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# Genomic Dissection of Bipolar Disorder and Schizophrenia, Including 28 Subphenotypes

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# Genomic dissection of bipolar disorder and schizophrenia including 28 subphenotypes

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## Summary

Schizophrenia and bipolar disorder are two distinct diagnoses that share symptomology. Understanding the genetic factors contributing to the shared and disorder-specific symptoms will be crucial for improving diagnosis and treatment. In genetic data consisting of 53,555 cases (20,129 BD, 33,426 SCZ) and 54,065 controls, we identified 114 genome-wide significant loci implicating synaptic and neuronal pathways shared between disorders. Comparing SCZ to BD (23,585 SCZ, 15,270 BD) identified four genomic regions including one with disorder-independent causal variants and potassium ion response genes as contributing to differences in biology between the disorders. Polygenic risk score (PRS) analyses identified several significant correlations within case-only phenotypes including SCZ PRS with psychotic features and age of onset in BD. For the first time, we discover specific loci that distinguish between BD and SCZ and identify polygenic components underlying multiple symptom dimensions. These results point to the utility of genetics to inform symptomology and potentially treatment.

## Introduction

Bipolar disorder (BD) and schizophrenia (SCZ) are severe psychiatric disorders and among the leading causes of disability worldwide(Whiteford et al., 2013). Both disorders have significant genetic components with heritability estimates ranging from 60-80%(Nöthen et al., 2010). Recent genetic and epidemiological studies have demonstrated substantial overlap between these two disorders with a genetic correlation from common variation near 0.6-0.7(Cross-Disorder Group of the Psychiatric Genomics Consortium, 2013) and high relative risks (RR) among relatives of both BD and SCZ patients (RRs for parent/offspring: BD/BD: 6.4, BD/SCZ: 2.4;

SCZ/BD: 5.2, SCZ/SCZ: 9.9)(Lichtenstein et al., 2009). Despite shared genetics and symptomology, the current diagnostic systems(“Diagnostic and Statistical Manual of Mental Disorders | DSM Library,” n.d.)(“WHO | International Classification of Diseases,” n.d.) adhere to historical distinctions from the late 19<sup>th</sup> century and represent BD and SCZ as independent categorical entities differentiated on the basis of their clinical presentation, with BD characterized by predominant mood symptoms, mood-congruent delusions and an episodic disease course and SCZ considered a prototypical psychotic disorder. Identifying genetic components contributing to both disorders provides insight into the biology underlying the shared symptoms of the disorders.

While the shared genetic component is substantial, studies to date have also implicated genetic architecture differences between these two disorders(Curtis et al., 2011; Ruderfer et al., 2014). A polygenic risk score created from a case only SCZ vs BD genome-wide association study (GWAS) significantly correlated with SCZ or BD diagnosis in an independent sample(Ruderfer et al., 2014), providing the first evidence that differences between the disorders also have a genetic basis. An enrichment of rare, moderate to highly penetrant copy number variants (CNVs) and *de novo* CNVs are seen in SCZ patients(CNV and Schizophrenia Working Groups of the Psychiatric Genomics Consortium, 2017; Gulsuner and McClellan, 2015; Kirov et al., 2012; Stone et al., 2008; Szatkiewicz et al., 2014), while, the involvement of CNVs in BD is less clear(Green et al., 2016). Although the role of *de novo* single nucleotide variants in BD and SCZ has been investigated in only a handful of studies, enrichment in pathways associated with the postsynaptic density has been reported for SCZ, but not BD(Fromer et al., 2014; Kataoka et al., 2016). Identifying disorder-specific variants and quantifying the contribution of genetic variation to specific symptom dimensions remain important open questions. Characterizing these genetic

differences will facilitate an understanding of the dimensions of the disorders instead of the dichotomous diagnosis. For example, we have shown that SCZ patients with greater manic symptoms have higher polygenic risk for BD (Ruderfer et al., 2014). These findings demonstrate shared genetic underpinnings for symptoms across disorders and may enable us to characterize patients by genetic liability to symptom dimensions thereby informing disease course and treatment.

Here, we utilize large collections of genotyped samples for BD and SCZ along with clinically-relevant measures identifying 28 subphenotypes to address three questions: 1) Are there specific variants, genes or pathways that are either shared by, or differentiate BD and SCZ? 2) Are the shared symptoms between these disorders driven by the same underlying genetic profiles? and 3) Can we demonstrate independent genetic signatures for subphenotypes within these disorders?

## **Results**

### **Shared genetic contribution to BD and SCZ**

We performed association analysis of BD and SCZ combined into a single phenotype, totaling 53,555 cases (20,129 BD, 33,426 SCZ) and 54,065 controls on 15.5 million SNP allele dosages imputed from 1000 genomes phase 3 (The 1000 Genomes Project Consortium, 2015). Logistic regression was performed controlling for 13 principal components of ancestry, study sites and genotyping platform. We identified 11,231 SNPs with p-value below our genome-wide significance (GWS) threshold of  $5 \times 10^{-8}$ . After grouping SNPs in linkage disequilibrium with each other ( $r^2 > 0.2$ ), 114 genomic risk loci remained. For the most significant variant in each of the 114 GWS loci, we performed conditional analysis with any GWS hit within 1Mb of the

extent of the locus from the previously performed single disease GWAS of SCZ(Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014) and BD(Stahl et al., 2017) and identified 32 loci that were independently significant defined strictly as no single disease locus within 1Mb or a GWS p-value after conditional analysis (Supplementary Table 1). We further performed gene-set based tests using MAGMA(Leeuw et al., 2015) across 10,891 curated pathways(Watanabe et al., 2017) and identified 8 pathways surpassing Bonferroni correction ( $p < 4.6 \times 10^{-6}$ ) with all but one pathway implicating synaptic and neuronal biology (Supplementary Table 2a). Establishing independent controls (see Methods) allowed us to perform disorder-specific GWAS in 20,129 BD cases vs 21,524 BD controls and 33,426 SCZ cases and 32,541 SCZ controls. Using these results, we compared effect sizes of these 114 loci across each disorder independently showing the subsets of variants that had larger effects in SCZ compared to BD and vice versa (Figure 1a).

### **Differentiating genetic contribution to BD and SCZ**

To identify loci with divergent effects on BD and SCZ, we performed an association analysis comparing 23,585 SCZ cases with 15,270 BD cases matched for shared ancestry and genotyping platform (see Methods, Figure 1b, Table 1). Two genome-wide significant loci were identified, the most significant of which was rs56355601 located on chromosome 1 at position 173,811,455 within an intron of *DARS2* (Supplementary Figure 1). The second most significant locus was rs200005157, a four base-pair insertion/deletion, on chromosome 20 at position 47638976 in an intron of *ARFGEF2* (Supplementary Figure 2). For both variants, the minor allele frequency was higher in BD cases than SCZ cases and disease-specific GWAS showed opposite directions of effect when compared to controls. We sought to identify additional disease-specific loci by

comprehensively incorporating expression information with association results to perform fine-mapping and identify novel variants (Gamazon et al., 2015; Giambartolomei et al., 2014; Gusev et al., 2016; He et al., 2013). Here, we applied the summary-data-based Mendelian randomization (SMR) method (Zhu et al., 2016) (see Methods) utilizing the cis-QTLs derived from peripheral blood (Westra et al., 2013), human dorsolateral prefrontal cortex (DLPFC) (Fromer et al., 2016) from the Common Mind Consortium and 11 brain regions from the GTEx consortium (Consortium, 2015). We identified one SNP-probe combination that surpassed the threshold for genome-wide significance in blood but was also the most significant finding in brain. We found that SNP rs4793172 in gene *DCAKD* is associated with SCZ vs BD analysis ( $p_{\text{GWAS}} = 2.8 \times 10^{-6}$ ) and is an eQTL for probe ILMN 1811648 ( $p_{\text{eQTL}} = 2.9 \times 10^{-168}$ ), resulting in  $p_{\text{SMR}} = 4.1 \times 10^{-6}$  in blood ( $p_{\text{eQTL}} = 2.9 \times 10^{-25}$ ,  $p_{\text{SMR}} = 2.0 \times 10^{-5}$  in DLFC, and  $p_{\text{eQTL}} = 4.6 \times 10^{-15}$ ,  $p_{\text{SMR}} = 6.0 \times 10^{-5}$  in GTEx cerebellar hemisphere) (Supplementary Table 3, Supplementary Figure 3) and shows no evidence of heterogeneity ( $p_{\text{HET}} = 0.66$ ) which implies only a single causal variant in the locus.

In an effort to prioritize genes for the two GWS loci from the GWAS, we performed fine-mapping (Benner et al., 2016) using an LD map derived from a majority of the control samples. We then performed SMR on each of the variants with causal probability greater than 1% using all eQTLs from the CommonMind Consortium DLPFC reference. All the most likely causal variants were shown to most significantly regulate the same gene suggesting *CSEIL* is the most likely relevant gene on chromosome 20 (rs200005157: causal probability=0.21,  $p_{\text{GWAS}} = 2.4 \times 10^{-8}$ ,  $p_{\text{eQTL}} = 3 \times 10^{-8}$ ,  $p_{\text{SMR}} = 8.5 \times 10^{-5}$ ,  $p_{\text{HET}} = 0.34$ ). For the locus on chromosome 1, *SLC9C2* is the most significantly regulated gene. However, a highly significant heterogeneity test indicates a complex genetic architecture making it difficult to infer a causal role for the associated SNP.



Therefore, *DARS2* presents as the most likely relevant gene on chromosome 1 (rs56355601:  $p_{\text{GWAS}}=5.6 \times 10^{-9}$ , causal probability=0.079,  $p_{\text{eQTL}} 7.4 \times 10^{-13}$ ,  $p_{\text{SMR}}=6.17 \times 10^{-6}$ ,  $p_{\text{HET}}=0.03$ ). We note however, that in both cases there are less associated variants that are stronger eQTLs for these genes complicating a straightforward causal interpretation. Finally, using the same gene-set test used for the combined analysis GO biological process “response to potassium ion” ( $p=1.6 \times 10^{-6}$ ) was the only pathway surpassing our Bonferroni corrected significance threshold (Supplementary Table 2b).

### **Regional joint association**

We expanded our efforts to identify disorder-specific genomic regions by jointly analyzing independent GWAS results from BD and SCZ (Pickrell et al., 2016). The genome was split into 1,703 previously defined approximately LD independent regions (Berisa and Pickrell, 2015). Thirteen percent, or 223 regions, had a posterior probability greater than 0.5 of having a causal variant for at least one disorder. Of these, 132 best fit the model of a shared causal variant influencing both BD and SCZ, 88 were most likely specific to SCZ, 3 demonstrated evidence of two independent variants (with one impacting each of the two disorders) and none were BD-specific. Of note, this approach calculates a prior probability that any given region is disease-specific and from these data the probability of having a BD specific region was 0.1% compared to 15% for SCZ, likely a result of increased power from the larger SCZ sample size and/or a difference in genetic architecture between these disorders.

The 114 GWS SNPs from the combined BD and SCZ GWAS localized into 99 independent regions (13 regions had multiple GWS SNPs), of which 78 (79%) were shared with a posterior probability of greater than 0.5. Sixty regions had at least one GWS SNP in the independent SCZ

GWAS, of which 30 (50%) are shared and 8 regions contained a GWS SNP in the independent BD GWAS, of which 6 (75%) are shared using the same definition. For the three regions showing evidence for independent variants, two had highly non-overlapping association signals in the same region stemming from independent variants. The third, on chromosome 19 presented a different scenario where association signals were overlapping. The most significant variant in BD was rs111444407 (chr19:19358207,  $p = 8.67 \times 10^{-10}$ ) and for SCZ was rs2315283 (chr19:19480575,  $p=4.41 \times 10^{-7}$ ). After conditioning on the most significant variant in the other disorder, the association signals of the most significant variant in BD and SCZ were largely unchanged (BD rs111444407  $=1.3 \times 10^{-9}$ , SCZ rs2315283  $p=6.7 \times 10^{-5}$ ). We further calculated the probability of each variant in the region being causal for both BD and SCZ (Benner et al., 2016) and found no correlation ( $r= -0.00016$ ). The most significant variants had the highest posterior probability of being causal (SCZ: rs2315283, prob = 0.02, BD: rs111444407, prob = 0.16). Both variants most significantly regulate the expression of *GATAD2A* in brain (Fromer et al., 2016) but in opposite directions (rs111444407  $p_{eQTL} = 6 \times 10^{-15}$ ,  $\beta = 0.105$ ; rs2315283  $p_{eQTL} = 1.5 \times 10^{-28}$ ,  $\beta = -0.11$ ).

### **Regional SNP-heritability estimation**

Across the genome, regional SNP-heritabilities ( $h^2_{\text{snp}}$ ) were estimated separately for SCZ and BD (Shi et al., 2016) and were found to be moderately correlated ( $r=0.25$ ). We next defined risk regions as those containing the most associated SNP for each GWS locus. In total, there were 101 SCZ risk regions from the 105 autosomal GWS loci reported previously (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014) and 29 BD risk regions from 30 GWS loci reported previously (Stahl et al., 2017). Ten regions were risk regions for both BD and

SCZ comprising 33% of BD risk regions and 10% of SCZ risk regions. We further stratified regional  $h^2_{\text{snp}}$  by whether a region was a risk region in one disorder, none or both (Supplementary Figure 4). Since the discovery data for the regions overlapped with the data used for the heritability estimation, we expected within-disorder analyses to show significant results. In risk regions specific to SCZ (n=91) there was a significant increase in regional  $h^2_{\text{snp}}$  in SCZ, as expected ( $p = 1.1 \times 10^{-22}$ ), but also in BD ( $p = 1.2 \times 10^{-6}$ ). In risk regions specific to BD (n=19), significantly increased regional  $h^2_{\text{snp}}$  was observed in BD, as expected ( $p = 0.0007$ ), but not in SCZ ( $p = 0.89$ ). Risk regions shared by both disorders had significantly higher  $h^2_{\text{snp}}$  in both disorders, as expected (BD  $p = 5.3 \times 10^{-5}$ , SCZ  $p = 0.006$ ), compared to non-risk regions. However, we observed a significant increase in BD  $h^2_{\text{snp}}$  in shared risk regions compared to BD risk regions (BD  $p = 0.003$ ) but not SCZ  $h^2_{\text{snp}}$  for shared risk regions compared to SCZ risk regions ( $p = 0.62$ ). Using a less stringent p-value threshold for defining risk regions ( $p < 5 \times 10^{-6}$ ), thereby substantially increasing the number of regions, resulted in similar results. Seven regions contributed to substantially higher  $h^2_{\text{snp}}$  in SCZ compared to BD but no region showed the inverse pattern. Of these regions, all but one was in the major histocompatibility region (MHC), the sole novel region was chr10:104380410-106695047 with regional  $h^2_{\text{snp}} = 0.0019$  in SCZ and  $h^2_{\text{snp}} = 0.00063$  in BD.

## **Polygenic dissection of subphenotypes**

Subphenotypes were collected for a subset of patients with either BD or SCZ (see Methods). For SCZ, we had clinical quantitative measurements of manic, depressive, positive and negative symptoms generated from factor analysis of multiple instruments as described previously (Ruderfer et al., 2014) but in larger sample sizes (n=6908, 6907, 8259, 8355

respectively). For BD, 24 subphenotypes were collected among nearly 13,000 cases in distinct categories including comorbidities, clinical information such as rapid cycling and psychotic features as well as additional disease course data such as age of onset and number of hospitalizations. For each BD or SCZ patient, we calculated a polygenic risk score (PRS) using all SNPs, from each of the four main GWAS analyses (BD+SCZ, BD, SCZ and SCZvsBD). We then used regression analysis including principal components and site to assess the relationship between each subphenotype and the 4 PRS. Specifically, we tested whether polygenic risk scores of BD+SCZ, BD, SCZ or SCZvsBD were correlated with each of these subphenotypes separately within BD and SCZ cases. When testing if the variance explained by the PRS was different from zero, we applied a significance cutoff of  $p < 0.0004$  based on Bonferroni correction for 112 tests. In total, we identified 6 significant results after correction (Figure 2, Table 2).

A significant positive correlation existed between BD PRS and manic symptoms in SCZ cases as seen previously (Ruderfer et al., 2014) ( $p=2 \times 10^{-5}$ ,  $t=4.26$ ) and BD PRS and psychotic features in BD patients ( $p=5.3 \times 10^{-5}$ ,  $t=4.04$ ). A significant increase in SCZ PRS was seen for BD cases with versus without psychotic features ( $p=1.2 \times 10^{-10}$ ,  $t=6.45$ ) and patients with increased negative symptoms in SCZ patients ( $p=3.60 \times 10^{-6}$ ,  $t=4.64$ ). The BD+SCZ vs controls PRS was significantly associated with psychotic features in BD ( $p=7.9 \times 10^{-13}$ ,  $t=7.17$ ) and negative symptoms in SCZ ( $p=1.5 \times 10^{-5}$ ,  $t=4.33$ ). The next two most significant results which did not survive our conservative correction were both indicative of a more severe course in BD: increased BD+SCZ PRS with increased numbers of hospitalizations in BD cases ( $p=4.2 \times 10^{-4}$ ,  $t=3.53$ ) and increased SCZ PRS with earlier onset of BD ( $p=7.9 \times 10^{-4}$ ,  $t=-3.36$ ). We assessed the role of BD subtype on the correlation between SCZ PRS and psychotic features and identified a

significant correlation when restricted to only BD type I cases indicating the result was not likely driven by BD patients with a schizoaffective subtype (BDI: 3,763 with psychosis, 2,629 without,  $p=1.55 \times 10^{-5}$ , Supplementary Table 4).

We performed a GWAS for all 8 quantitative subphenotypes and 9 binary subphenotypes with at least 1,000 cases and calculated heritability and genetic correlation with BD and SCZ. Only two subphenotypes had significant  $h^2_{\text{snp}}$  estimates using LD-score regression (Bulik-Sullivan et al., 2015) both in BD: psychotic features in BD ( $h^2_{\text{snp}}=0.15$ ,  $SE=0.06$ ) and suicide attempt ( $h^2_{\text{snp}}=0.25$ ,  $SE=0.1$ ). Only psychotic features demonstrated a significant genetic correlation with SCZ ( $r_g=0.34$ ,  $SE=0.13$ ,  $p=0.009$ ). The significant genetic correlation demonstrates a genome-wide relationship between common variants contributing to SCZ risk and those contributing to psychotic features in BD cases. We tested whether the most significantly associated SCZ loci contributed directly to psychotic features in BD. One hundred of the 105 autosomal genome-wide significant SCZ SNPs previously published (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014) were in our dataset after QC and 60 were in the same direction of effect for risk of psychotic features in BD ( $p=0.028$ , one-sided binomial-test).

## Discussion

Here we present a genetic dissection of bipolar disorder and schizophrenia from over 100,000 genotyped subjects. Consistent with earlier results (Cross-Disorder Group of the Psychiatric Genomics Consortium, 2013), we found extensive genetic sharing between these two disorders, identifying 114 genome-wide significant loci contributing to both disorders of which 32 are novel. These findings point to the relevance of neuronal and synaptic biology for the shared

genetic substrate of these disorders. However, despite this degree of sharing, we identified several loci that significantly differentiated between the two disorders, having opposite directions of effect. We also found polygenic components that significantly correlated from one disorder to symptoms of the other.

Two GWS loci were identified from the case only SCZ versus BD analysis providing opportunities to inform the underlying biological distinctions between BD and SCZ. The most significant locus implicates *DARS2* (coding for the mitochondrial Aspartate-tRNA ligase) which is highly expressed in the brain and significantly regulated by the most significant SNP rs56355601 ( $p_{eQTL}=2.5 \times 10^{-11}$ ). Homozygous mutations in *DARS2* are responsible for leukoencephalopathy with brainstem and spinal cord involvement and lactate elevation (LBSL), which was characterized by neurological symptoms such as psychomotor developmental delay, cerebellar ataxia and delayed mental development (Yamashita et al., 2013, p. 2). Based on methylation analysis from the prefrontal cortex of stress models (rats and monkeys) and from peripheral samples (in monkeys and human newborns), *DARS2*, among others, has been suggested as a potential molecular marker of early-life stress and vulnerability to psychiatric disorders (Luoni et al., 2016). The second most significant locus implicates *CSE1L*, a nuclear transport factor that plays a role in cellular proliferation as well as in apoptosis (Bera et al., 2001). Intronic SNPs in *CSE1L* have been associated with subjective well-being (Okbay et al., 2016) and, nominally to antidepressant response (Li et al., 2016). More interestingly, *CSE1L* is a potential target gene of miR-137, one of the well-known schizophrenia risk loci (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014), which is able to negatively regulate *CSE1L* by interacting with complementary sequences in the 3' UTR of *CSE1L* (Li et al.,

2013). Although falling short of genome-wide significance, the third most significant locus implicates *ARNTL* (Aryl Hydrocarbon Receptor Nuclear Translocator Like), which is a core component of the circadian clock. *ARNTL* has been previously hypothesized for relevance in bipolar disorder,(Yang et al., 2008) although human genetic evidence is currently limited(Byrne et al., 2014).

The ability to generate transcriptional data on multiple tissues across many individuals using RNA-sequencing has provided detailed information on the role common variants play in regulating expression of specific genes in specific tissues. These eQTLs can be integrated with the genetic association data from GWAS to inform on the relationship between variant association and variant regulation of expression for each gene. Performing this integration, we identified a third genome-wide significant finding in *DCAKD*. The gene codes for Dephospho-CoA Kinase Domain Containing protein, a member of the human postsynaptic density proteome from human neocortex(Bayés et al., 2011). In the mouse cortical synaptoproteome *DCAKD* is among the proteins with the highest changes between juvenile postnatal days and adult stage, suggesting a putative role in brain development(Gonzalez-Lozano et al., 2016; Moczulska et al., 2014). Discerning between pleiotropy (variant independently regulates expression and alters risk to disease) from causality (variant regulates expression which thereby alters risk to disease) through statistical analysis alone is difficult, this analytical approach is stringent in excluding loci where colocalised SNP-phenotype and SNP-expression associations may reflect confounding driven by linkage disequilibrium (LD) (one variant regulates expression and a different variant alters risk but the variants in the region are in LD). Hence, this approach utilizes currently available data to prioritize genes, including direction of effect, for functional follow-up.

These analyses will become more powered with increased sample sizes for both phenotype and eQTL data sets.

Performing pathway analysis based on the full association results shows enrichment of genes involved in response to potassium ions, including potassium voltage-gated channel subfamily members and a number of genes regulated by cellular potassium concentration. This is in line with previous genetic evidence pointing to a key etiologic role of potassium channels, in particular, in BD(Judy and Zandi, 2013), which could be explained by their role in multiple neurobiological mechanisms involved in the development of psychiatric disorders such as regulation of the dopaminergic circuits, synaptic plasticity, and myelination(Balaraman et al., 2015).

We further assessed the contribution of regions of the genome to each disorder through joint regional association and heritability estimation. These results point to an additional locus that may contribute differentially to liability to BD and SCZ. The region on chr19 shows overlapping association peaks that are driven by independent causal variants for each disorder. Both variants significantly regulate the same gene *GATAD2A* but in opposite directions. *GATAD2A* is a transcriptional repressor, which is targeted by *MBD2* and is involved in methylation-dependent gene silencing. The protein is part of the large NuRD (nucleosome remodeling and deacetylase) complex, for which also HDAC1/2 are essential components. NurD complex proteins have been associated with autism(Li et al., 2015). Their members, including *GATAD2A*, display preferential expression in fetal brain development(Li et al., 2015) and in recent work has been implicated in SCZ through open chromatin(Fullard et al., n.d.). Further, p66 $\alpha$  (mouse *GATAD2A*) was recently



shown to participate in memory preservation through long-lasting histone modification in hippocampal memory-activated neurons(Ding et al., 2017). SNP-heritability appears to be consistently shared across regions and chromosomes between these two disorders. Regions with GWS loci often explain higher proportions of heritability as expected. When looking at the effect on heritability of the presence of a GWS locus in the other disorder, we identified a significant increase in BD heritability for regions containing a GWS locus for SCZ but no significant increase in SCZ heritability in regions having a BD one. This result suggests a directionality to the genetic sharing of these disorders with a larger proportion of BD loci being specific to BD. However, we cannot exclude that the asymmetry of results may reflect less power of discovery for BD than SCZ. The degree to which power and subphenotypes contribute to this result requires further examination.

We note that as with nearly all GWAS findings, the calculated population-based effect sizes of the variants identified here are small and independently explain only a modest fraction to the heritability of these disorders. The identification of these variants is dependent on the ability to have highly accurate allele frequency estimates that can only be ascertained from large sample sizes. As sample sizes get larger the power to identify variants of smaller effect increases meaning that increasing sample size results in the identification of variants of smaller effect. However, a small population effect size does not exclude the possibility of a substantially larger effect on molecular phenotypes nor does it preclude the utility of association regions in understanding biology or having a clinical impact. Efforts following up GWAS results to date have demonstrated the value of these findings in pointing to genes that can aid in understanding the underlying biology of the trait(Claussnitzer et al., 2015; Mohanan et al., 2018; Sekar et al.,

2016). Further, there is a clear relationship between GWAS results of a phenotype and gene targets of drugs that treat that phenotype pointing to the potential for improved therapeutic understanding(Nelson et al., 2015; Ruderfer et al., 2016). A major challenge of GWAS is the sheer number of findings and the substantial time/cost required for functional follow up of these findings in the classical paradigms used for genes causal for monogenic disorders. In silico bioinformatic analyses (such as SMR used here) that integrate GWAS results with ‘omics data (transcription, protein, epigenetic, etc.) have the potential to put a clearer biological focus on GWAS results. Such analyses can become more complex as more reference omics data sets (with genome-wide genotyping) become available. Additional analytical efforts will be required to facilitate the transition from GWAS to biology but substantial data has shown there is much to be learned from these variants despite their small effects(Visscher et al., 2017).

We have now identified multiple genomic signatures that correlate between one disorder and a clinical symptom in the other disorder, illustrating genetic components underlying particular symptom dimensions within these disorders. Medical symptoms, including those seen in psychiatric disorders, can manifest through a multitude of causes. The classic example often used is headache for which many different paths lead to the same symptom. Psychiatric symptoms also have many potential causes. For example, symptoms of psychosis can be the result of highly heritable diseases such as BD and SCZ but also infectious and neurodegenerative diseases, sleep/sensory deprivation or psychedelic drugs. Demonstrating a shared biological underpinning to these symptoms suggests they could be treated through modulating the same pathway. As previously shown, we find a significant positive correlation between the PRS of BD and manic symptoms in SCZ. We also demonstrate that BD cases with psychotic features carry a

significantly higher SCZ PRS than BD cases without psychotic features and this result is not driven by the schizoaffective BD subtype. Further, we show that increased PRS is associated with more severe illness. This is true for BD with psychotic features having increased SCZ PRS, earlier onset BD having higher SCZ PRS and cases with higher BD+SCZ PRS having a larger number of hospitalizations. We demonstrated that psychotic features within BD is a heritable trait and GWS loci for SCZ have a consistent direction of effect in psychotic features in BD, demonstrating the potential to study psychosis more directly to identify variants contributing to that symptom dimension.

This work illustrates the utility of genetic data, in aggregate, at dissecting symptom heterogeneity among related disorders and suggests that further work could aid in characterizing patients for more personalized treatment. Genetic risk scores have demonstrated their ability to inform and predict pathology (Cleyne et al., 2016) and more recently have been shown to be able to identify patients with risk equivalent to monogenic variants (Khera et al., 2017). In psychiatry, we lack objective biological measurements (biomarkers) with which to assess the ability of a genetic signature to predict or inform. Lacking diagnostic pathology for psychiatric disorders leaves a genuine opportunity for the genetics to drive diagnosis and treatment to a much larger degree than in other domains. One potential model assumes that each individual has a quantitative loading of a series of symptom dimensions (i.e. manic, psychotic, cognitive, etc.) and that these symptoms can be assessed at the genetic level to characterize a patient's dysfunction and used to inform disease course and optimal treatment. Making this a reality will require more detailed information on disease course and outcomes. For example, if treatment response data existed for these samples one could ask whether a genetic loading for psychosis

was correlated with response to treatment. Initial work has already shown the potential of this approach using a SCZ PRS to inform lithium response in BD (Amare et al., 2018). Ultimately, the goal will be to quantify multiple genetic loadings of each individual's illness and use those measures to inform treatment based on the outcomes of previous individuals with similar profiles.

In conclusion, we present a detailed genetic dissection of BD and SCZ pointing to substantial shared genetic risk but also demonstrating that specific loci contribute to the phenotypic differences of these disorders. We show that genetic risk scores can correspond to symptoms within and across disorders. Finally, we present data that points to these disorders being neither independent nor the same but sharing particular symptom dimensions that can be captured from the genetics and used to characterize patients to ultimately inform diagnosis and treatment.

#### **Author Contributions:**

DMR, PS and KSK managed and organized the group. DMR, SR, JB, EAS, JMWP, NM, AWC, APSO, LMOL and VT contributed to analyses. Subphenotype collection and organization was led by AM and AHF. Initial manuscript was drafted by DMR, ED, ADF, SP, JLK. Manuscript contributions and interpretation of results was provided by DMR, ED, SHL, MCO, PFS, RAO, NRW, PS and KSK. The remaining authors contributed to the recruitment, genotyping, or data processing for the contributing components of the study. All other authors saw, had the opportunity to comment on, and approved the final draft.

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## Declaration of Interests

The authors declare no competing interests.

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## Figure Legends

### Figure 1. Associated Genomic Loci Shared and Divergent Between BD and SCZ

a) Odds ratios (OR) from independent data sets of BD (blue) and SCZ (red) for each of the 114 genome-wide significant variants in the BD and SCZ vs controls GWAS. b) Manhattan plot for SCZ vs BD GWAS.

### Figure 2. Polygenic Risk Score Dissection of Clinical Symptom Dimensions

Effect size (calculated by dividing regression estimate by standard error) from regression analysis including ancestry covariates for each subphenotype and PRS for BD (x-axis) and SCZ (y-axis). Point size represents  $-\log_{10}(\text{p-value})$  with SCZ (red) and BD (blue). Numbered subphenotypes are 1) comorbid migraine, 2) panic attacks 3) suicide attempt 4) mixed states 5) rapid cycling 6) comorbid eating disorder 7) comorbid OCD 8) year of birth 9) suicide ideation 10) panic disorder 11) number of suicide attempts 12) depressive symptoms (SCZ) 13) episodes depressive 14) episodes total 15) positive symptoms (SCZ) 16) irritable mania 17) age of onset depression 18) family history 19) episodes mixed mania 20) unipolar mania 21) alcohol substance dependence 22) age of onset mania 23) age at interview 24) number of hospitalizations. All subphenotypes are in BD except those labeled (SCZ).

## **Table Legends**

### **Table 1. Most Significant Associated Loci from SCZ vs BD GWAS**

Association results for the five most significant variants in the SCZ vs BD GWAS with the top two being genome-wide significant. Each variant includes results from the independent BD vs controls and SCZ vs controls GWAS and the comparable p-value from a heterogeneity test when performing a two cohort meta-analysis of SCZ and BD.

### **Table 2. Complete Results of Polygenic Risk Score Dissection Analysis**

Polygenic scoring results of all four GWAS phenotypes (BD+SCZ vs controls, BD vs controls, SCZ vs controls and SCZ vs BD) and 24 subphenotypes from BD and 4 subphenotypes from SCZ, rows without case/control counts are quantitative measures. Significance and effects are from regression analysis of subphenotype on PRS including principal components of ancestry and site as covariates. Effect is the regression estimate divided by the standard error.

## **Supplementary Figure Legends**

**Figure S1. Related to Figure 1b. Regional Association Plot and Forest Plot for the First Genome-wide Significant Hit in the SCZ vs BD GWAS.**

**Figure S2. Related to Figure 1b. Regional Association Plot and Forest Plot for the Second Genome-wide Significant Hit in the SCZ vs BD GWAS.**

**Figure S3. Related to Summary-data-based Mendelian Randomization. Detailed Association of DCAKD from SMR.**

Results at the *DCAKD* locus from SMR analysis of SCZ vs BD. Top plot, brown dots represent the *P* values for SNPs from SCZ vs BD GWAS, diamonds represent the *P* values for probes from the SMR test. Bottom plot, the eQTL *P* values of SNPs from the Westra study for the ILMN\_1811648 probe tagging *DCAKD*. The top and bottom plots include all the SNPs available in the region in the GWAS and eQTL summary data, respectively, rather than only the SNPs common to both data sets. Highlighted in red is the gene (*DCAKD*) that passed the SMR and HEIDI tests.

**Figure S4. Related to Regional SNP-heritability estimation. Heritability Estimates for BD and SCZ in Genome-wide Significant Regions of BD and SCZ.**

Regional SNP-heritability estimates for SCZ and BD stratified by whether the region contains the most significant variant in a genome-wide significant locus in BD, SCZ, neither or both.

**STAR Methods**

**CONTACT FOR REAGENT AND RESOURCE SHARING**

Genotype and phenotype data use is restricted and governed by the Psychiatric Genetics Consortium. Further information and requests for analytical results or additional information should be directed to and will be fulfilled by the Lead Contact, Douglas Ruderfer ([douglas.ruderfer@vanderbilt.edu](mailto:douglas.ruderfer@vanderbilt.edu)).



## **SUBJECT DETAILS**

### **Genotyped Sample Description**

SCZ samples are a substantial subset of those analyzed previously (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014). BD samples are the newest collection from Psychiatric Genomics Consortium Bipolar Disorder Working Group (Stahl et al., 2017).

Below we provide information on the individual samples used here as provided by the original PGC disorder publications. Additionally, most studies have been described in detail in the citations provided. The boldfaced first line for each sample is study PI, PubMed ID, country (study name), and the PGC internal tag or study identifier.

### **European ancestry, case-control design**

#### *Schizophrenia*

**Adolfsson, R | NP | Umeå, Sweden | scz\_umeb\_eur**

**Adolfsson, R | NP | Umeå, Sweden | scz\_umes\_eur**

Cases of European ancestry were ascertained from multiple different studies of schizophrenia (1992-2009). The diagnostic processes were similar between studies, and the final diagnosis is a best-estimate consensus lifetime diagnosis based on multiple sources of information such as clinical evaluation by research psychiatrists, different types of semi-structured interviews made by trained research nurses and research psychiatrists, medical records, course of the disease and data from multiple informants. Diagnosis was made in accordance with the Diagnostic and Statistical Manual of Mental Disorders-Version IV (DSM-IV) or International Classification of Diseases, 10th Revision (ICD-10) criteria. Controls were recruited from the Betula study, an ongoing longitudinal, prospective, population-based study from the same geographic area (North

1004 Sweden) that is studying aging, health, and cognition in adults. All subjects (cases and controls)  
1005 participated after giving written informed consent and the regional Ethical Review Board at the  
1006 University of Umeå approved all original studies and participation in the PGC. GWAS  
1007 genotyping was performed at Broad Institute.

1008 **Andreassen, O | 19571808 | Norway (TOP) | scz\_top8\_eur**

1009 In the TOP study (Tematisk område psykoser), cases of European ancestry, born in Norway,  
1010 were recruited from psychiatric hospitals in the Oslo region. Patients were diagnosed according  
1011 to SCID and further ascertainment details have been reported. Healthy control subjects were  
1012 randomly selected from statistical records of persons from the same catchment area as the patient  
1013 groups. All participants provided written informed consent and the human subjects protocol was  
1014 approved by the Norwegian Scientific-Ethical Committee and the Norwegian Data Protection  
1015 Agency.

1016 **Blackwood, D | 19571811 | Edinburgh, UK | scz\_edin\_eur**

1017 Cases and controls were recruited from the southeast of Scotland, and ascertainment has been  
1018 previously described as part of the International Schizophrenia Consortium studies. All  
1019 participating subjects gave written, informed consent and the human subjects protocol was  
1020 approved by the Scotland A Research Ethics Committee. DNA samples were genotyped at the  
1021 Broad Institute.

1022 **Børglum, A | 19571808 | Denmark | scz\_aarh\_eur**

1023 DNA samples for all subjects were collected from blood spots systematically collected by the  
1024 Danish Newborn Screening Biobank), with case/control status established using the Danish  
1025 Psychiatric Central Register. Cases were diagnosed clinically according to ICD-10 criteria.

1026 Controls were selected to match the cases by birth cohort. The Danish Data Protection Agency  
1027 and the ethics committees in Denmark approved the human subjects protocol.

1028 **Bramon | 23871474 | Seven countries (PEIC, WTCCC2) | scz\_pewb\_eur**

1029 **Bramon | 23871474 | Spain (PEIC, WTCCC2) | scz\_pewb\_eur**

1030 The Psychosis Endophenotypes International Consortium (PEIC) was part of WTCCC2. Samples  
1031 were collected through seven centers in Europe and Australia (the Institute of Psychiatry, King's  
1032 College London, London; GROUP (consisting of the University of Amsterdam, Amsterdam; the  
1033 University of Groningen, Groningen; Maastricht University Medical Centre, Maastricht; and the  
1034 University of Utrecht, Utrecht); the University of Western Australia, Perth; the Universidad de  
1035 Cantabria, Santander; the University of Edinburgh, Edinburgh; Heidelberg University,  
1036 Heidelberg and Ludwig-Maximilians-Universität München, Munich). To allow for a DSM-IV  
1037 diagnosis to be ascertained or ruled out, all participants (including controls and unaffected family  
1038 members) underwent a structured clinical interview with the Schedule for Affective Disorders  
1039 and Schizophrenia (SADS), the Structured Clinical Interview for DSM Disorders (SCID), or the  
1040 Schedules for Clinical Assessment in Neuropsychiatry (SCAN). We included cases with  
1041 schizophrenia and schizoaffective disorder. Participants in all groups were excluded if they had a  
1042 history of neurological disease or head injury resulting in loss of consciousness.

1043 **Buxbaum, J | 20489179 | New York, US & Israel | scz\_msaf\_eur**

1044 Samples contributed by Mount Sinai were derived from three cohorts. In all cohorts, ethical  
1045 approval was obtained from all participating sites, and all subjects provided informed consent.  
1046 Two of the cohorts were in a prior paper on copy number variation. One of the cohorts was from  
1047 the Mount Sinai brain bank, where DNA was extracted from postmortem samples, and another

1048 comprised of patients ascertained in Israel. The third cohort included subjects more recently  
1049 recruited through the Mount Sinai Conte Center.

1050 **Corvin, A | 19571811 | Ireland | scz\_dubl\_eur**

1051 The case sample was collected primarily in the Dublin area and the ascertainment procedure has  
1052 been previously described. The controls were recruited, from the same region through the Irish  
1053 Blood Transfusion Services. All participants gave written, informed consent and the collections  
1054 were approved through the Federated Dublin Hospitals and Irish Blood Transfusion Services  
1055 Research Ethics Committees, respectively. DNA samples were genotyped at the Broad Institute.

1056 **Corvin, A; Riley, B | 22883433 | Ireland (WTCCC2) | scz\_irwt\_eur**

1057 The case sample was recruited from the Republic of Ireland and Northern Ireland. All cases had  
1058 four Irish grandparents and ascertainment details have been reported elsewhere. Ethics approval  
1059 was obtained from all participating hospitals and centers. Controls were blood donors from the  
1060 Irish Blood Transfusion Service, whose Ethics Committee approved the human subjects  
1061 protocol. All participants gave written informed consent. Samples were genotyped at Affymetrix  
1062 (Santa Clara, California, US) laboratory as part of the WTCCC2 genotyping pipeline.

1063 **Ehrenreich, H | 20819981 | Germany (GRAS) | scz\_gras**

1064 The Gottingen Research Association for Schizophrenia (GRAS) collection included cases  
1065 recruited across 23 German hospitals. Controls were unscreened blood donors recruited at the  
1066 Georg-August-University according to national blood donation guidelines. Cases completed a  
1067 structured clinical interview and were diagnosed with DSM-IV schizophrenia or schizoaffective  
1068 disorder. The study was approved by the Georg-August-University ethics committee and local  
1069 internal review boards of the participating centers. All participants gave written informed  
1070 consent.

1071 **Esko, T | 15133739 | Estonia (EGCUT) | scz\_egcu\_eur**

1072 The Estonian cohort comes from the population-based biobank of the Estonian Genome Project

1073 of University of Tartu (EGCUT). The project was conducted according to the Estonian Gene

1074 Research Act and all participants provided informed consent (www.biobank.ee). In total, 52,000

1075 individuals aged 18 years or older participated in this cohort (33% men, 67% women). The

1076 population distributions of the cohort reflect those of the Estonian population (83% Estonians,

1077 14% Russians and 3% other). General practitioners (GP) and physicians in the hospitals

1078 randomly recruited the participants. A Computer-Assisted Personal interview was conducted

1079 over 1-2 ours at doctors' offices. Data on demographics, genealogy, educational and

1080 occupational history, lifestyle and anthropometric and physiological data were assessed.

1081 Schizophrenia was diagnosed prior to the recruitment by a psychiatrist according to ICD-10

1082 criteria and identified from the Estonian Biobank phenotype database. Controls were drawn from

1083 a larger pool of genotyped biobank samples by matching on gender, age and genetic ancestry.

1084 All the controls were population-based and have not been sampled for any specific disease.

1085 **Esko, T; Li, Q; Dominici E | 15133739, 24166486 | J&J and Roche cases, EGCUT controls |**

1086 **scz\_jr3a\_eur**

1087 **Esko, T; Li, Q; Domenici E | 15133739, 24166486 | J&J and Roche cases, EGCUT controls |**

1088 **scz\_jr3b\_eur**

1089 **Esko, T; Li, Q; Domenici E | 15133739, 24166486 | J&J and Roche cases, EGCUT controls |**

1090 **scz\_jri6\_eur**

1091 **Esko, T; Li, Q; Dominici E | 15133739, 24166486 | J&J and Roche cases cases, EGCUT**

1092 **controls | scz\_jrsa\_eur**

1093 Cases were collected by Johnson and Johnson (J&J) and Roche as part of clinical collaborations  
1094 with hospitals and outpatient centers. Cases were diagnosed according to DSMIV criteria, with  
1095 medical record review by a trained psychiatrist. There were reliability trials across centers for the  
1096 J&J studies. The J&J cases were mostly collected in Eastern Europe, with most coming from  
1097 Estonian and Russia (>100); intermediate numbers from Austria, the Czech Republic, Latvia,  
1098 Lithuania, and Spain (50-100); and smaller collections from Bulgaria, Hungary, and Poland  
1099 (<50). The Roche cases were assessed with a structured psychiatric assessment by trained  
1100 interviewers. Most of the Eastern European controls were from the Estonian Biobank project  
1101 (EGCUT) and were ancestrally matched with cases from the J&J sample.

1102 **Gejman, P | 19571809 | US, Australia (MGS) | scz\_mgs2\_eur**

1103 European ancestry case samples were collected by the Molecular Genetics of Schizophrenia  
1104 (MGS) collaboration across multiple sites in the USA and Australia as described in detail  
1105 elsewhere. Cases gave written informed consent, and IRBs at each collecting site approved the  
1106 human subjects protocol. A survey company (Knowledge Networks, under MGS guidance)  
1107 collected the European ancestry control sample and ascertainment is described in detail  
1108 elsewhere. DNA samples were genotyped at the Broad Institute.

1109 **Gurling, H | 19571811 | London, UK | scz\_uclo\_eur**

1110 All cases and controls were collected by University College London and had both parents from  
1111 England, Scotland or Wales. All participants gave written informed consent and the U.K.  
1112 National Health Service multicenter and local research ethics committee approved the human  
1113 subjects protocol. Further details on ascertainment are available elsewhere. The samples were  
1114 genotyped at the Broad Institute.

1115 **Jönsson, E | 19571808 | Sweden (Hubin) | scz\_ersw\_eur**

1116 Cases were recruited from northwestern Stockholm County and ascertainment has been  
1117 described previously. Cases gave informed consent and the human subjects protocol was  
1118 approved by the ethical committees of the Karolinska Hospital and the Stockholm Regional  
1119 Ethical Committee. Controls were recruited either among subjects previously participating in  
1120 biological research at the Karolinska Institute or drawn from a representative register of the  
1121 population of Stockholm County. All participants provided informed consent.

1122 **Kirov, G | Not published | Bulgaria | scz\_buls\_eur**

1123 All cases were recruited from Bulgaria and had a history of hospitalization for treatment of  
1124 schizophrenia. Controls were recruited from the two largest cities in Bulgaria as previously  
1125 described. All participants gave written informed consent and the study was approved by local  
1126 ethics committees at the participating centers.

1127 **Knight, J; Collier DA; Nisenbaum L| Not published | Canada (Toronto) -US(Lilly)-US**  
1128 **(MIGen)| scz\_lktu\_eur**

1129 Toronto cases were recruited by referral and advertisement. Diagnoses were made according to  
1130 DSM-III or DSM-IV criteria following interview and medical record review. US cases were  
1131 recruited from schizophrenia clinical trials in a range of settings as part of a trial with Eli Lilly.  
1132 Diagnoses were made according to DSM-III or DSM-IV criteria following interview by  
1133 psychiatrist and medical record review. No controls were sampled as part of the study, and  
1134 ancestrally-matched controls were chosen from the Myocardial Infarction Genetics Consortium  
1135 (MIGen, dbGaP ID phs000294.v1.p1) that was genotyped with the same SNP array.

1136 **Lencz, T; Darvasi A | 23325106 | Israel | scz\_ajsz\_eur**

1137 Cases and controls were sampled from an Ashkenazi Jewish repository (Hebrew University  
1138 Genetic Resource, <http://huqr.huji.ac.il>). Patients were recruited from hospitalized inpatients at 7

1139 medical centers in Israel and were diagnosed with DSM-IV schizophrenia or schizoaffective  
1140 disorder. Controls were sampled through the Israeli Blood Bank and did not report any chronic  
1141 disease or regularly prescribed medication at the time of assessment. Full ascertainment details  
1142 have previously been reported. Local ethics committees and the National Genetic Committee of  
1143 the Israeli Ministry of Health approved the studies and all participants gave informed, written  
1144 consent.

1145 **Levinson, D | 22885689 | Six countries, WTCCC controls | scz\_lacw\_eur**

1146 Cases collected as part of a larger pedigree-based study were partitioned into two subsamples.  
1147 Cases with two genotyped parents were analyzed as trios (see PI Levinson, ms.scz\_lemu\_eur in  
1148 the Trio section below). Unrelated cases who could not be used as part of a trio were included as  
1149 a separate case-control analysis, using independent controls, matched by ancestry and  
1150 genotyping array, from the Wellcome Trust Case Control Consortium. Cases were identified  
1151 from different clinical settings (e.g. inpatients, outpatients and community facilities) in six  
1152 countries (Australia, France, Germany, Ireland, UK, and the US). Diagnoses were established  
1153 using semi-structured interviews, psychiatric records and informant reports. Case subjects were  
1154 diagnosed with schizophrenia or schizoaffective disorder according to DSM-III-R criteria. All  
1155 protocols were approved by loci IRBs, and all cases provided written informed consent.

1156 **Malhotra, A | 17522711 | New York, US | scz\_zhh1\_eur**

1157 The case and control subjects were recruited in the New York metropolitan area and  
1158 ascertainment methods have been described previously. All participants gave written, informed  
1159 consent and the IRB of the North Shore-Long Island Jewish Health System approved the human  
1160 subjects protocols. DNA was genotyped at Zucker Hillside.

1161 **Mowry, B | 21034186 | Australia | scz\_asrb\_eur**



1162 These subjects were part of the Australian Schizophrenia Research Bank. The case sample was  
1163 recruited in four Australian States (New South Wales, Queensland, Western Australia and  
1164 Victoria) through hospital inpatient units, community mental health services, outpatient clinics  
1165 and rehabilitation services, non-government mental illness support organizations, and, in the  
1166 initial stages, through a large-scale, national, multi-media advertising campaign. This sample is  
1167 comprised of 509 cases from larger metropolitan centers of Brisbane, Newcastle, Sydney,  
1168 Melbourne, and Perth. Cases gave written informed consent, and the human subjects protocol  
1169 was initially approved by the Hunter New England Area Health Research Committee and  
1170 subsequently approved by relevant Institutional Ethics Committees in Brisbane, Sydney,  
1171 Melbourne and Perth. Healthy controls were recruited through multi-media advertisements, and  
1172 other sources. Controls were from the metropolitan centers of Brisbane, Newcastle, Sydney,  
1173 Melbourne, and Perth. Controls gave written informed consent, and the human subjects protocol  
1174 was approved by the Hunter New England Area Health Research Committee and Institutional  
1175 Ethics Committees in Brisbane, Sydney, Melbourne and Perth. The samples were genotyped in  
1176 two stages at the Hunter Medical Research Institute, University of Newcastle, Newcastle,  
1177 Australia.

1178 **O'Donovan, M: Owen, M | 19571811 | Cardiff, UK | scz\_caws\_eur**

1179 The case sample included European ancestry schizophrenia cases recruited in the British Isles  
1180 and described previously. All cases gave written informed consent to. The study was approved  
1181 by the Multicentre Research Ethics Committee in Wales and Local Research Ethics Committees  
1182 from all participating sites. The control sample used the Wellcome Trust CaseControl  
1183 Consortium (WTCCC) sample described elsewhere, but included similar numbers of individuals

1184 from the 1958 British Birth Cohort and a panel of consenting blood donors (UK Blood Service).  
1185 Samples were genotyped at Affymetrix service lab (San Francisco, USA).

1186 **O'Donovan, M: Owen, M: Walters, J | 22614287 | UK (CLOZUK) | scz\_clm2\_eur**

1187 **O'Donovan, M: Owen, M: Walters, J | 22614287 | UK (CLOZUK) | scz\_clo3\_eur**

1188 CLOZUK cases were taking the antipsychotic clozapine and had received a clinical diagnosis of  
1189 treatment-resistant schizophrenia. Patients taking clozapine provide blood samples to allow  
1190 detection of adverse drug-effects. Through collaboration with Novartis (the manufacturer of a  
1191 proprietary form of clozapine, Clozaril), we acquired blood from people with treatment-resistant  
1192 schizophrenia according to the clozapine registration forms completed by treating psychiatrists  
1193 as previously reported. The samples were genotyped at the Broad Institute. The UK Multicentre  
1194 Research Ethics Committee (MREC) approved the study. The controls were drawn from the  
1195 WTCCC2 control samples (~3,000 from the 1958 British Birth Cohort and ~3,000 samples from  
1196 the UK Blood Service Control Group). An additional 900 controls, held by Cardiff University,  
1197 were recruited from the UK National Blood Transfusion Service. They were not specifically  
1198 screened for psychiatric illness. All control samples were from participants who provided  
1199 informed consent.

1200 **Ophoff, R | 19571808 | Netherlands | scz\_ucla\_eur**

1201 The case sample consisted of inpatients and outpatients recruited through psychiatric hospitals  
1202 and institutions throughout the Netherlands. Cases with DSM-IV schizophrenia were included in  
1203 the analysis. Further details on ascertainment are provided elsewhere. Controls came from the  
1204 University Medical Centre Utrecht and were volunteers with no psychiatric history. Ethical  
1205 approval was provided by local ethics committees and all participants gave written informed  
1206 consent.

1207 **Palotie, A | 19571808 | Finland | scz\_fi3m\_eur**

1208 **Palotie, A | Not published | Finnish | scz\_fii6\_eur**

1209 Finnish cases were drawn from a nationwide collection of families with schizophrenia spectrum  
 1210 disorders. The control sample was derived from the Finnish Health 2000 survey. All participants  
 1211 provided written informed consent and approval was obtained from the ethics committees at each  
 1212 location.

1213 **Pato, C | 19571811 | Portugal | scz\_port\_eur**

1214 Cases and controls lived in Portugal, the Azorean and Madeiran islands, or were the direct  
 1215 (first or second-generation) Portuguese immigrant population in the US, as previously described.  
 1216 Controls were not biologically related to cases. All participants gave written informed consent  
 1217 and the IRB of SUNY Upstate Medical University approved the protocol. The samples were  
 1218 genotyped at the Broad Institute.

1219 **Petryshen, T | 24424392 | Boston, US (CIDAR) | scz\_cims\_eur**

1220 Cases were recruited from inpatient and outpatient settings in the Boston area by clinician  
 1221 referral, through review of medical records, or through advertisements in local media. Cases  
 1222 were diagnosed with DSM-IV schizophrenia through a structured clinical interview (SCID) by  
 1223 trained interviewers with review of medical records and a best estimate diagnostic procedure  
 1224 including reliability trials across interviewers. A psychiatrist or a PhD-level mental health  
 1225 professional made the final diagnostic determination. Controls were ascertained through local  
 1226 advertisements from the same geographical area. Ethical approval was provided by local ethics  
 1227 committees and all participants gave written informed consent.

1228 **Rietschel/Rujescu/Nöthen | 19571808 | Bonn/Mannheim, Germany | scz\_boco\_eur**

1229 These German samples were collected by separate groups within the MoodS Consortium in  
1230 Mannheim, Bonn, Munich and Jena. For the PGC analyses, the samples were combined by chip  
1231 and ancestry. In Bonn/Mannheim, cases were ascertained as previously described. Controls were  
1232 drawn from three population-based epidemiological studies (PopGen), the Cooperative Health  
1233 Research in the Region of Augsburg (KORA) study, and the Heinz Nixdorf Recall (HNR) study.  
1234 All participants gave written informed consent and the local ethics committees approved the  
1235 human subjects protocols. Additional controls were randomly selected from a Munich-based  
1236 community sample and screened for the presence of anxiety and affective disorders using the  
1237 Composite International Diagnostic Screener. Only individuals negative for the above mentioned  
1238 disorders were included in the sample.

1239 **Rujescu, D | 19571808 | Munich, Germany | scz\_munc\_eur**

1240 For the Munich sample, cases were ascertained from the Munich area of Germany, as described  
1241 previously. The controls were unrelated volunteers randomly selected from the general  
1242 population of Munich. All were screened to exclude a history of psychosis/central neurological  
1243 disease either personally or in a first-degree relative. All participants gave written informed  
1244 consent and the local ethics committees approved the human subjects protocols.

1245 **St Clair, D | 19571811 | Aberdeen, UK | scz\_aber\_eur**

1246 Ascertainment and inclusion/exclusion criteria for cases and controls have been previously  
1247 described. All participating subjects were born in the UK (95% Scotland) and gave written  
1248 informed consent. Both local and multiregional academic ethical committee approved the human  
1249 subjects protocol. The samples were genotyped at the Broad Institute.

1250 **Sullivan, PF | 18347602 | US (CATIE) | scz\_cati\_eur**

1251 Cases were collected as part of the Clinical Antipsychotics Trials of Intervention Effectiveness  
1252 (CATIE) project and ascertainment was previously described. Participants were recruited from  
1253 multiple sites in the USA with informed written consent and approval from the IRBs at each  
1254 CATIE site and the University of North Carolina (Chapel Hill). The control subjects were  
1255 collected by MGS (described above) and gave online informed consent and were fully  
1256 anonymized. There was no overlap with controls included in the MGS collaboration sample.

1257 **Sullivan, PF; Sklar P; Hultman C | 23974872 | Sweden | scz\_swe1\_eur**

1258 **Sullivan, PF; Sklar P; Hultman C | 23974872 | Sweden | scz\_s234\_eur**

1259 **Sullivan, PF; Sklar P; Hultman C | 23974872 | Sweden | scz\_swe5\_eur**

1260 **Sullivan, PF; Sklar P; Hultman C | 23974872 | Sweden | scz\_swe6\_eur**

1261 Samples from the Swedish Schizophrenia Study were collected in a multi-year project and  
1262 genotypes in six batches (sw1-6). All procedures were approved by ethical committees at the  
1263 Karolinska Institutet and the University of North Carolina, and all subjects provided written  
1264 informed consent (or legal guardian consent and subject assent). All samples were genotyped at  
1265 the Broad Institute. Cases with schizophrenia were identified via the Swedish Hospital Discharge  
1266 Register which captures all public and private inpatient hospitalizations. The register is complete  
1267 from 1987 and is augmented by psychiatric data from 1973-1986. The register contains  
1268 International Classification of Disease discharge diagnoses made by attending physicians for  
1269 each hospitalization. Case inclusion criteria included  $\geq 2$  hospitalizations with a discharge  
1270 diagnosis of schizophrenia, both parents born in Scandinavia and age  $\geq 18$  years. Case exclusion  
1271 criteria included hospital register diagnosis of any medical or psychiatric disorder mitigating a  
1272 confident diagnosis of schizophrenia as determined by expert review. The validity of this case  
1273 definition of schizophrenia was strongly supported by clinical, epidemiological, genetic

1274 epidemiological and genetic evidence. Controls were selected at random from Swedish  
1275 population registers, with the goal of obtaining an appropriate control group and avoiding ‘super-  
1276 normal’ controls. Control inclusion criteria included never being hospitalized for schizophrenia  
1277 or bipolar disorder (given evidence of genetic overlap with schizophrenia), both parents born in  
1278 Scandinavia and age of  $\geq 18$  years.

1279 **Walters, J | 21850710 | Cardiff, UK (CogUK) | scz\_cou3\_eur**

1280 Cases were recruited from community mental health teams in Wales and England on the basis of  
1281 a clinical diagnosis of schizophrenia or schizoaffective disorder (depressed sub-type) as  
1282 described previously. 35 Diagnosis was confirmed following a SCAN interview and review of  
1283 case notes followed by consensus diagnosis according to DSM-IV criteria. The samples were  
1284 genotyped at the Broad Institute. The UK Multicentre Research Ethics Committee (MREC)  
1285 approved the study and all participants provided valid informed consent.

1286 **Weinberger, D | 11381111 | NIMH CBDB | scz\_lie2\_eur**

1287 **Weinberger, D | 11381111 | NIMH CBDB | scz\_lie5\_eur**

1288 Subjects were recruited from the Clinical Brain Disorders Branch of the NIMH ‘Sibling Study’  
1289 as previously described. In brief, cases and controls gave informed consent and only participants  
1290 of European ancestry were included in the current analysis. Cases completed a structured clinical  
1291 interview and were diagnosed with schizophrenia-spectrum disorders. Samples were genotyped  
1292 at the NIMH.

1293 **Wendland/Schubert | Pfizer | Not Published | Multiple countries | scz\_pfla\_eur**

1294 Pfizer contributed anonymized individual genotypes for cases from seven multi-center  
1295 randomized, double-blind efficacy and safety clinical trials (A1281063, A1281134, A1281148,  
1296 A245-102, NRA7500001, NRA7500002, NRA7500003, and NRA7500004) as well as a set of

purchased samples (NRA9000099). Trial samples were collected for antipsychotic medications across outpatient and inpatient treatment settings. All participating cases had a diagnosis of schizophrenia and were assessed using a structural clinical interview by trained interviewers, with systematic procedures to quality-control diagnostic accuracy and reliability trials across participating sites in the United States and internationally. Purchased blood samples were obtained from PrecisionMed International by Pharmacia and Upjohn Corporation, and were collected from diagnosed subjects with schizophrenia and schizoaffective disorder. All studies were reviewed by both central and local institutional review boards, depending on the study site, before recruitment of subjects started. Protocol amendments were approved while the study was in progress and before the data were unblinded. The studies were conducted in conformity with the U.S. Food and Drug Administration Code of Federal Regulations (21CFR, Part 50) and the Declaration of Helsinki and its amendments, and were consistent with Good Clinical Practice and the applicable regulatory requirements. Participants provided written informed consent before enrollment. An optional blood sample was collected from clinical trial subjects for pharmacogenetic analysis to investigate potential associations between genetic variant drug response and general characteristics of schizophrenia and related disorders. Sample collection was not required for participation in the original clinical trials. The controls (A9011027) were recruited in a multi-site, cross-sectional, non-treatment prospective trial to collect data, including DNA, from cognitive normal and free of psychiatric diseases elderly subjects in the US. Subjects were specifically recruited to match the gender, age, and ethnicity information from the LEADe and UCSD MCI studies. The study described here is within the scope of patient consent.

**Werge, T | 19571808 | Denmark | scz\_denm\_eur**

1319 Cases were ascertained through psychiatric departments and twin pair studies, and were of  
1320 Danish parentage for at least the prior three generations. The controls were collected at the  
1321 University of Aarhus, and included 500 medical students, all of Danish parentage for at least  
1322 three generations. All subjects gave written informed consent and the Danish Data Protection  
1323 Agency and the ethics committees of Denmark approved the human subjects protocol.

1324

1325 *Bipolar Disorder*

1326 **Adolfsson, R | Not published | Umeå, Sweden | bip\_ume4\_eur**

1327 Clinical characterization of the patients included the Mini-International Neuropsychiatric  
1328 Interview (MINI), the Diagnostic Interview for Genetic Studies (DIGS), the Family Interview for  
1329 Genetic Studies (FIGS) and the Schedules for Clinical Assessment in Neuropsychiatry (SCAN).  
1330 The final diagnoses were made according to the DSM-IV-TR and determined by consensus of 2  
1331 research psychiatrists. The unrelated Swedish control individuals, consisting of a large  
1332 population-based sample representative of the general population of the region, were randomly  
1333 selected from the ‘Betula study’.

1334 **Alda, M; Smoller, J | Not published | Nova Scotia, Canada; I2B2 controls | bip\_hal2\_eur**

1335 The case samples were recruited from patients longitudinally followed at specialty mood  
1336 disorders clinics in Halifax and Ottawa (Canada). Cases were interviewed in a blind fashion with  
1337 the Schedule of Affective Disorders and Schizophrenia-Lifetime version (SADS-L) and  
1338 consensus diagnoses were made according to DSM-IV and Research Diagnostic Criteria (RDC).  
1339 Protocols and procedures were approved by the local Ethics Committees and written informed  
1340 consent was obtained from all patients before participation in the study. Control subjects were  
1341 drawn from the I2B2 (Informatics for Integrating Biology and the Bedside) project. The study



1342 consists of de-identified healthy individuals recruited from a healthcare system in the Boston,  
1343 MA, US area. The de-identification process meant that the Massachusetts General Hospital  
1344 Institutional Review Board elected to waive the requirement of seeking informed consent as  
1345 detailed by US Code of Federal Regulations, Title 45, Part 46, Section 116 (46.116).

1346 **Andreassen, OA | PMID:21926972 [PGC1], PMID:20451256 | Norway (TOP) |**  
1347 **bip\_top7\_eur**

1348 In the TOP study (Tematisk område psykoser), cases of European ancestry, born in Norway,  
1349 were recruited from psychiatric hospitals in the Oslo region. Patients were diagnosed according  
1350 to the SCID and further ascertainment details have been reported. Healthy control subjects were  
1351 randomly selected from statistical records of persons from the same catchment area as the patient  
1352 groups. The control subjects were screened by interview and with the Primary Care Evaluation  
1353 of Mental Disorders (PRIME-MD). None of the control subjects had a history of  
1354 moderate/severe head injury, neurological disorder, mental retardation or an age outside the age  
1355 range of 18-60 years. Healthy subjects were excluded if they or any of their close relatives had a  
1356 lifetime history of a severe psychiatric disorder. All participants provided written informed  
1357 consent and the human subjects protocol was approved by the Norwegian Scientific-Ethical  
1358 Committee and the Norwegian Data Protection Agency.

1359 **Andreassen, OA | Not published | Norway (TOP) | bip\_top8\_eur**

1360 The TOP8 bipolar disorder cases and controls were ascertained in the same way as the  
1361 bip\_top7\_eur (TOP7) samples described above, and recruited from hospitals across Norway.

1362 **Biernacka, JM; Frye, MA | 27769005 | Mayo Clinic, USA | bip\_may1\_eur**

1363 Bipolar cases were drawn from the Mayo Clinic Bipolar Biobank. Enrolment sites included  
1364 Mayo Clinic, Rochester, Minnesota; Lindner Center of HOPE/University of Cincinnati College

1365 of Medicine, Cincinnati, Ohio; and the University of Minnesota, Minneapolis, Minnesota.  
1366 Enrolment at each site was approved by the local Institutional Review Board approval, and all  
1367 participants consented to use of their data for future genetic studies. Participants were identified  
1368 through routine clinical appointments, from in-patients admitted in mood disorder units, and  
1369 recruitment advertising. Participants were required to be between 18 and 80 years old and be able  
1370 to speak English, provide informed consent, and have DSM-IV-TR diagnostic confirmation of  
1371 type 1 or 2 bipolar disorder or schizoaffective bipolar disorder as determined using the SCID.  
1372 Controls were selected from the Mayo Clinic Biobank. Potential controls with ICD9 codes for  
1373 bipolar disorder, schizophrenia or related diagnoses in their electronic medical record were  
1374 excluded.

1375 **Blackwood, D | 18711365 [PGC1] | Edinburgh, UK | bip\_edi1\_eur**

1376 This sample comprised Caucasian individuals contacted through the inpatient and outpatient  
1377 services of hospitals in South East Scotland. A BD-I diagnosis was based on an interview with  
1378 the patient using the SADS-L supplemented by case note review and frequently by information  
1379 from medical staff, relatives and caregivers. Final diagnoses, based on DSM-IV criteria were  
1380 reached by consensus between two trained psychiatrists. Ethnically-matched controls from the  
1381 same region were recruited through the South of Scotland Blood Transfusion Service. Controls  
1382 were not directly screened to exclude those with a personal or family history of psychiatric  
1383 illness. The study was approved by the Multi-Centre Research Ethics Committee for Scotland  
1384 and patients gave written informed consent for the collection of DNA samples for use in genetic  
1385 studies.

1386 **Breen, G; Vincent, JB | 24387768; 19416921; 21926972 [PGC1] | London, UK; Toronto,**  
1387 **Canada [BACC] | bip\_bac1\_eur**

1388 The total case/control cohort (N=1922) includes 871 subjects from Toronto, Canada (N=431  
1389 cases (160 male; 271 female); N=440 controls (176 male; 264 female)), 1051 subjects from  
1390 London, UK (N=538 cases (180 male; 358 female); N=513 controls (192 male; 321 female)). A  
1391 summary of mean and median age at interview, age of onset (AOO), diagnostic subtypes (BD 1  
1392 versus BD 2), presence of psychotic symptoms, suicide attempt and family history of psychiatric  
1393 disorders has been provided previously for both the Toronto and London cohorts. From the  
1394 Toronto site (Centre for Addiction & Mental Health (CAMH)), BD individuals and unrelated  
1395 healthy controls matched for age, gender and ethnicity were recruited. Inclusion criteria for  
1396 patients: a) diagnosed with DSMIV/ICD 10 BD 1 or 2; b) 18 years old or over; c) Caucasian, of  
1397 Northern and Western European origin, and three out of four grandparents also N.W. European  
1398 Caucasian. Exclusion criteria include: a) Use of intravenous drugs; b) Evidence of intellectual  
1399 disability; c) Related to an individual already in the study; d) Manias that only ever occurred in  
1400 relation to or resulting from alcohol or substance abuse/dependence, or medical illness; e)  
1401 Manias resulting from non-psychotropic substance usage. The SCAN interview (Schedule for  
1402 Clinical Assessments in Neuropsychiatry) was used for subject assessment. Using the SCAN  
1403 interview along with case note review, each case was assigned DSM-IV and ICD 10 diagnoses  
1404 by two independent diagnosticians, according to lifetime consensus best-estimate diagnosis.  
1405 Lifetime occurrence of psychiatric symptoms was also recorded using the OPCRIT checklist,  
1406 modified for use with mood disorders. Similar methods and criteria were also used to collect a  
1407 sample of 538 BD cases and 513 controls for the London cohort (King's College London; KCL).  
1408 Both studies were approved by respective institutional research ethics committees (the CAMH  
1409 Research Ethics Board (REB) in Toronto, and the College Research Ethics Committee (CREC)  
1410 at KCL), and informed written consent was obtained from all participants. GWAS results have

1411 previously been published for the entire KCL/CAMH cohort.

1412 **Corvin, A | 18711365 [PGC1] | Ireland | bip\_dub1\_eur**

1413 Samples were collected as part of a larger study of the genetics of psychotic disorders in the  
 1414 Republic of Ireland, under protocols approved by the relevant IRBs and with written informed  
 1415 consent that permitted repository use. Cases were recruited from Hospitals and Community  
 1416 psychiatric facilities in Ireland by a psychiatrist or psychiatric nurse trained to use the SCID.  
 1417 Diagnosis was based on the structured interview supplemented by case note review and collateral  
 1418 history where available. All diagnoses were reviewed by an independent reviewer. Controls were  
 1419 ascertained with informed consent from the Irish GeneBank and represented blood donors who  
 1420 met the same ethnicity criteria as cases. Controls were not specifically screened for psychiatric  
 1421 illness.

1422 **Rietschel, M; Nöthen, MM, Cichon, S | 21926972 [PGC1] | BOMA-Germany I |**  
 1423 **bip\_bonn\_eur**

1424 Cases for the BOMA-Bipolar Study were ascertained from consecutive admissions to the  
 1425 inpatient units of the Department of Psychiatry and Psychotherapy at the University of Bonn and  
 1426 at the Central Institute for Mental Health in Mannheim, University of Heidelberg, Germany.  
 1427 DSM-IV lifetime diagnoses of bipolar I disorder were assigned using a consensus best-estimate  
 1428 procedure, based on all available information, including a structured interview with the SCID  
 1429 and SADS-L, medical records, and the family history method. In addition, the OPCRIT checklist  
 1430 was used for the detailed polydiagnostic documentation of symptoms. Controls were ascertained  
 1431 from three population-based studies in Germany (PopGen, KORA, and Heinz-Nixdorf-Recall  
 1432 Study). The control subjects were not screened for mental illness. Study protocols were reviewed  
 1433 and approved in advance by Institutional Review Boards of the participating institutions. All

1434 subjects provided written informed consent.

1435 **Rietschel, M; Nöthen, MM; Schulze, TG; Reif, A; Forstner, AJ | 24618891 | BOMA-**  
1436 **Germany II | bip\_bmg2\_eur**

1437 Cases were recruited from consecutive admissions to psychiatric in-patient units at the  
1438 University Hospital Würzburg. All cases received a lifetime diagnosis of BD according to the  
1439 DSM-IV criteria using a consensus best-estimate procedure based on all available information,  
1440 including semi-structured diagnostic interviews using the Association for Methodology and  
1441 Documentation in Psychiatry, medical records and the family history method. In addition, the  
1442 OPCRIT system was used for the detailed polydiagnostic documentation of symptoms.

1443 Control subjects were ascertained from the population-based Heinz Nixdorf Recall (HNR) Study.  
1444 The controls were not screened for a history of mental illness. Study protocols were reviewed  
1445 and approved in advance by Institutional Review Boards of the participating institutions. All  
1446 subjects provided written informed consent.

1447 **Rietschel, M; Nöthen, MM; Schulze, TG; Bauer, M; Forstner, AJ; Müller-Myhsok, B |**  
1448 **24618891 | BOMA-Germany III | bip\_bmg3\_eur**

1449 Cases were recruited at the Central Institute of Mental Health in Mannheim, University of  
1450 Heidelberg, and other collaborating psychiatric hospitals in Germany. All cases received a  
1451 lifetime diagnosis of BD according to the DSM-IV criteria using a consensus best-estimate  
1452 procedure based on all available information including structured diagnostic interviews using the  
1453 AMDP, Composite International Diagnostic Screener (CID-S), SADS-L and/or SCID, medical  
1454 records, and the family history method. In addition, the OPCRIT system was used for the  
1455 detailed polydiagnostic documentation of symptoms. Controls were selected randomly from a  
1456 Munich-based community sample and recruited at the Max-Planck Institute of Psychiatry. They

were screened for the presence of anxiety and mood disorders using the CID-S. Only individuals without mood and anxiety disorders were collected as controls. Study protocols were reviewed and approved in advance by Institutional Review Boards of the participating institutions. All subjects provided written informed consent.

**Hauser, J; Lissowska, J; Forstner, AJ | 24618891 | BOMA-Poland | bip\_bmpo\_eur**

Cases were recruited at the Department of Psychiatry, Poznan University of Medical Sciences, Poznan, Poland. All cases received a lifetime diagnosis of BD according to the DSM-IV criteria on the basis of a consensus best-estimate procedure and structured diagnostic interviews using the SCID. Controls were drawn from a population-based case-control sample recruited by the Cancer-Center and Institute of Oncology, Warsaw, Poland and a hospital-based case-control sample recruited by the Nofer Institute of Occupational Medicine, Lodz, Poland. The Polish controls were produced by the International Agency for Research on Cancer (IARC) and the Centre National de Génotypage (CNG) GWAS Initiative for a study of upper aerodigestive tract cancers. The controls were not screened for a history of mental illness. Study protocols were reviewed and approved in advance by Institutional Review Boards of the participating institutions. All subjects provided written informed consent.

**Rietschel, M; Nöthen, MM; Rivas, F; Mayoral, F; Kogevinas, M; others | 24618891 | BOMA-Spain | bip\_bmsp\_eur**

Cases were recruited at the mental health departments of the following five centers in Andalusia, Spain: University Hospital Reina Sofia of Córdoba, Provincial Hospital of Jaen; Hospital of Jerez de la Frontera (Cádiz); Hospital of Puerto Real (Cádiz); Hospital Punta Europa of Algeciras (Cádiz); and Hospital Universitario San Cecilio (Granada). Diagnostic assessment was performed using the SADS-L; the OPCRIT; a review of medical records; and interviews with

1480 first and/or second degree family members using the Family Informant Schedule and Criteria  
1481 (FISC). Consensus best estimate BD diagnoses were assigned by two or more independent senior  
1482 psychiatrists and/or psychologists, and according to the RDC, and the DSM-IV. Controls were  
1483 Spanish subjects drawn from a cohort of individuals recruited in the framework of the European  
1484 Community Respiratory Health Survey (ECRHS, <http://www.ecrhs.org/>). The controls were not  
1485 screened for a history of mental illness. Study protocols were reviewed and approved in advance  
1486 by Institutional Review Boards of the participating institutions. All subjects provided written  
1487 informed consent.

1488 **Fullerton, J.M.; Mitchell, P.B.; Schofield, P.R.; Martin N.G.; Cichon, S. | 24618891 |**  
1489 **BOMA-Australia | bip\_bmau\_eur**

1490 Cases were recruited at the Mood Disorder Unit, Prince of Wales Hospital in Sydney. All cases  
1491 received a lifetime diagnosis of BD according to the DSM-IV criteria on the basis of a consensus  
1492 best-estimate procedure and structured diagnostic interviews using the DIGS, FIGS, and the  
1493 SCID. Controls were parents of unselected adolescent twins from the Brisbane Longitudinal  
1494 Twin Study. The controls were not screened for a history of mental illness. Study protocols were  
1495 reviewed and approved in advance by Institutional Review Boards of the participating  
1496 institutions. All subjects provided written informed consent.

1497 **Grigoriu-Serbanescu, M; Nöthen, MM | 21353194 | BOMA-Romania | bip\_rom3\_eur**

1498 Cases were recruited from consecutive admissions to the Obregia Clinical Psychiatric Hospital,  
1499 Bucharest. Patients were administered the DIGS and FIGS interviews. Information was also  
1500 obtained from medical records and close relatives. The diagnosis of BP-I was assigned according  
1501 to DSM-IV criteria using the best estimate procedure. All patients had at least two hospitalized  
1502 illness episodes. Population-based controls were evaluated using the DIGS to exclude a lifetime

1503 history of major affective disorders, schizophrenia, schizoaffective disorders, and other  
1504 psychoses, obsessive-compulsive disorder, eating disorders, and alcohol or drug addiction.

1505 **Craddock, N, Jones, I, Jones, L | 17554300 | WTCCC | bip\_wtcc\_eur\_sr-qc**

1506 Cases were all over the age of 17 yr, living in the UK and of European descent. Recruitment was  
1507 undertaken throughout the UK and included individuals who had been in contact with mental  
1508 health services and had a lifetime history of high mood. After providing written informed  
1509 consent, participants were interviewed by a trained psychologist or psychiatrist using a semi-  
1510 structured lifetime diagnostic psychiatric interview (Schedules for Clinical Assessment in  
1511 Neuropsychiatry) and available psychiatric medical records were reviewed. Using all available  
1512 data, best-estimate life-time diagnoses were made according to the RDC. In the current study we  
1513 included cases with a lifetime diagnosis of RDC bipolar 1 disorder, bipolar 2 disorder or schizo-  
1514 affective disorder, bipolar type. Controls were recruited from two sources: the 1958 Birth Cohort  
1515 study and the UK Blood Service (blood donors) and were not screened for history of mental  
1516 illness. All cases and controls were recruited under protocols approved by the appropriate IRBs.  
1517 All subjects gave written informed consent.

1518 **Kelsoe, J | 21926972 [PGC1] | USA (GAIN) | bip\_gain\_eur**

1519 *Genetic Association Information Network (GAIN)/ The Bipolar Genome Study (BiGS)* The BD  
1520 sample was collected under the auspices of the NIMH Genetics Initiative for BD  
1521 (<http://zork.wustl.edu/nimh/>), genotyped as part of GAIN and analyzed as part of a larger GWAS  
1522 conducted by the BiGS consortium. Approximately half of the GAIN sample was collected as  
1523 multiplex families or sib pair families (waves 1-4), the remainder were collected as individual  
1524 cases (wave 5). Subjects were ascertained at 11 sites: Indiana University, John Hopkins  
1525 University, the NIMH Intramural Research Program, Washington University at St. Louis,



1526 University of Pennsylvania, University of Chicago, Rush Medical School, University of Iowa,  
1527 University of California, San Diego, University of California, San Francisco, and University of  
1528 Michigan. All investigations were carried out after the review of protocols by the IRB at each  
1529 participating institution. At all sites, potential cases were identified from screening admissions to  
1530 local treatment facilities and through publicity programs or advocacy groups. Potential cases  
1531 were evaluated using the DIGS, FIGS, and information from relatives and medical records. All  
1532 information was reviewed through a best estimate diagnostic procedure by two independent and  
1533 non-interviewing clinicians and a consensus best-estimate diagnosis was reached. In the event of  
1534 a disagreement, a third review was done to break the tie. Controls were from the NIMH Genetic  
1535 Repository sample obtained by Dr. P. Gejman through a contract to Knowledge Networks, Inc.  
1536 Only individuals with complete or near-complete psychiatric questionnaire data who did not  
1537 fulfill diagnostic criteria for major depression and denied a history of psychosis or BD were  
1538 included as controls for BiGS analyses. Controls were matched for gender and ethnicity to the  
1539 cases.

1540 **Kelsoe, J; Sklar, P; Smoller, J | [PGC1 Replication] | USA (FAT2; FaST, BiGS, TGEN) |**  
1541 **bip\_fat2\_eur**

1542 Cases were collected from individuals at the 11 U.S. sites described for the GAIN sample.  
1543 Eligible participants were age 18 or older meeting DSM-IV criteria for BD-I or BD-II by  
1544 consensus diagnosis based on interviews with the Affective Disorders Evaluation (ADE) and  
1545 MINI. All participants provided written informed consent and the study protocol was approved  
1546 by IRBs at each site. Collection of phenotypic data and DNA samples were supported by NIMH  
1547 grants MH063445 (JW Smoller); MH067288 (PI: P Sklar), and MH63420 (PI: V Nimgaonkar).  
1548 The control samples were NIMH controls that were using the methods described in that section.

1549 The case and control samples were independent of those included in the GAIN sample.

1550 **Kirov, G | 25055870 | Bulgarian trios | bip\_butr\_eur**

1551 All cases were recruited in Bulgaria from psychiatric inpatient and outpatient services. Each  
1552 proband had a history of hospitalisation and was interviewed with an abbreviated version of the  
1553 SCAN. Consensus best-estimate diagnoses were made according to DSM-IV criteria by two  
1554 researchers. All participants gave written informed consent and the study was approved by local  
1555 ethics committees at the participating centers.

1556 **Kirov, G | 25055870 | UK trios | bip\_uktr\_eur**

1557 The BD subjects were recruited from lithium clinics and interviewed in person by a senior  
1558 psychiatrist, using abbreviated version of the SCAN. Consensus best-estimate diagnoses were  
1559 made based on the interview and hospital notes. Ethics committee approval for the study was  
1560 obtained from the relevant research ethics committees and all individuals provided written  
1561 informed consent for participation.

1562 **Landén, M; Sullivan, PF; Sklar, P | [ICCBD] | Sweden (ICCBD) | bip\_swa2\_eur**

1563 The BD subjects were identified using the Swedish National Quality Register for Bipolar  
1564 Disorders (Bipolär) and the Swedish National Patient Register (using a validated algorithm  
1565 requiring at least two hospitalizations with a BD diagnosis). A confirmatory telephone interview  
1566 with a diagnostic review was conducted. Additional subjects were recruited from the St. Göran  
1567 Bipolar Project (Affective Center at Northern Stockholm Psychiatry Clinic, Sweden), enrolling  
1568 new and ongoing patients diagnosed with BD using structured clinical interviews. Diagnoses  
1569 were made according to the DSM-IV criteria (Bipolär and St. Göran Bipolar Project) and ICD-  
1570 10 (National Patient Register). The control subjects used were the same as for the SCZ analyses  
1571 described above. All ascertainment procedures were approved by the Regional Ethical

1572 Committees in Sweden.

1573 **Landén, M; Sullivan, PF; Sklar, P | [ICCBD] | Sweden (ICCBD) | bip\_swei\_eur**

1574 The cases and controls in the bip\_swei\_eur sample were recruited using the same ascertainment  
1575 methods described for the bip\_swa2\_eur sample.

1576 **Leboyer, M | [PGC1 replication] | France | bip\_fran\_eur**

1577 Cases with BD1 or BD2 and control samples were recruited as part of a large study of genetics of  
1578 BD in France (Paris-Creteil, Bordeaux, Nancy) with a protocol approved by relevant IRBs and  
1579 with written informed consent. Cases were of French descent for more than 3 generations were  
1580 assessed by a trained psychiatrist or psychologist using structured interviews supplemented by  
1581 medical case notes, mood scales and self-rating questionnaire assessing dimensions.

1582 **Li, Q | 24166486; 27769005 | USA (Janssen), SAGE controls | bip\_jst5\_eur**

1583 The study included unrelated patients with bipolar 1 disorder from 6 clinical trials (IDs:  
1584 NCT00253162, NCT00257075, NCT00076115, NCT00299715, NCT00309699, and  
1585 NCT00309686). Participant recruitment was conducted by Janssen Research & Development,  
1586 LLC (formerly known as Johnson & Johnson Pharmaceutical Research & Development, LLC) to  
1587 assess the efficacy and safety of risperidone. Bipolar cases were diagnosed according to DSM-  
1588 IV-TR criteria. The diagnosis of bipolar disorder was confirmed by the Schedule for Affective  
1589 Disorders and Schizophrenia for School-Age Children-Present and Lifetime Version (K-SADS-  
1590 PL) in NCT00076115, by the SCID in NCT00257075 and NCT00253162, or by the MINI in  
1591 NCT00299715 and NCT00309699, and NCT00309686, respectively. Additional detailed  
1592 descriptions of these clinical trials can be found at ClinicalTrials.gov. Only patients of European  
1593 ancestry with matching controls were included in the current analysis. Controls subjects were  
1594 drawn from the Study of Addiction: Genetics and Environment (SAGE, dbGaP Study Accession:

1595 phs000092.v1.p1). Control subjects did not have alcohol dependence or drug dependence  
1596 diagnoses; however, mood disorders were not an exclusion criterion.

1597 **McQuillin, A; Gurling, H | 18317468 [PGC1] | UCL (University College London), London,**  
1598 **UK | bip\_uclo\_eur**

1599 The UCL sample comprised Caucasian individuals who were ascertained and received clinical  
1600 diagnoses of bipolar 1 disorder according to UK National Health Service (NHS) psychiatrists at  
1601 interview using the categories of the International Classification of Disease version 10. In  
1602 addition bipolar subjects were included only if both parents were of English, Irish, Welsh or  
1603 Scottish descent and if three out of four grandparents were of the same descent. All volunteers  
1604 read an information sheet approved by the Metropolitan Medical Research Ethics Committee  
1605 who also approved the project for all NHS hospitals. Written informed consent was obtained  
1606 from each volunteer. The UCL control subjects were recruited from London branches of the  
1607 National Blood Service, from local NHS family doctor clinics and from university student  
1608 volunteers. All control subjects were interviewed with the SADS-L to exclude all psychiatric  
1609 disorders.

1610 **Craddock, N; Jones, I; Jones, L | [ICCBD] | Cardiff and Worcester, UK (ICCBD-BDRN) |**  
1611 **bip\_icuk\_eur**

1612 Cases were all over the age of 17 yr, living in the UK and of European descent. Cases were  
1613 recruited via systematic and not systematic methods as part of the Bipolar Disorder Research  
1614 Network project ([www.bdrn.org](http://www.bdrn.org)), provided written informed consent and were interviewed  
1615 using a semi-structured diagnostic interview, the Schedules for Clinical Assessment in  
1616 Neuropsychiatry. Based on the information gathered from the interview and case notes review,  
1617 best-estimate lifetime diagnosis was made according to DSM-IV. Inter-rater reliability was

1618 formally assessed using 20 randomly selected cases (mean  $\kappa$  Statistic = 0.85). In the current  
1619 study we included cases with a lifetime diagnosis of DSM-IV bipolar disorder or schizo-affective  
1620 disorder, bipolar type. The BDRN study has UK National Health Service (NHS) Research Ethics  
1621 Committee approval and local Research and Development approval in all participating NHS  
1622 Trusts/Health Boards. Controls were part of the Wellcome Trust Case Control Consortium  
1623 common control set, which comprised healthy blood donors recruited from the UK Blood  
1624 Service and samples from the 1958 British Birth Cohort. Controls were not screened for a history  
1625 of mental illness. All cases and controls were recruited under protocols approved by the  
1626 appropriate IRBs. All subjects gave written informed consent.

1627 **Ophoff, RA | Not Published | Netherlands | bip\_ucla\_eur**

1628 The case sample consisted of inpatients and outpatients recruited through psychiatric hospitals  
1629 and institutions throughout the Netherlands. Cases with DSM-IV bipolar disorder, determined  
1630 after interview with the SCID, were included in the analysis. Controls were collected in parallel  
1631 at different sites in the Netherlands and were volunteers with no psychiatric history after  
1632 screening with the (MINI). Ethical approval was provided by UCLA and local ethics committees  
1633 and all participants gave written informed consent.

1634 **Paciga, S | [PGC1] | USA (Pfizer) | bip\_pf1e\_eur**

1635 This sample comprised Caucasian individuals recruited into one of three Geodon (ziprasidone)  
1636 clinical trials (NCT00141271, NCT00282464, NCT00483548). Subjects were diagnosed by a  
1637 clinician with a primary diagnosis of Bipolar 1 Disorder, most recent episode depressed, with or  
1638 without rapid cycling, without psychotic features, as defined in the DSM-IV-TR (296.5x) and  
1639 confirmed by the MINI (version 5.0.0). Subjects also were assessed as having a HAM-D-17  
1640 total score of >20 at the screening visit. The trials were conducted in accordance with the

1641 protocols, International Conference on Harmonization of Good Clinical Practice Guidelines, and  
1642 applicable local regulatory requirements and laws. Patients gave written informed consent for the  
1643 collection of blood samples for DNA for use in genetic studies.

1644 **Pato, C | [ICCBD] | Los Angeles, USA (ICCBD-GPC)| bip\_usc2\_eur**

1645 Genomic Psychiatry Consortium (GPC) cases and controls were collected via the University of  
1646 Southern California healthcare system, as previously described. Using a combination of focused,  
1647 direct interviews and data extraction from medical records, diagnoses were established using the  
1648 OPCRIT and were based on DSM-IV-TR criteria. Age and gender-matched controls were  
1649 ascertained from the University of Southern California health system and assessed using a  
1650 validated screening instrument and medical records.

1651 **Scott, L; Myer, RM; Boehnke, M | 19416921 [PGC1] | Michigan, USA (Pritzker and**  
1652 **NIMH) | bip\_mich\_eur**

1653 The Pritzker Neuropsychiatric Disorders Research Consortium (NIMH/Pritzker) case and  
1654 controls samples were from the NIMH Genetics Initiative Genetics Initiative Repository. Cases  
1655 were diagnosed according to DMS-III or DSM-IV criteria using diagnostic interviews and/or  
1656 medical record review. Cases with low confidence diagnoses were excluded. From each wave 1-  
1657 5 available non-Ashkenazi European-origin family, two BD1 siblings were included when  
1658 possible and the proband was preferentially included if available (n=946 individuals in 473  
1659 sibling pairs); otherwise a single BD1 case was included (n=184). The bipolar sibling pairs were  
1660 retained within the NIMH/Pritzker sample when individuals in more than one study were  
1661 uniquely assigned to a study set. Controls had non-Ashkenazi European-origin, were aged 20-70  
1662 years and reported no diagnosis with or treatment for BD or schizophrenia, and that they had not  
1663 heard voices that others could not hear. Individuals with suspected major depression were

1664 excluded based on answers to questions related to depressive mood. NIMH controls were further  
1665 selected as the best match(es) to NIMH cases based on self-reported ancestry.

1666 **Sklar, P; Smoller, J | 18317468 [PGC1] | USA (STEP1) | bip\_stp1\_eur**

1667 The Systematic Treatment Enhancement Program for Bipolar Disorder (STEP-BD) was a seven-  
1668 site, national U.S., longitudinal cohort study designed to examine the effectiveness of treatments  
1669 and their impact on the course of BD that enrolled 4,361 participants who met DSM-IV criteria  
1670 for BD1, BD2, bipolar not otherwise specified (NOS), schizoaffective manic or bipolar type, or  
1671 cyclothymic disorder based on diagnostic interviews. From the parent study, 2,089 individuals  
1672 who were over 18 years of age with BD1 and BD2 diagnoses consented to the collection of  
1673 blood samples for DNA. BD samples with a consensus diagnosis of BD1 were selected for  
1674 inclusion in STEP1. Two groups of controls samples from the NIMH repository were used. One  
1675 comprised DNA samples derived from US Caucasian anonymous cord blood donors. The  
1676 second were controls who completed the online self-administered psychiatric screen and were  
1677 ascertained as described above, by Knowledge Networks Inc. For the second sample of controls  
1678 only those without history of schizophrenia, psychosis, BD or major depression with functional  
1679 impairment were used.

1680 **Sklar, P; Smoller, J | 18711365 [PGC1] | USA (STEP2) | bip\_stp2\_eur**

1681 The STEP2 sample included BD-1 and BD-2 samples from the STEP-BD study described above  
1682 along with BD-2 subjects from UCL study also described above. The controls samples for this  
1683 study were from the NIMH repository as described above for the STEP1 study.

1684

1685 **European ancestry, trio design**

1686 *Schizophrenia*

1687 **Kirov, G: Owen M | 22083728| Bulgaria | ms.scz\_butr\_eur**

1688 Families from Bulgaria were recruited if a proband had schizophrenia or schizoaffective  
1689 disorder, both parents were available, and all members of the trio agreed to participate in the  
1690 study. Recruitment took place between 1999 and 2004 in several psychiatric hospitals in  
1691 Bulgaria. Ethical Committee approval was obtained from each of these hospitals. All probands  
1692 and all parents received an Information Sheet and signed Informed Consent Forms. All  
1693 participants had attended mainstream schools, which at the time in Bulgaria, excluded people  
1694 with mental retardation. Probands were either in- or out-patients at the time of the study but each  
1695 had a history of hospitalization. A team of psychiatrists was trained in using the rating scales and  
1696 methods of the study. We used the SCAN instrument to perform an interview for psychotic and  
1697 mood symptoms. This instrument has been translated into Bulgarian and validated by one of its  
1698 authors (A. Jablensky). Consensus diagnoses were made according to DSM-IV criteria on the  
1699 basis of an interview and inspection of hospital notes by two clinicians. If consensus was not  
1700 attained, the patient was re-interviewed by a research interview trained clinician and was  
1701 excluded if consensus could still not be reached. In addition, approximately 23% of the sample  
1702 was selected at random and re-interviewed by a research interview trained clinician. Hospital  
1703 notes were also collected for affected relatives in order to confirm diagnoses.

1704 **Levinson, D | 22885689 | Six countries | ms.scz\_lemu\_eur**

1705 Schizophrenia cases were included from the family sample of European-ancestry pedigrees  
1706 described by Levinson et al. Participants and their families in this trio study, probands were  
1707 ascertained and recruited from different clinical settings (e.g. inpatients, outpatients and  
1708 community facilities) in six countries (Australia, France, Germany, Ireland, UK, and the US).  
1709 (Unrelated individuals were included as part of a case-control design, see Levinson, D,



1710 scz\_lacw\_eur above.) Diagnoses were established using semi-structured interviews, psychiatric  
1711 records and informant reports. Case probands were diagnosed with schizophrenia or  
1712 schizoaffective disorder according to DSM-III-R criteria. The trio-based analysis included  
1713 families where there was at least one affected proband and two available parents. Each affected  
1714 sibling in such families was included, with the parents, as an independent trio. All protocols were  
1715 approved by local IRBs, and all cases provided written informed consent.

1716 **Kirov, G: Owen, M | Not Published | Bulgaria | ms.scz\_uktr\_eur**

1717 All cases and parents were recruited from UK and had a history of hospitalization for treatment  
1718 of schizophrenia. Diagnosis was confirmed following a SCAN interview and review of case  
1719 notes followed by consensus diagnosis according to DSM-IV criteria. The samples were  
1720 genotyped at the Broad Institute. All participants gave written informed consent and the study  
1721 was approved by local ethics committees at the participating centers. The samples were  
1722 genotyped at the Broad Institute.

1723

#### 1724 **Genotype Quality Control**

1725 To ensure independence of the data sets, individuals were excluded until no individual showed a  
1726 relatedness ( $\pi_{\text{hat}}$ ) value greater than 0.2 to any other individual in the collection, while  
1727 preferentially keeping the case over the control for case-control related pairs. In total 1,795 BD  
1728 cases, 1,165 SCZ cases and 27,274 controls were removed (most of which were previously  
1729 known), leaving 20,129 BD cases 33,426 SCZ cases and 54,065 controls for the final meta-  
1730 analysis.

1731 For analyses directly comparing BD and SCZ, we matched cases from both phenotypes on  
1732 genotyping platform and ancestry, resulting in 15,270 BD cases versus 23,585 SCZ cases.

Hence, we were able to match 76% of BD cases and 71% of SCZ cases for this case vs case analysis.

Among our entire dataset, 44% of the sample was female, 51% was male and 5% were unreported by the collection site. This work focused explicitly on the autosomes and sought maximal power across the analyses, sex was not used except for during quality control and sex-specific analyses were not performed in this effort. Individual ages were not provided. For a subset of cases, we had information for age of onset which were used in subphenotype specific analyses only.

### **Sub-phenotype Description**

BD sub-phenotypes were collected by each study site using a combination of diagnostic instruments, case records and participant interviews. Ascertainment details for each study site are described in the supplementary data of the PGC Bipolar Working Group paper(Stahl et al., 2017). The selection of phenotypes for collection by this group was determined by literature searches in order to determine phenotypes with prior evidence for heritability. It was further refined dependent on the availability of phenotype data across a range of study sites and the consistency by which the phenotypes were defined. Schizophrenia subphenotypes represent quantitative traits extracted using factor analysis from a set of standard psychiatric assessments and represent four symptom dimensions (manic, depressive, positive and negative). These subphenotypes were used previously(Ruderfer et al., 2014) but in this work we have increased the sample size with additional cohorts being added.

### **METHOD DETAILS**

## QUANTIFICATION AND STATISTICAL ANALYSIS

### Quality Control, Imputation, Association Analysis and Polygenic Risk Score Testing

Quality control and imputation were performed on each of the study cohort datasets ( $n=81$ ), according to standards established by the Psychiatric Genomics Consortium (PGC). The quality control parameters for retaining SNPs and subjects were: SNP missingness  $< 0.05$  (before sample removal); subject missingness ( $p < 0.02$ ); autosomal heterozygosity deviation ( $|F_{het}| < 0.2$ ); SNP missingness  $< 0.02$  (after sample removal); difference in SNP missingness between cases and controls  $< 0.02$ ; and SNP Hardy-Weinberg equilibrium ( $p > 10^{-6}$  in controls or  $p > 10^{-10}$  in cases). Genotype imputation was performed using the pre-phasing/imputation stepwise approach implemented in IMPUTE2(Howie et al., 2011) / SHAPEIT(Delaneau et al., 2013) (chunk size of 3 Mb and default parameters). The imputation reference set consisted of 2,186 phased haplotypes from the full 1000 Genomes Project dataset (August 2012, 30,069,288 variants, release “v3.macGT1”), all variants align to human genome build 19 (hg19). After imputation, we used the best guess genotypes (genotype probability  $> 0.8$ ), for further robust relatedness testing and population structure analysis. Here we required very high imputation quality (INFO  $> 0.8$ ) and low missingness ( $< 1\%$ ) for further quality control. After linkage disequilibrium (LD) pruning ( $r^2 < 0.02$ ) and frequency filtering (MAF  $> 0.05$ ), there were 14,473 autosomal SNPs in the data set. Principal component estimation was done with the same collection of autosomal SNPs. We tested the first 20 principal components for phenotype association (using logistic regression with study indicator variables included as covariates) and evaluated their impact on the genome-wide test statistics using  $\lambda$ . Thirteen principal components

namely 1,2,3,4,5,6,7,8,10,12,15,18,20 were included in all association analyses ( $\lambda=1.45$ ). Analytical steps were repeated for SCZ vs BD analysis.

We performed four main association analyses (Figure 1), i.e. (i) GWAS of BD and SCZ as a single combined case phenotype, as well as disorder-specific GWAS using independent control sets in (ii) BD cases vs BD controls and (iii) SCZ cases vs SCZ controls, and (iv) association analysis of SCZ cases vs BD cases. For all GWS loci from the GWAS of BD and SCZ vs controls we identified any GWS loci within 1Mb from the extent of the locus in the previously published PGC SCZ vs controls (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014) and the most recent PGC GWAS of BD vs controls (Stahl et al., 2017) and performed conditional analysis. Specifically, we transformed the genotype probabilities of the disease variant into dosages and used it as an additional covariate for the association analysis for the BD+SCZ vs controls index SNP. This was done within each cohort and an OR based inverse SE weighted meta-analysis was performed for the final result. All datasets were included except for those with trios.

#### **Summary-data-based Mendelian Randomization (SMR)**

SMR (Zhu et al., 2016) is a method that integrates summary level GWAS data with gene expression quantitative trait loci (eQTL) identified in independent data sets. This integration aims to identify variants that have pleiotropic effects on expression of a given gene and the phenotype. While significant findings may indeed reflect a causal path from variant to phenotype through expression, it is impossible to discern statistically between pleiotropy and causality. However, the method can remove linkage as driving the result, and uses the data available to prioritise amongst genes in genomic regions that show association with disease. We used SMR

as a statistical fine-mapping tool applied to the SCZ vs BD GWAS results to identify loci with strong evidence of causality via gene expression. SMR analysis is limited to significant ( $FDR < 0.05$ ) cis SNP-expression quantitative trait loci (eQTLs) with  $MAF > 0.01$ . eQTLs passing these thresholds were combined with GWAS results in the SMR test, with significance ( $p_{SMR}$ ) reported at a Bonferroni-corrected threshold for each eQTL data set. The eQTL architecture may differ between genes. For example, through LD, many SNPs can generate significant associations with the same gene, but in some instances multiple SNPs may be independently associated with the expression of a gene. After identification of significant SNP-expression-trait association through the SMR test, a follow-up heterogeneity test aims to prioritize variants by excluding regions for which there is conservative evidence for multiple causal loci ( $p_{HET} < 0.05$ ). SMR analyses were conducted using eQTL data from whole peripheral blood(Westra et al., 2013), dorsolateral prefrontal cortex generated by the CommonMind Consortium<sup>8</sup>, and 11 brain sub-regions from the GTEx consortium(Consortium, 2015).

## **Regional joint GWAS**

Summary statistic Z-scores were calculated for each marker in each of the four main GWAS results, using the logistic regression coefficient and its standard error. Rare SNPs ( $MAF < 0.01$ ), and SNPs with a low INFO score ( $< 0.3$ ) in either dataset were removed. The causal variant relationships between SCZ and BD were investigated using the Bayesian method software *pw-gwas* (v0.2.1), with quasi-independent regions determined by estimate LD blocks in an analysis of European individuals ( $n=1,703$ )(Berisa and Pickrell, 2015; Pickrell et al., 2016). Briefly, *pw-gwas* takes a Bayesian approach to determine the probability of five independent models of association. (1) There is no causal variant in BD or SCZ; (2) a causal variant in BD, but not SCZ

(3); a causal variant in SCZ, but not BD; (4) a shared causal variant influencing both BD and SCZ; (5) two causal variants where one influences BD, and one influences SCZ (Figure 2). The posterior probability of each model is calculated using model priors, estimated empirically within pw-gwas. Regions were considered to support a particular model when the posterior probability of the model was greater than 0.5.

### **Regional SNP-heritability estimation**

We calculated local SNP-heritability independently for SCZ and BD using the Heritability Estimator from Summary Statistics (HESS) software(Shi et al., 2016) for each of the independent regions defined above. The sum of these regional estimates is the total SNP-heritability of the trait. To calculate local SNP-heritability HESS requires reference LD matrices representative of the population from which the GWAS samples were drawn. We utilized the 1000 genomes European individuals as the reference panel(The 1000 Genomes Project Consortium, 2015). Unlike pw-gwas(Pickrell et al., 2016), HESS does not assume that only one causal variant can be present in each region.

### **DATA AND SOFTWARE AVAILABILITY**

Summary statistics from GWAS are publically available at <https://www.med.unc.edu/pgc/results-and-downloads/downloads>.