70**, 1977-1994 (2006).
- 93 Alberti, M., Fürsich, F. T., Abdelhady, A. A. & Andersen, N. Middle to Late Jurassic equatorial seawater temperatures and latitudinal temperature gradients based on stable isotopes of brachiopods and oysters from Gebel Maghara, Egypt. *Palaeogeography, palaeoclimatology, palaeoecology* **468**, 301-313 (2017).
- 94 Scotese, C. in *GSA Annual Meeting Denver, Colorado, Abstracts with Programs*. 74-31.
- 95 Chekroun, M. D., Simonnet, E. & Ghil, M. Stochastic climate dynamics: Random attractors and time-dependent invariant measures. *Physica D: Nonlinear Phenomena* **240**, 1685-1700 (2011).
- 96 Cherns, L. & Wright, V. P. Missing molluscs as evidence of large-scale, early skeletal aragonite dissolution in a Silurian sea. *Geology* **28**, 791-794 (2000).
- 97 Wright, V. P. & Cherns, L. How far did feedback between biodiversity and early diagenesis affect the nature of Early Palaeozoic sea floors? *Palaeontology* **59**, 753-765 (2016).
- 98 Hendy, A. J. The influence of lithification on Cenozoic marine biodiversity trends. *Paleobiology* **35**, 51-62 (2009).
- 99 Paul, C. R. C. in *Problems of phylogenetic reconstruction* (Academic Press, 1982).
- 100 Liow, L. H., Reitan, T. & Harnik, P. G. Ecological interactions on macroevolutionary time scales: clams and brachiopods are more than ships that pass in the night. *Ecology Letters* **18**, 1030-1039 (2015).
- 101 Carter, J. Environmental and biological controls of bivalve shell mineralogy and microstructure. *Skeletal growth of aquatic organisms* (1980).
- 102 Harper, E. M. & Checa, A. Physiological versus Biological Control in Bivalve Calcite Prisms: Comparison of Euheterodonts and Pteriomorphs. *The Biological Bulletin* **232**, 19-29 (2017).
- 103 Teichert, C. Times of crisis in the evolution of the Cephalopoda. *Paläontologische Zeitschrift* **60**, 227-243 (1986).
- 104 Von Euw, S. *et al.* Biological control of aragonite formation in stony corals. *Science* **356**, 933-938 (2017).
- 105 Zapalski, M. K. Evidence of photosymbiosis in Palaeozoic tabulate corals. *Proceedings of the Royal Society of London B: Biological Sciences* **281**, 20132663 (2014).
- 106 Ries, J. B., Stanley, S. M. & Hardie, L. A. Scleractinian corals produce calcite, and grow more slowly, in artificial Cretaceous seawater. *Geology* **34**, 525-528 (2006).
- 107 Kiessling, W., Simpson, C. & Foote, M. Reefs as cradles of evolution and sources of biodiversity in the Phanerozoic. *Science* **327**, 196-198 (2010).

- 108 Kiessling, W. Geologic and biologic controls on the evolution of reefs. *Annual Review of Ecology, Evolution, and Systematics* **40**, 173-192 (2009).
- 109 Porter, S. M. Seawater chemistry and early carbonate biomineralization. *Science* **316**, 1302-1302 (2007).
- 110 Zhuravlev, A. Y. & Wood, R. A. Eve of biomineralization: Controls on skeletal mineralogy. *Geology* **36**, 923-926 (2008).
- 111 Knoll, A. H. Biomineralization and evolutionary history. *Reviews in mineralogy and geochemistry* **54**, 329-356 (2003).
- 112 Reitner, J. A new calcitic sphinctozoan sponge belonging to the Demospongiae from the Cassian Formation (Lower Carnian; Dolomites, Northern Italy) and its phylogenetic relationship. *Geobios* **20**, 571-589 (1987).
- 113 Wray, J. L. in *Introduction to marine Micropaleontology* 171-187 (Elsevier, 1998).
- 114 Martin, R. E. Cyclic and secular variation in microfossil biomineralization: clues to the biogeochemical evolution of Phanerozoic oceans. *Global and Planetary Change* **11**, 1-23 (1995).
- 115 Smith, A. M., Key, M. M. & Gordon, D. P. Skeletal mineralogy of bryozoans: taxonomic and temporal patterns. *Earth-Science Reviews* **78**, 287-306 (2006).
- 116 Bandel, K. Shell structure of the Gastropoda excluding Archaeogastropoda. *Skeletal biomineralization: patterns, processes and evolutionary trends* **1**, 117-134 (1990).
- 117 van Dijk, I., de Nooijer, L. J., Hart, M. B. & Reichart, G. J. The long-term impact of magnesium in seawater on foraminiferal mineralogy: Mechanism and consequences. *Global Biogeochemical Cycles* **30**, 438-446 (2016).
- 118 Tappan, H. & Loeblich Jr, A. R. Foraminiferal evolution, diversification, and extinction. *Journal of Paleontology*, 695-714 (1988).
- 119 Carter, J. G., Barrera, E. & Tevesz, M. J. Thermal potentiation and mineralogical evolution in the Bivalvia (Mollusca). *Journal of Paleontology* **72**, 991-1010 (1998).
- 120 Lowenstam, H. A. Factors affecting the aragonite: calcite ratios in carbonate-secreting marine organisms. *The Journal of Geology* **62**, 284-322 (1954).
- 121 Stolarski, J., Meibom, A., Przeniosło, R. & Mazur, M. A Cretaceous scleractinian coral with a calcitic skeleton. *Science* **318**, 92-94 (2007).
- 122 De Baets, K. & Munnecke, A. Evidence for Palaeozoic orthoconic cephalopods with bimineralic shells. *Palaeontology* (2017).
- 123 Janiszewska, K., Mazur, M., Escrig, S., Meibom, A. & Stolarski, J. Aragonitic scleractinian corals in the Cretaceous calcitic sea. *Geology* **45**, 319-322 (2017).
- 124 Horton, T. *et al.* World Register of Marine Species (WoRMS). *WoRMS Editorial Board*, <http://www.marinespecies.org> (2018).
- 125 Chamberlain, S. A. & Szöcs, E. taxize: taxonomic search and retrieval in R. *F1000Research* **2**, <http://f1000research.com/articles/2-191/v2> (2013).
- 126 Gradstein, F. M., Ogg, J. G., Schmitz, M. & Ogg, G. *The geologic time scale 2012*. (elsevier, 2012).
- 127 Cohen, K., Finney, S., Gibbard, P. & Fan, J.-X. The ICS international chronostratigraphic chart. *Episodes* **36**, 199-204 (2013, updated).