

2010-05-10

# CBD & D9-THC - the two faces of cannabis

Zeissler,

<http://hdl.handle.net/10026.1/12049>

---

Cell Science

---

*All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Please cite only the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author.*

## CBD & $\Delta^9$ -THC - the two faces of cannabis

Marie-Louise Zeissler, John P. Zajicek & Camille B. Carroll



Peninsula College of Medicine & Dentistry, University of Plymouth, Dept Clinical Neurobiology, John Bull Building, Tamar Science Park, Plymouth, Devon, PL6 8BU, UK.

Received 10th May © Cell Science 2010

**Cannabidiol (CBD) and  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) are phytocannabinoids produced by the plant *Cannabis sativa* which have both shown promising therapeutic value as anti-inflammatory and neuroprotective agents in a variety of neurological disorders. However, the molecular mechanisms involved in their protective function are thought to be distinctly different probably due to their divergent receptor pharmacology.  $\Delta^9$ -THC is a potent agonist at the cannabinoid receptors 1 and 2 through which it is thought to modulate synaptic plasticity and inflammatory responses respectively. CBD on the other hand exhibits negligible affinity for both receptors prompting research into cannabinoid receptor independent mechanisms of action such as modulation of endogenous cannabinoid tone and calcium signalling. This review aims to give an introduction into the pharmacological and biological effects of  $\Delta^9$ -THC and CBD with particular reference to neurological disease.**

### Introduction

After alcohol and tobacco, cannabis is one of the most commonly used recreational drugs worldwide. Although its deleterious effects on cognitive brain function have been well described, cannabis has also been shown to have potential as an anti-inflammatory, anxiolytic and neuroprotective agent. Cannabidiol (CBD) and  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) are the two most abundant compounds out of over 70 phytocannabinoids produced by the plant *Cannabis sativa* [1]. Because of its apparent psychoactivity,  $\Delta^9$ -THC was the first cannabinoid to be studied in detail and led to the discovery of the first cannabinoid receptors, CB1 and CB2 [2, 3]. Synthetic as well as endogenous cannabinoids (endocannabinoids) are defined by their characteristic ability to bind to either of these receptors whereas phytocannabinoids are identified by carrying a distinct C<sub>21</sub> group [1].

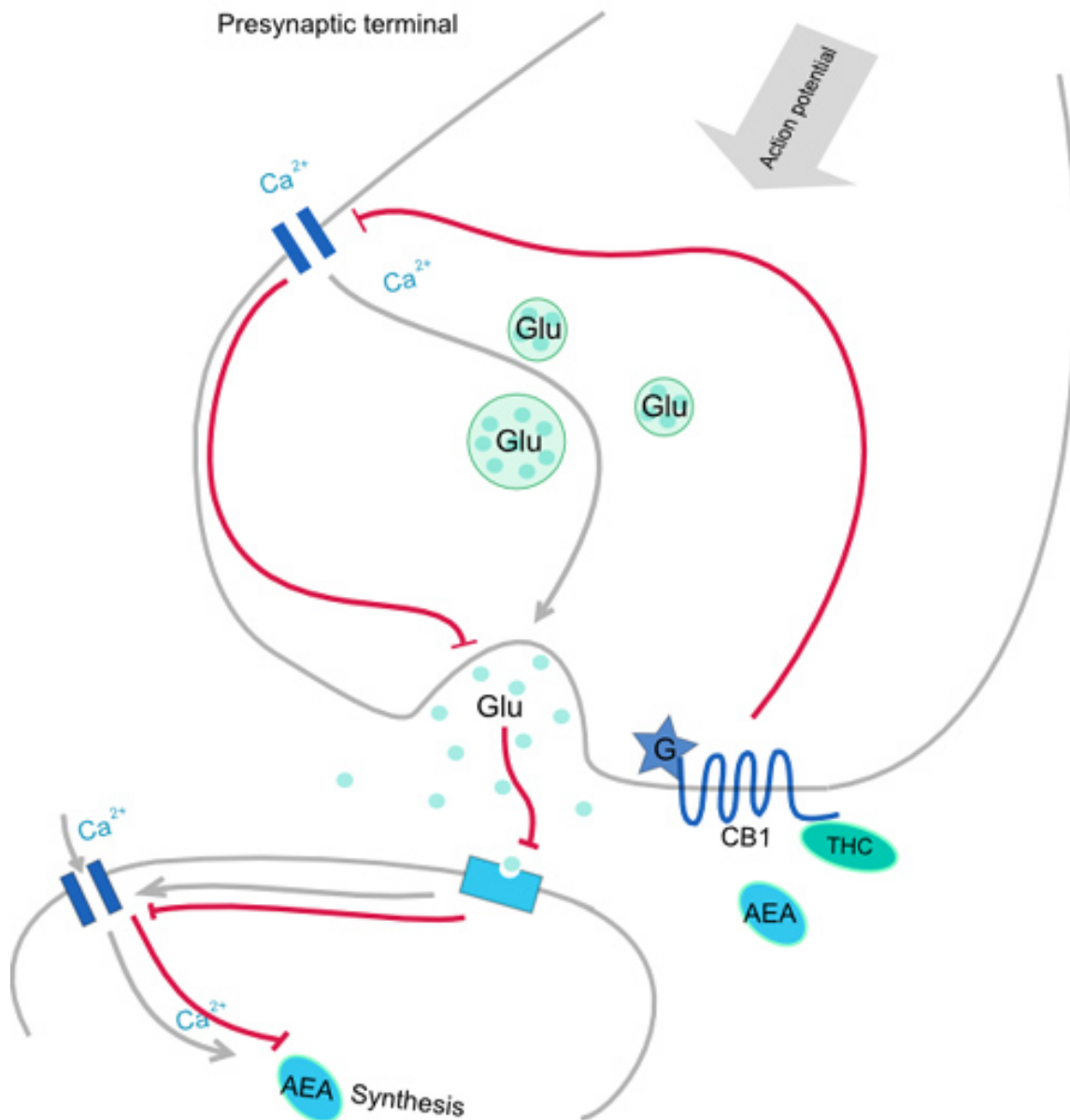
The therapeutic potential of cannabinoids has been widely recognised and they have been clinically trialled in the treatment of a range of neurological symptoms and disorders including

spasticity [4], neurogenic pain [5], dyskinesia [6], tremor [7], schizophrenia [8, 9] and psychosis [10-12].

Although both  $\Delta^9$ -THC and CBD have anti-inflammatory and neuroprotective properties, their effects are thought to be mediated through distinctly different pathways. A key difference between  $\Delta^9$ -THC and CBD is that while  $\Delta^9$ -THC readily agonises CB1 and CB2 receptors, CBD exhibits negligible affinity for both. Furthermore, the binding properties of  $\Delta^9$ -THC and CBD differ for the newly identified G-protein receptor GPR55 receptor which is agonised by both the endocannabinoid anandamide (AEA) and  $\Delta^9$ -THC but antagonised by CBD [13]. In this review, we will examine the differential pharmacological and biological effects of  $\Delta^9$ -THC and CBD, mainly as a result of this discrepancy, with particular reference to neurological disease. The role of GPR55 in neuroprotective mechanisms is thus far unknown and will therefore not be further discussed in this review.

## **2. The psychoactivity and cognitive effects of $\Delta^9$ -THC**

The potentially long lasting detrimental effects on cognitive function induced by the use of cannabis are thought to be due to  $\Delta^9$ -THC, its major psychoactive component [14-16]. Concern regarding psychotropic and cognitive side-effects has limited therapeutic use of cannabinoids in the treatment of neurodegenerative diseases. It has been found that  $\Delta^9$ -THC can influence memory formation, impair psychomotor control, trigger anxiety and psychosis and affect social interaction [17, 18]. It has been shown that regular exposure of rats to  $\Delta^9$ -THC as well as a single injection of as little as 1  $\mu\text{g}/\text{kg}$   $\Delta^9$ -THC into the cerebellum can lead to significant impairment of spatial learning [19, 20]. It is widely accepted that these psychoactive activities are CB1 receptor mediated as behavioural symptoms as well as cognitive impairment can be mimicked by CB1 receptor agonists whereas specific antagonists such as SR141716A are protective [14, 19, 21-23]. Activation of CB1 receptors modulates long term potentiation and depression (LTP and LTD) and thus has a profound effect on synaptic plasticity and memory formation. This is in accordance with reports of high CB1 receptor expression levels in brain regions associated with cognitive function [24].  $\Delta^9$ -THC has been reported to manipulate LTP and LTD directly through CB1 receptor activation but may also affect the retrograde signalling of endogenous CB1 ligands (endocannabinoids) [25, 26]. When acting directly through the CB1 receptor, a depression of neurotransmitter release is initiated as a result of receptor coupling to inhibitory G-proteins preventing the activation of voltage-gated  $\text{Ca}^{2+}$  [27-29] and  $\text{K}^+$  channels [30, 31]. The nature of the long term effect is dependent on the neurotransmitter released at the synapse. Thus CB1 receptor stimulation leads to inhibition of LTD in GABAergic synapses and inhibition of LTP at glutamatergic synapses [32-35].



**Figure 1.** Activation of CB1 at the presynaptic terminal by THC leads to an inhibition of neurotransmitter release and postsynaptic endocannabinoid; Glutamate (glu), G-protein (G).

Endocannabinoids can be released either as a result of postsynaptic depolarisation stimulated by the resultant Ca<sup>2+</sup> influx (depolarization induced suppression of inhibition (DSI) or depolarisation induced suppression of excitation (DSE)) or through activation of postsynaptic G-coupled receptors such as the metabotropic glutamate receptor 1 (mGlu1). Using hippocampal and nucleus accumbens tissue slices it has been shown that a single injection of  $\Delta^9$ -THC leads to a short term but significant reduction of DSI and DSE as well as mGlu1-mediated LTD [25]; this effect was rescued with the specific CB1 receptor antagonist SR141716A [25]. Interestingly, chronic exposure to  $\Delta^9$ -THC led to down-regulation of CB1 receptor expression and reduced coupling efficiency, thereby desensitising the brain to activation of the receptor by the drug as well as its endogenous ligands [36, 37]. Additionally,  $\Delta^9$ -THC can lead to a desensitisation of the CB1 receptor even if applied in doses that do not trigger internalisation of the receptor [25, 37]. Therefore, given their central role in the mediation of psychoactivity, it has been proposed that

the activity of CB1 receptors is tightly linked to tolerance of and dependence on cannabinoids.

The molecular differences observed following acute versus chronic administration could be the major factor in the modulation of cannabinoid tolerance as well as dose-dependent differences [38]. These differences are particularly pronounced in the activation of downstream signalling pathways. The extracellular-regulated kinase (ERK), cyclic adenosine monophosphate (cAMP) and mitogen-activated protein kinase (MAPK) pathways all lead to the phosphorylation of CREB (cAMP-response element-binding protein) which is an activator of transcription. Low doses of  $\Delta^9$ -THC activate ERK 1/2, which play an important role in the regulation of cell survival, whereas high doses do not [20]. Similarly, acute treatment significantly increases CREB phosphorylation [33, 39] whereas chronic treatment with  $\Delta^9$ -THC reduces CREB phosphorylation, which mediates transcription by binding to cAMP response elements (CRE) via CB1 receptors, both in the hippocampus and the cerebellum [38]. However, both chronic and acute treatments with  $\Delta^9$ -THC lead to an increase in ERK1/2 protein levels and activation of ERK1/2 in the dorsal striatum, nucleus accumbens and the hippocampus [38, 40, 41]. Thus CB1 activation leads to ERK1/2 activation [41]. During the process of desensitisation of the receptor, CB1 is phosphorylated, inhibiting G-protein coupling and promoting  $\beta$ -arrestin binding to the receptor which in turn initiates internalisation [42]. Phosphorylation of the CB1 receptor is critical for inactivating ERK1/2. By contrast, internalisation of the receptor has no effect on ERK1/2 activation patterns, a phenomenon that may be cell specific [43, 44].

Unlike  $\Delta^9$ -THC, CBD does not appear to affect behavioural and cognitive brain function, which is not surprising considering the key role CB1 plays in these mechanisms. However, CBD may be able to influence the psychoactivity of  $\Delta^9$ -THC via as yet unknown mechanisms [45]. For example CBD inhibits up to 50% of the breakdown of  $\Delta^9$ -THC into its metabolites by hepatic microsomes [46]. In the 1970s a series of studies reported the ability of CBD to attenuate the detrimental effect of  $\Delta^9$ -THC on cognitive function, as well as  $\Delta^9$ -THC-induced symptoms such as anxiety and paranoia [47-49]. Additionally, CBD is able to reverse perceptive changes caused by nabilone, a structural analogue of  $\Delta^9$ -THC [50]. Furthermore, mice treated with CBD-supplemented  $\Delta^9$ -THC performed better in spatial memory tests than when treated with  $\Delta^9$ -THC alone, whereas CBD alone had no effect [45, 51]. Dosage seems to be important as the modulating effects of CBD on  $\Delta^9$ -THC seem to be dependent on the ratio of CBD to  $\Delta^9$ -THC [12, 51, 52]. A recent study using functional magnetic resonance imaging (fMRI) in humans confirmed that CBD and  $\Delta^9$ -THC have distinctly opposite effects on neuronal activity during cognitive tasks [53]. CBD is thus considered a possible treatment for cognitive impairments associated with marijuana consumption.

CBD has also been suggested as a treatment of psychosis such as occurs in schizophrenia and drug-induced psychosis. A pilot study found that CBD significantly reduced levodopa-induced psychosis in Parkinson's disease patients [10] and may thus be a useful treatment as it was well tolerated with almost no side effects. However, it was found that CBD monotherapy for treatment resistant schizophrenia had no effect in 2 out of three cases [54].

Commonly, anti-psychotic drugs induce *c-fos* expression in the limbic system which is mimicked by CBD providing molecular evidence of the anti-psychotic activity of CBD [55].

On a cellular level the ability of CBD to attenuate or even reverse the psychoactive effects of  $\Delta^9$ -THC may be linked to its CB1 receptor pharmacology. Although CBD is a weak agonist at the CB1 receptor it has been repeatedly reported that CBD may be a partial antagonist at the receptor, marked by the displacement of the CB1 receptor agonists CP559540 and Win55212-2, but lacking receptor activation. Furthermore, agonists at the CB1 receptor are usually able to increase binding of the G-protein GTP $\gamma$ S. CBD inhibits the binding of GTP $\gamma$ S to mouse brain membranes thus displaying signs of inverse agonism at the receptor at a concentration of 10 $\mu$ M [56-58]. Although this finding remains controversial, as others have reported GTP $\gamma$ S binding to be unaffected by cannabidiol (reviewed in [59]), this may provide a possible mechanism by which CBD is able to attenuate some of the CB1 receptor-mediated psychoactive properties of  $\Delta^9$ -THC.

## Neuroprotection

The diverse mechanisms of action underlying the neuroprotective properties of cannabinoids can be divided into three broad categories:

- (1) Receptor-mediated. In the case of  $\Delta^9$ -THC this can be either CB1 or CB2 mediated, whereby CB1 activation is usually protective against excitotoxicity [60, 61] and CB2 activation is associated with an anti-inflammatory response.
- (2) Anti-oxidant activity. Both CBD and  $\Delta^9$ -THC are protective, at least in part, due to their inherent antioxidant activity conferred by the structural presence of a phenolic ring.
- (3) Modulation of endocannabinoid tone and other mechanisms. There is evidence that CBD, which has negligible affinity at the CB1 and CB2 receptor, may affect endocannabinoid tone and intracellular calcium signalling, which could be relevant to its neuroprotective effects.

### (1) Receptor-mediated neuroprotection

Studies in a range of *in vivo* and *in vitro* excitotoxicity models have suggested that the neuroprotective effect of cannabinoids may be mediated via the CB1 receptor [62-69]. CB1 receptors are widely expressed throughout the brain [24, 70, 71] where they co-localise with glutamatergic and GABAergic synapses [70, 72]. In motor diseases such as Huntington's disease (HD) and Parkinson's disease (PD) CB1 expression pattern changes, implicating an important role for the receptor [73-75].

Excitotoxicity occurs as a result of intense synaptic firing which leads to changes in dendrite morphology, synapse loss and can promote cell death. Even non-toxic increases in synaptic activity can cause an increase in dendrites and alter the shape of synaptic spines which can be crucial in pathological processes [61, 76-78]. One mechanism by which  $\Delta^9$ -THC may be exerting its protective effects is via CB1 receptor-mediated presynaptic inhibition of glutamate release and suppression of excitotoxic mechanisms [28, 64, 67], although another study found a direct post-synaptic CB1 receptor-mediated protective effect of  $\Delta^9$ -THC against kainate-induced

excitotoxicity [65].

Excitotoxicity is thought to be a relevant mechanism of cell death in conditions such as epilepsy, multiple sclerosis and cerebral ischaemia, in models of which CB1-receptor-mediated protective effects have been demonstrated [63, 66, 69]. However, the therapeutic use of  $\Delta^9$ -THC or other CB1 agonists in excitotoxicity-induced neurodegeneration may be limited by agonist-induced desensitisation of the CB1 receptor due to internalisation as well as reduced coupling efficiency [36, 37], which has been shown to occur within 24hrs in a cell culture model [60]. Excitotoxicity has also been postulated to be relevant in the pathogenesis of neurodegenerative conditions such as motor neurone disease [79]. However, in the case of Parkinson's disease (PD), it is likely that other mechanisms of cell death are more relevant and in this case the protective effects of cannabinoids appear to be non-CB1 receptor-mediated [80-83].

The role of the CB2 receptor in mediating neuro-protective effects via modulation of inflammation is discussed in Section 4 below.

## (2) Antioxidant effects

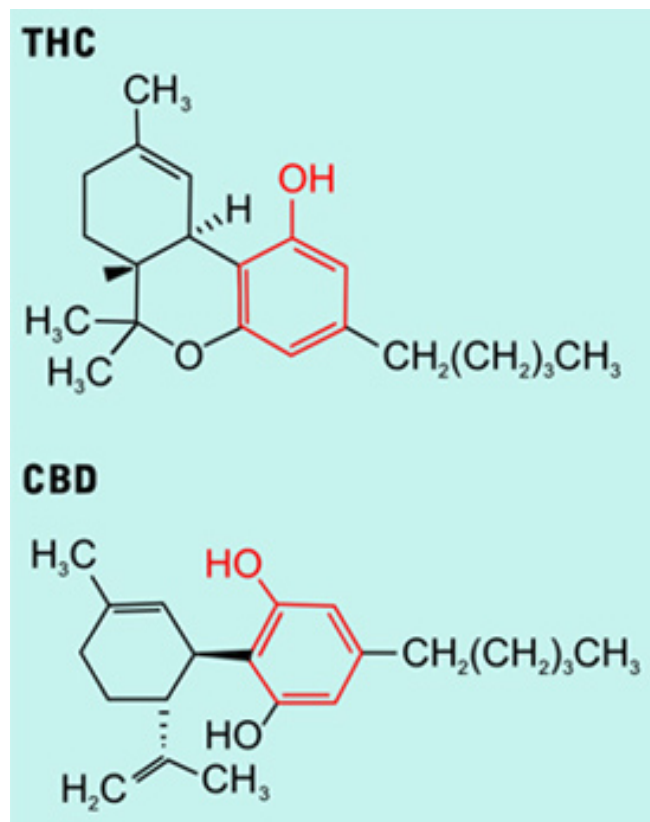
It is likely that some of the neuroprotective effects of  $\Delta^9$ -THC are non-CB1 receptor mediated. This has particularly been demonstrated in models of PD, where *in vivo* and *in vitro* studies have demonstrated neuroprotective effects of  $\Delta^9$ -THC that could not be blocked by specific CB1 receptor antagonists [80-82]. A *Drosophila* model of PD also implicated non-receptor mediated mechanisms of neuroprotection as CB1 receptor agonists were neuroprotective even though *Drosophila* do not express CB1 or CB2 receptors [83].

**Figure 2.** Antioxidant properties of THC and CBD are conferred by a phenol ring structure (red).

Commonly, neurodegenerative diseases lead to the formation of reactive oxygen species (ROS). Both CBD and  $\Delta^9$ -THC as well as many other cannabinoids are inherently antioxidant due to the presence of a phenolic ring in their chemical structure [80, 81, 83, 84] (see figure 2) and it has been suggested that this underlies their neuroprotective effects.

## (3) Modulation of endocannabinoid tone and other mechanisms

Modulation of endocannabinoid tone may be a relevant neuroprotective mechanism. In some conditions an increase in endocannabinoid level has been observed,

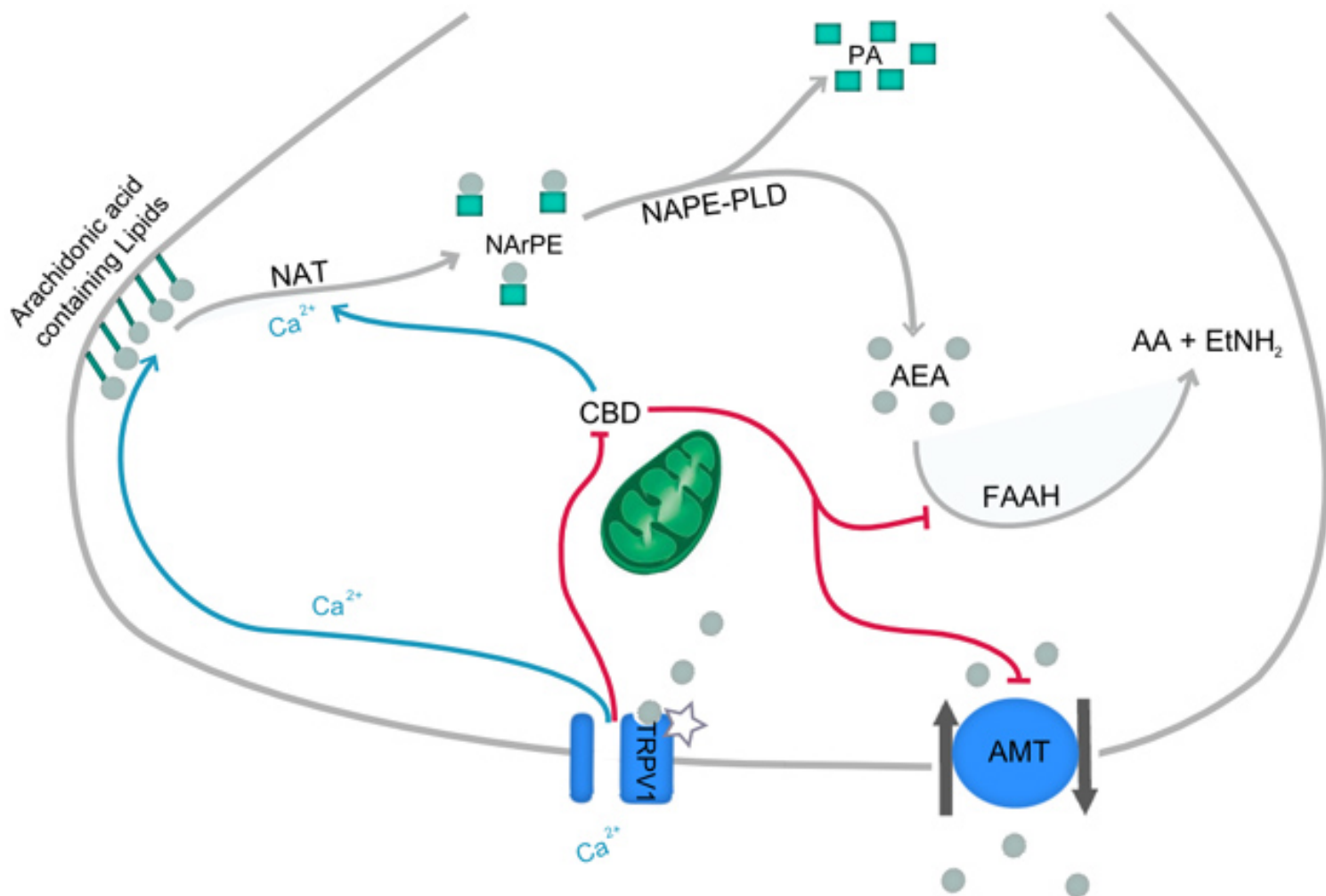


and it is thought that this may be a protective response. For example in a model of epilepsy, kainate-induced excitotoxicity acutely increased anandamide levels within the mouse brain and application of the CB1 receptor antagonist, SR141716A, increased seizure severity, suggesting a CB1-receptor-mediated protective effect of AEA against excitotoxicity. Furthermore, the AEA uptake inhibitor UCM707 was also protective [69]. Increase in endocannabinoid level has also been demonstrated in PD [85, 86].

In contrast, in animal models of another neurodegenerative disease, Huntington's disease (HD), endocannabinoid levels are markedly reduced [87]. In this condition transient receptor potential cation channel, subfamily V, member 1 (TRPV1) receptor activation has been shown to reduce motor symptoms such as hyperkinesia [88]. Although a neuroprotective effect may be exerted by CB1 receptor agonists as described above, there is loss of CB1 receptor-expressing neurones in the later stages of the disease [75, 88-90]; however, AEA uptake inhibitors as well as inhibitors of the hydrolysing enzyme FAAH have been shown to be neuroprotective [91]. This illustrates that in neurodegenerative conditions such PD and HD, cannabinoids may exert both neuroprotective effects as well as relieving symptoms - effects mediated by different receptors or non-receptor dependent mechanisms. Greater clinical benefit may therefore be achieved by modulating endocannabinoid tone rather than targeting specific receptors.

Although CBD has a limited direct effect on CB1 receptors due to its low affinity, it modulates endocannabinoid tone by increasing AEA levels which in turn may act at the CB1 receptor. However, the exact mechanism by which CBD modulates endocannabinoid tone remains unclear. One possibility is that CBD acts via the intracellular vanilloid receptor, TRPV1, which is a non-selective cation channel that facilitates the entry of  $Ca^{2+}$  into the cell [92]. Activation of the TRPV1 receptor can trigger  $Ca^{2+}$  dependent synthesis of endocannabinoids such as anandamide (AEA) which are also ligands at the receptor [93-95]. CBD leads to an influx in intracellular  $Ca^{2+}$ . However, there is evidence that CBD may not be acting via the TRPV1 receptor as the  $Ca^{2+}$  rise it generates is not blocked by the specific TRPV1 antagonist capsazepine [96]. On the contrary, the  $Ca^{2+}$  influx significantly increased when the TRPV1 receptor was blocked indicating that TRPV1 receptor activation causes downregulation of the CBD-induced  $Ca^{2+}$  rise [96]. Thus AEA, which is a potent activator of the TRPV1 receptor and causes the opening of its cation channel allowing  $Ca^{2+}$  to enter [93], inhibited the CBD-mediated  $Ca^{2+}$  response [97, 98]. This led to the hypothesis that in diseases where an increase of AEA levels is observed, CBD may be less effective. However, this is the subject of ongoing investigation as CBD is neuroprotective in models of PD [80], a disease in which AEA levels are enhanced [85, 86].





**Figure 3.** Endocannabinoid system of the postsynaptic terminal. N-acyltransferase (NAT) transfers arachidonic acid (AA) onto NArPE in a calcium dependent manner which is then cleaved by NAPE-specific phospholipase D (NAPE-PLD) into AEA and phosphatidic acid (PA). Fatty acid amide hydrolase (FAAH) hydrolyses AEA into AA and EtNH<sub>2</sub>. The existence of an AEA membrane transporter (AMT) is still under debate but it may aid the movement of AEA across the plasma membrane. Grey lines - basic metabolism of AEA as described above, red lines – inhibitory actions of CBD on AEA hydrolysis and transport as well as inhibitory action of TRPV1 activation on mitochondrial calcium release, blue lines - release of calcium from mitochondria and TRPV1.

Evidence now suggests that Ca<sup>2+</sup> may be released by CBD from mitochondrial stores [99]. Presumably as a response to CBD, AEA levels are increased either due to CBD's inhibitory function on AEA hydrolysis or due to Ca<sup>2+</sup> influx resulting in a negative feedback cycle. Interestingly, the application of specific fatty-acid amide hydrolase inhibitors (FAAHI) (which inhibit hydrolysis of AEA and to some extent 2AG) does not cause a reduction in CBD response which may be due to the fact that CBD already acts as an inhibitor of hydrolysis [97] (summarised in Figure 3).

Therefore by increasing AEA levels within the cell, CBD may indirectly increase CB1 receptor activation, thus providing a potential mechanism by which CBD exerts its neuroprotective effects.

However, a more specific protective effect of CBD against mitochondrial toxins has also been proposed. A detailed study was published in 2007 evaluating the effect of CBD on 3-nitropropionic acid (3NP)-induced GABAergic neurone depletion [100]. 3NP is a mitochondrial toxin, inhibiting mitochondrial complex 2. It is used to model Huntington's disease (HD) as loss of function of the mitochondrial complex is its primary genetic cause [101]. In Sagredo's study, CBD completely reversed toxin-induced GABA depletion, whereas the specific CB1 receptor agonist, ACEA, and the specific CB2 agonist, HU-308, were not protective. Furthermore, coapplication of specific TRPV1 antagonists had no effect on the actions of CBD. Thus it is unlikely that the protective effect of CBD was exerted via the TRPV1 receptor despite its ability to agonise it [100]. This supports the findings of Ryan *et al* that CBD does not act via the TRPV1 receptor [102]. They have demonstrated that CBD prevents the membrane depolarisation caused by mitochondrial toxins. Furthermore, protection afforded by CBD against the mitochondrial toxin FCCP was potentiated by coapplication of the antioxidant BHT, indicating that the protective effect of CBD was not primarily exerted via its antioxidant properties [102].

In summary, CBD may not exert neuroprotective effects mediated by TRPV1 or antioxidant mechanisms but may uniquely protect against mitochondria-depolarising toxins through its calcium signalling abilities or through manipulating endocannabinoid tone. By contrast, unlike CBD where alternative modes of action were pursued due to its inability to bind CB1 or CB2 receptor, very little is known about the action of  $\Delta^9$ -THC on intracellular calcium and endocannabinoids. Nevertheless it has been shown that  $\Delta^9$ -THC modulates intracellular calcium levels in smooth muscle cells [103] and in the brain [104].

#### 4. Inflammation

Inflammation plays an important part in many neurodegenerative diseases. Both  $\Delta^9$ -THC and CBD have anti-inflammatory actions and might therefore be useful therapeutic agents in these conditions.

The anti-inflammatory action of  $\Delta^9$ -THC is thought to be mediated through the CB2 receptor which is mainly located on cells of the immune system [105], with the highest levels being found in B lymphocytes and natural killer cells. The main CB2 expressing cells of the CNS are microglia, which are related to macrophages. Like macrophages, they are attracted to sites of injury where they are activated and proliferate [106, 107]. Once activated, microglia start to display a more phagocytic phenotype with amoeboid morphology. At this stage they produce various inflammatory cytokines including IL-1, IL-6 and TNF- $\alpha$  and express major histocompatibility complex class 1 and 2 as well as the complement receptors CD11/CD18. The recruitment of microglia plays a major role in many neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease and multiple sclerosis.

The expression pattern of CB2 by microglia is dependent on the activation status of the cells: inactivated, resting microglia express low levels of CB2 which is markedly increased in intermediate activation states where cells are responsive and primed, and reduced in fully activated glial cells [108]. Cannabinoids are known to suppress some of the immune functions of microglia such as phagocytosis, cytolysis and cytokine secretion and may therefore potentially

reduce potentiation of neurodegeneration attributable to microglial activation [109-111].

The cannabinoids  $\Delta^9$ -THC and AEA are able to inhibit the secretion of the inflammatory cytokines TNF- $\alpha$ , IL- $\alpha$ , IL- $\beta$  and IL-6 by activated microglia [112, 113]. It is thought that cannabinoid-induced reduction in microglial chemotaxis and motility is primarily mediated by CB2 receptor activation [114-117].  $\Delta^9$ -THC has also been shown to reduce the production of inflammatory cytokines by peripheral immune cells in a mouse model of autoimmune hepatitis; this response was CB2 receptor dependent as shown by the co-application of specific antagonists [118]. Blocking FAAH activity also had an immune-suppressive effect [118]. Thus it is thought that anti-inflammatory actions of cannabinoids are mainly mediated via the CB2 receptor which actively prevents the activation of glial cells [109]. However, even though specific CB2 receptor agonists such as BML-190 and JWH-015 as well as  $\Delta^9$ -THC and AEA are able to protect neuronal cells from co-cultured, stimulated microglia, BML-190 caused a distinctly different cytokine profile compared with the other compounds indicating that regulation of inflammatory cytokine production may not directly influence neuronal cell viability [113].

Anti-inflammatory responses may also be partly mediated via the CB1 receptor, as indicated by a study using CB1 specific agonists which showed a dose dependent inhibition of nitric oxide production (a marker of microglial activation) [119]. Others have reported that CB1 agonists are able to reverse the morphological change of macrophages to a mobile amoeboid phenotype in a CB1 receptor-dependent manner [120, 121]. However, unlike in the rat, human microglia do not appear to express CB1 receptors which may limit the relevance of these findings to human disease [113].

Another potential mediator of  $\Delta^9$ -THC-associated anti-inflammatory effects is the peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ), a receptor involved in lipid homeostasis, which is widely expressed in neuronal and glial cells [122]. PPAR- $\gamma$  is a nuclear receptor that through binding to DNA promoter regions can regulate microglial activation [123], expression of inflammatory cytokines [124], nitric oxide synthase [125] and antioxidative enzymes [126]. In a mouse model of PD, the PPAR- $\gamma$  receptor agonist, pioglitazone, prevented dopaminergic cell loss and attenuated glial activation [127]. Both  $\Delta^9$ -THC and anandamide activate PPAR- $\gamma$  [128, 129]. Although AEA itself is a PPAR- $\gamma$  receptor agonist, evidence suggests that it inhibits IL-2 secretion via its COX-2 metabolites, an effect that can be attenuated by co-application of PPAR- $\gamma$  antagonists [124, 130]. Thus it is unclear whether PPAR- $\gamma$  activation alone is sufficient to mediate some of its anti-inflammatory effects.

CBD is a poor agonist at the CB2 receptor and thus mediates its anti-inflammatory actions through other, cannabinoid receptor-independent, mechanisms or through modulation of endocannabinoid levels as previously described. Carrier *et al* have proposed that CBD may act through modifying adenosine signalling [131, 132]. They showed that CBD inhibited the uptake of adenosine by nucleoside transporters in murine microglia, thereby increasing the activation of adenosine A2A receptors. A2A knock-out mice and antagonism of the A2A receptor lead to attenuation of the CBD-mediated effect, suggesting its importance in CBD-mediated anti-inflammatory actions [131, 132]. Further downstream, CBD prevents MAPK activation and

translocation of the nuclear factor NF- $\kappa$ B into the nucleus thus preventing the expression of inflammatory cytokines and nitric oxide synthase [133].

## 5. Pharmacokinetics

There are currently three cannabis-based medications clinically available, two of which contain  $\Delta^9$ -THC, whose licensed indications vary between countries. These include dronabinol (trade name Marinol®), formulated as capsules containing synthetic  $\Delta^9$ -THC; and Nabiximols oral spray (trade name Sativex®), a mixture of both CBD and  $\Delta^9$ -THC. The third medication is Nabilone, a synthetic  $\Delta^9$ -THC analogue. The potential therapeutic benefit of any drug is dependent on its pharmacokinetics which determines the onset, magnitude and duration of any effect depending on drug absorption by tissues, hepatic metabolism and excretion in bodily fluids and faeces. Thus, the route of administration makes a considerable difference to the bioavailability of  $\Delta^9$ -THC such that smoked cannabis leads to a quicker and higher peak of drug plasma levels compared with oral intake. Compared with intravenous administration of  $\Delta^9$ -THC, bioavailability by the oral route ranges between 4% and 20% [134-136], probably as a result of first pass metabolism by the liver into its over 80 different metabolites, the most abundant of which are the psychoactive metabolite 11-OH-THC and its inactive carboxylated form THC-COOH [135]. When smoked, mean plasma concentration of  $\Delta^9$ -THC peaks after approximately 6 minutes, followed by a rapid decrease to less than a quarter of its maximum concentration [134]. This effect, accompanied by a steady increase in 11-OH-THC, led researchers to believe that it was its metabolite rather than  $\Delta^9$ -THC itself that causes psychotic symptoms after consumption of the drug [137]. However, it is now known that, due its lipophilic properties, the majority of  $\Delta^9$ -THC is quickly absorbed by adipose tissue and then slowly released back into the bloodstream [138].

The extent to which CBD exerts clinically relevant positive effects when co-administered with  $\Delta^9$ -THC is the subject of ongoing debate, extensively reviewed by Russo *et al* [12]; the attenuating effect of CBD on the psychoactive effects of  $\Delta^9$ -THC is discussed in section 2. Nabiximols (Sativex®) contains  $\Delta^9$ -THC and CBD in almost equal proportions (2.7mg  $\Delta^9$ -THC to 2.5mg CBD ratio per spray) and is composed of the standardized  $\Delta^9$ -THC and CBD extracts Tetranabinex® and Nabidiolex® respectively. However, this does not mean that plasma levels of the drugs are equal following dose administration as both may be metabolised at different rates. Indeed, lower plasma concentrations of CBD have been found in comparison with  $\Delta^9$ -THC after administration of equal amounts of both compounds despite the fact that CBD is metabolised at a slower rate and, unlike  $\Delta^9$ -THC, the majority of CBD is excreted unchanged [134]. However, there is evidence that CBD may inhibit the conversion of  $\Delta^9$ -THC into its metabolite 11-OH-THC by the enzyme CYP450 [46, 134, 139, 140]. Accordingly it was found that metabolism of  $\Delta^9$ -THC to 11-OH-THC was reduced when CBD and  $\Delta^9$ -THC extracts were co-administered. Furthermore a 1:1 mixture delayed  $\Delta^9$ -THC absorption compared with  $\Delta^9$ -THC high ratios [141, 142].

## 6. Conclusion

Here we have discussed the diverse mechanisms of action of  $\Delta^9$ -THC and CBD and how these relate to their potential therapeutic role in neurological disorders. Although both compounds exhibit neuroprotective and anti-inflammatory effects their receptor pharmacology distinctly differs, indicating divergent mechanisms of action. These are probably mediated by the CB1 and CB2 receptors in the case of  $\Delta^9$ -THC, but are much less clear in relation to CBD as the compound displays negligible affinity for both cannabinoid receptors. Thus CBD research has focused on non-receptor mediated mechanisms of actions such as  $\text{Ca}^{2+}$  and endocannabinoid tone modulation whereas the effect of  $\Delta^9$ -THC on these two variables is uncertain. The clinical utility of  $\Delta^9$ -THC may be limited by its psychoactive potential, whereas CBD not only appears to have antipsychotic effects but may also counteract the psychoactive properties of  $\Delta^9$ -THC. Thus it may be of interest to study the pharmacological properties of these two compounds not only separately but also in conjunction to further evaluate their clinical potential.

## Abbreviations

2AG - 2-acylglycerol  
 3NP - 3-nitropropionic acid  
 A2A - Adenosine A2A receptor  
 AEA - Anandamide  
 BHT - Butylated hydroxytoluene  
 CBD - Cannabidiol  
 CNS - Central nervous system  
 DSE - Depolarisation induced suppression of excitation  
 DSI - Depolarisation induced suppression of inhibition  
 FAAH - Fatty acid amide hydrolase  
 FCCP - carbonylcyanide-*p*-trifluoromethoxyphenylhydrazone  
 fMRI - Functional magnetic resonance imaging  
 HD - Huntington's Disease  
 LTD - Long term depression  
 LTP - Long term potentiation  
 mGlu1 - Metabotropic glutamate receptor 1  
 PD - Parkinson's disease  
 TRPV1 - transient receptor potential cation channel, subfamily V, member 1  
 $\Delta^9$ -THC -  $\Delta^9$ -Tetrahydrocannabinol

## References

1. ElSohly, M.A. and D. Slade, Chemical constituents of marijuana: The complex mixture of natural cannabinoids. *Life Sciences*, 2005. 78(5): p. 539-548.
2. Matsuda, L.A., *et al.*, Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature*, 1990. 346(6284): p. 561-564.
3. Munro, S., K.L. Thomas, and M. Abushaar, Molecular characterization of a peripheral receptor for cannabinoids.

Nature, 1993. 365(6441): p. 61-65.

4. Zajicek, J., *et al.*, Cannabinoids for treatment of spasticity and other symptoms related to multiple sclerosis (CAMS study): multicentre randomised placebo-controlled trial. *Lancet*, 2003. 362(9395): p. 1517-1526.

5. Wade, D.T., *et al.*, A preliminary controlled study to determine whether whole-plant cannabis extracts can improve intractable neurogenic symptoms. *Clinical Rehabilitation*, 2003. 17(1): p. 21-29.

6. Carroll, C.B., *et al.*, Cannabis for dyskinesia in Parkinson disease - A randomized double-blind crossover study. *Neurology*, 2004. 63(7): p. 1245-1250.

7. Fox, P., *et al.*, The effect of cannabis on tremor in patients with multiple sclerosis. *Neurology*, 2004. 62(7): p. 1105-1109.

8. Cohen, M., N. Solowij, and V. Carr, Cannabis, cannabinoids and schizophrenia: integration of the evidence. *Australian and New Zealand Journal of Psychiatry*, 2008. 42(5): p. 357-368.

9. Zuardi, A.W., *et al.*, Cannabidiol, a *Cannabis sativa* constituent, as an antipsychotic drug. *Brazilian Journal of Medical and Biological Research*, 2006. 39(4): p. 421-429.

10. Zuardi, A.W., *et al.*, Cannabidiol for the treatment of psychosis in Parkinson's disease. *Journal of Psychopharmacology*, 2009. 23(8): p. 979-983.

11. Croxford, J.L., Therapeutic potential of cannabinoids in CNS disease. *Cns Drugs*, 2003. 17(3): p. 179-202.

12. Russo, E. and G.W. Guy, A tale of two cannabinoids: The therapeutic rationale for combining tetrahydrocannabinol and cannabidiol. *Medical Hypotheses*, 2006. 66(2): p. 234-246.

13. Ryberg, E., *et al.*, The orphan receptor GPR55 is a novel cannabinoid receptor. *British Journal of Pharmacology*, 2007. 152(7): p. 1092-1101.

14. Lichtman, A.H., K.R. Dimen, and B.R. Martin, Systemic or intrahippocampal cannabinoid administration impairs spatial memory in rats. *Psychopharmacology*, 1995. 119(3): p. 282-290.

15. Bolla, K.I., *et al.*, Dose-related neurocognitive effects of marijuana use. *Neurology*, 2002. 59(9): p. 1337-1343.

16. Pope, H.G., *et al.*, Cognitive measures in long-term cannabis users. *Journal of Clinical Pharmacology*, 2002. 42(11): p. 41S-47S.

17. D'Souza, D.C., *et al.*, Delta-9-tetrahydrocannabinol effects in schizophrenia: Implications for cognition, psychosis, and addiction. *Biological Psychiatry*, 2005. 57(6): p. 594-608.

18. D'Souza, D.C., R.A. Sewell, and M. Ranganathan, Cannabis and psychosis/schizophrenia: human studies. *European Archives of Psychiatry and Clinical Neuroscience*, 2009. 259(7): p. 413-431.

19. Senn, R., *et al.*, Long-term cognitive deficits induced by a single, extremely low dose of tetrahydrocannabinol (THC): Behavioral, pharmacological and biochemical studies in mice. *Pharmacology Biochemistry and Behavior*, 2008. 88(3): p. 230-237.

20. Amal, H., *et al.*, Long-term consequences of a single treatment of mice with an ultra-low dose of Delta(9)-

tetrahydrocannabinol (THC). *Behavioural Brain Research*, 2010. 206(2): p. 245-253.

21. Brodtkin, J. and J.M. Moerschbaecher, SR141716A antagonizes the disruptive effects of cannabinoid ligands on learning in rats. *Journal of Pharmacology and Experimental Therapeutics*, 1997. 282(3): p. 1526-1532.

22. Lichtman, A.H. and B.R. Martin, Delta(9)-tetrahydrocannabinol impairs spatial memory through a cannabinoid receptor mechanism. *Psychopharmacology*, 1996. 126(2): p. 125-131.

23. Mallet, P.E. and R.J. Beninger, The cannabinoid CB1 receptor antagonist SR141716A attenuates the memory impairment produced by Delta(9)-tetrahydrocannabinol or anandamide. *Psychopharmacology*, 1998. 140(1): p. 11-19.

24. Herkenham, M., *et al.*, Cannabinoid receptor localization in brain. *Proceedings of the National Academy of Sciences of the United States of America*, 1990. 87(5): p. 1932-1936.

25. Mato, S., *et al.*, A single in-vivo exposure to Delta 9THC blocks endocannabinoid-mediated synaptic plasticity. *Nature Neuroscience*, 2004. 7(6): p. 585-586.

26. Hoffman, A.F., *et al.*, Opposing actions of chronic Delta 9-tetrahydrocannabinol and cannabinoid antagonists on hippocampal long-term potentiation. *Learning & Memory*, 2007. 14(1-2): p. 63-74.

27. Shen, M.X. and S.A. Thayer, The cannabinoid agonist Win55,212-2 inhibits calcium channels by receptor-mediated and direct pathways in cultured rat hippocampal neurons. *Brain Research*, 1998. 783(1): p. 77-84.

28. Twitchell, W., S. Brown, and K. Mackie, Cannabinoids inhibit N- and P/Q-type calcium channels in cultured rat hippocampal neurons. *Journal of Neurophysiology*, 1997. 78(1): p. 43-50.

29. Mackie, K. and B. Hille, Cannabinoids inhibit N-type calcium channels in neuroblastoma glioma-cells. *Proceedings of the National Academy of Sciences of the United States of America*, 1992. 89(9): p. 3825-3829.

30. Deadwyler, S.A., *et al.*, CANNABINOIDS MODULATE POTASSIUM CURRENT IN CULTURED HIPPOCAMPAL-NEURONS. *Receptors & Channels*, 1993. 1(2): p. 121-134.

31. Mackie, K., *et al.*, Cannabinoids activate an inwardly rectifying potassium conductance and inhibit Q-type calcium currents in ATT20 cells transfected with rat-brain cannabinoid receptor. *Journal of Neuroscience*, 1995. 15(10): p. 6552-6561.

32. Wilson, R.I. and R.A. Nicoll, Endogenous cannabinoids mediate retrograde signalling at hippocampal synapses. *Nature*, 2001. 410(6828): p. 588-592.

33. Fan, N., *et al.*, Reduced expression of glutamate receptors and phosphorylation of CREB are responsible for *in vivo* Delta 9-THC exposure-impaired hippocampal synaptic plasticity. *Journal of Neurochemistry*, 2010. 112(3): p. 691-702.

34. Wilson, R.I. and R.A. Nicoll, Neuroscience - Endocannabinoid signaling in the brain. *Science*, 2002. 296(5568): p. 678-682.

35. Hashimoto-dani, Y., T. Ohno-Shosaku, and M. Kano, Endocannabinoids and synaptic function in the CNS. *Neuroscientist*, 2007. 13(2): p. 127-137.

36. Breivogel, C.S., *et al.*, Chronic Delta(9)-tetrahydrocannabinol treatment produces a time-dependent loss of cannabinoid receptors and cannabinoid receptor-activated G proteins in rat brain. *Journal of Neurochemistry*, 1999. 73(6): p. 2447-2459.
37. Sim-Selley, L.J. and B.R. Martin, Effect of chronic administration of R-(+)-[2,3-dihydro-5-methyl-3-[(morpholinyl)methyl]pyrrolo[1,2,3-de]-1,4 -benzoxazinyl]-(1-naphthalenyl)methanone mesylate (IN55,212-2) or Delta(9)-tetrahydrocannabinol on cannabinoid receptor adaptation in mice. *Journal of Pharmacology and Experimental Therapeutics*, 2002. 303(1): p. 36-44.
38. Rubino, T., *et al.*, Modulation of extracellular signal-regulated kinases cascade by chronic Delta(9)-tetrahydrocannabinol treatment. *Molecular and Cellular Neuroscience*, 2004. 25(3): p. 355-362.
39. Casu, M.A., *et al.*, Effect of Delta(9)-tetrahydrocannabinol on phosphorylated CREB in rat cerebellum: An immunohistochemical study. *Brain Research*, 2005. 1048(1-2): p. 41-47.
40. Derkinderen, P., *et al.*, Regulation of extracellular signal-regulated kinase by cannabinoids in hippocampus. *Journal of Neuroscience*, 2003. 23(6): p. 2371-2382.
41. Valjent, E., *et al.*, Delta 9-tetrahydrocannabinol-induced MAPK/ERK and Elk-1 activation *in vivo* depends on dopaminergic transmission. *European Journal of Neuroscience*, 2001. 14(2): p. 342-352.
42. Gainetdinov, R.R., *et al.*, Desensitization of G protein-coupled receptors and neuronal functions. *Annual Review of Neuroscience*, 2004. 27: p. 107-144.
43. Daigle, T.L., C.S. Kearn, and K. Mackie, Rapid CB1 cannabinoid receptor desensitization defines the time course of ERK1/2 MAP kinase signaling. *Neuropharmacology*, 2008. 54(1): p. 36-44.
44. Turu, G. and L. Hunyady, Signal transduction of the CB1 cannabinoid receptor. *Journal of Molecular Endocrinology*. 44(2): p. 75-85.
45. Hayakawa, K., *et al.*, Cannabidiol potentiates pharmacological effects of Delta(9)-tetrahydrocannabinol via CB1 receptor-dependent mechanism. *Brain Research*, 2008. 1188: p. 157-164.
46. Bornheim, L.M., *et al.*, Effect of cannabidiol pretreatment on the kinetics of tetrahydrocannabinol metabolites in mouse brain. *Drug Metabolism and Disposition*, 1995. 23(8): p. 825-831.
47. Karniol, I.G., *et al.*, Cannabidiol interferes with effects of delta-9-tetrahydrocannabinol in man. *European Journal of Pharmacology*, 1974. 28(1): p. 172-177.
48. Dalton, W.S., *et al.*, Influence of cannabidiol on delta-9-tetrahydrocannabinol effects. *Clinical Pharmacology & Therapeutics*, 1976. 19(3): p. 300-309.
49. Zuardi, A.W., *et al.*, Action of cannabidiol on the anxiety and other effects produced by delta-9-THC in normal subjects. *Psychopharmacology*, 1982. 76(3): p. 245-250.
50. Leweke, F.M., *et al.*, Cannabidiol as an antipsychotic agent. *European Psychiatry*, 2007. 22: p. S21-S21.
51. Fadda, P., *et al.*, Differential effects of THC or CBD-rich cannabis extracts on working memory in rats. *Neuropharmacology*, 2004. 47(8): p. 1170-1179.



52. Zuardi, A.W. and I.G. Karniol, Effects on variable-interval performance in rats of delta-9-tetrahydrocannabinol and cannabidiol, separately and in combination. *Brazilian Journal of Medical and Biological Research*, 1983. 16(2): p. 141-146.
53. Bhattacharyya, S., *et al.*, Opposite effects of delta-9-tetrahydrocannabinol and cannabidiol on human brain function and psychopathology. *Neuropsychopharmacology*, 2009. 35(3): p. 764-74.
54. Zuardi, A.W., *et al.*, Cannabidiol monotherapy for treatment-resistant schizophrenia. *Journal of Psychopharmacology*, 2006. 20(5): p. 683-686.
55. Guimaraes, V.M.C., *et al.*, Cannabidiol increases Fos expression in the nucleus accumbens but not in the dorsal striatum. *Life Sciences*, 2004. 75(5): p. 633-638.
56. Thomas, A., *et al.*, Cannabidiol displays unexpectedly high potency as an antagonist of CB1 and CB2 receptor agonists *in vitro*. *British Journal of Pharmacology*, 2007. 150(5): p. 613-623.
57. Petitet, F., *et al.*, Complex pharmacology of natural cannabinoids: Evidence for partial agonist activity of Delta (9)-tetrahydrocannabinol and antagonist activity of cannabidiol on rat brain cannabinoid receptors. *Life Sciences*, 1998. 63(1): p. PL1-PL6.
58. Jones, N.A., *et al.*, Cannabidiol Displays Antiepileptiform and Antiseizure Properties In Vitro and In Vivo. *Journal of Pharmacology and Experimental Therapeutics*. 332(2): p. 569-577.
59. Pertwee, R.G., The diverse CB1 and CB2 receptor pharmacology of three plant cannabinoids: Delta(9)-tetrahydrocannabinol, cannabidiol and Delta(9)-tetrahydrocannabivarin. *British Journal of Pharmacology*, 2008. 153(2): p. 199-215.
60. Gilbert, G.L., *et al.*, Delta(9)-Tetrahydrocannabinol protects hippocampal neurons from excitotoxicity. *Brain Research*, 2007. 1128(1): p. 61-69.
61. Kim, H.J., J.J. Waataja, and S.A. Thayer, Cannabinoids inhibit network-driven synapse loss between hippocampal neurons in culture. *Journal of Pharmacology and Experimental Therapeutics*, 2008. 325(3): p. 850-858.
62. Panikashvili, D., *et al.*, An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature*, 2001. 413(6855): p. 527-531.
63. Pryce, G., *et al.*, Cannabinoids inhibit neurodegeneration in models of multiple sclerosis. *Brain*, 2003. 126: p. 2191-2202.
64. Gilbert, G.L., *et al.*, [Delta]9-Tetrahydrocannabinol protects hippocampal neurons from excitotoxicity. *Brain Research*, 2007. 1128: p. 61-69.
65. Abood, M.E., *et al.*, Activation of the CB1 cannabinoid receptor protects cultured mouse spinal neurons against excitotoxicity. *Neuroscience Letters*, 2001. 309(3): p. 197-201.
66. Nagayama, T., *et al.*, Cannabinoids and neuroprotection in global and focal cerebral ischemia and in neuronal cultures. *Journal of Neuroscience*, 1999. 19(8): p. 2987-2995.
67. Shen, M.X. and S.A. Thayer, Cannabinoid receptor agonists protect cultured rat hippocampal neurons from excitotoxicity. *Molecular Pharmacology*, 1998. 54(3): p. 459-462.

68. van der Stelt, M., *et al.*, Neuroprotection by Delta(9)-tetrahydrocannabinol, the main active compound in marijuana, against ouabain-induced *in vivo* excitotoxicity. *Journal of Neuroscience*, 2001. 21(17): p. 6475-6479.
69. Marsicano, G., *et al.*, CB1 cannabinoid receptors and on-demand defense against excitotoxicity. *Science*, 2003. 302(5642): p. 84-88.
70. Herkenham, M., *et al.*, Characterization and localization of cannabinoid receptors in rat brain - A quantitative *in vitro* autoradiographic study. *Journal of Neuroscience*, 1991. 11(2): p. 563-583.
71. Hohmann, A.G. and M. Herkenham, Localization of cannabinoid CB1 receptor mRNA in neuronal subpopulations of rat striatum: A double-label *in situ* hybridization study. *Synapse*, 2000. 37(1): p. 71-80.
72. Domenici, M.R., *et al.*, Cannabinoid receptor type 1 located on presynaptic terminals of principal neurons in the forebrain controls glutamatergic synaptic transmission. *Journal of Neuroscience*, 2006. 26(21): p. 5794-5799.
73. Romero, J., *et al.*, Unilateral 6-hydroxydopamine lesions of nigrostriatal dopaminergic neurons increased CB1 receptor mRNA levels in the caudate-putamen. *Life Sciences*, 2000. 66(6): p. 485-494.
74. Garcia-Arencibia, M., *et al.*, Cannabinoid CB1 Receptors are Early DownRegulated Followed by a Further UpRegulation in the Basal Ganglia of Mice with Deletion of Specific Park Genes. *Journal of Neural Transmission-Supplement*, 2009(73): p. 269-275.
75. Glass, M., R.L.M. Faull, and M. Dragunow, Loss of cannabinoid receptors in the substantia-nigra in Huntington's disease. *Neuroscience*, 1993. 56(3): p. 523-527.
76. Olney, J.W., T. Fuller, and T. Degubareff, Acute dendrotoxic changes in the hippocampus of kainate treated rats. *Brain Research*, 1979. 176(1): p. 91-100.
77. Fiala, J.C., J. Spacek, and K.M. Harris, Dendritic spine pathology: Cause or consequence of neurological disorders? *Brain Research Reviews*, 2002. 39(1): p. 29-54.
78. Waataja, J.J., *et al.*, Excitotoxic loss of post-synaptic sites is distinct temporally and mechanistically from neuronal death. *Journal of Neurochemistry*, 2008. 104(2): p. 364-375.
79. Doble, A., The role of excitotoxicity in neurodegenerative disease: Implications for therapy. *Pharmacology & Therapeutics*, 1999. 81(3): p. 163-221.
80. Garcia-Arencibia, M., *et al.*, Evaluation of the neuroprotective effect of cannabinoids in a rat model of Parkinson's disease: Importance of antioxidant and cannabinoid receptor-independent properties. *Brain Research*, 2007. 1134(1): p. 162-170.
81. Hampson, A.J., *et al.*, Cannabidiol and (-)-Delta(9)-tetrahydrocannabinol are neuroprotective antioxidants. *Proceedings of the National Academy of Sciences of the United States of America*, 1998. 95(14): p. 8268-8273.
82. Price, D.A., *et al.*, WIN55,212-2, a cannabinoid receptor agonist, protects against nigrostriatal cell loss in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *European Journal of Neuroscience*, 2009. 29(11): p. 2177-2186.
83. Jimenez-Del-Rio, M., A. Daza-Restrepo, and C. Velez-Pardo, The cannabinoid CP55,940 prolongs survival and improves locomotor activity in *Drosophila melanogaster* against paraquat: Implications in Parkinson's disease. *Neuroscience Research*, 2008. 61(4): p. 404-411.

84. Marsicano, G., *et al.*, Neuroprotective properties of cannabinoids against oxidative stress: role of the cannabinoid receptor CB1. *Journal of Neurochemistry*, 2002. 80(3): p. 448-456.
85. Pisani, A., *et al.*, High endogenous cannabinoid levels in the cerebrospinal fluid of untreated Parkinson's disease patients. *Annals of Neurology*, 2005. 57(5): p. 777-779.
86. Maccarrone, M., *et al.*, Levodopa treatment reverses endocannabinoid system abnormalities in experimental parkinsonism. *Journal of Neurochemistry*, 2003. 85(4): p. 1018-1025.
87. Lastres-Becker, I., *et al.*, Changes in endocannabinoid transmission in the basal ganglia in a rat model of Huntington's disease. *Neuroreport*, 2001. 12(10): p. 2125-2129.
88. Lastres-Becker, I., *et al.*, Compounds acting at the endocannabinoid and/or endovanilloid systems reduce hyperkinesia in a rat model of Huntington's disease. *Journal of Neurochemistry*, 2003. 84(5): p. 1097-1109.
89. Fernandez-Ruiz, J., The endocannabinoid system as a target for the treatment of motor dysfunction. *British Journal of Pharmacology*, 2009. 156(7): p. 1029-1040.
90. Glass, M., M. Dragunow, and R.L.M. Faull, The pattern of neurodegeneration in Huntington's disease: A comparative study of cannabinoid, dopamine, adenosine and GABA(A) receptor alterations in the human basal ganglia in Huntington's disease. *Neuroscience*, 2000. 97(3): p. 505-519.
91. Lastres-Becker, I., *et al.*, Alleviation of motor hyperactivity and neurochemical deficits by endocannabinoid uptake inhibition in a rat model of Huntington's disease. *Synapse*, 2002. 44(1): p. 23-35.
92. Bisogno, T., *et al.*, Molecular targets for cannabidiol and its synthetic analogues: effect on vanilloid VR1 receptors and on the cellular uptake and enzymatic hydrolysis of anandamide. *British Journal of Pharmacology*, 2001. 134(4): p. 845-852.
93. van der Stelt M., *et al.*, Anandamide acts as an intracellular messenger amplifying Ca<sup>2+</sup> influx via TRPV1 channels. *The EMBO Journal*, 2005. 24: p. 3026-3037.
94. Evans, D.M., M.R. Johnson, and A.C. Howlett, Ca<sup>2+</sup>-dependent release from rat-brain of cannabinoid receptor-binding activity. *Journal of Neurochemistry*, 1992. 58(2): p. 780-782.
95. Di Marzo, V., *et al.*, Formation and inactivation of endogenous cannabinoid anandamide in central neurons. *Nature*, 1994. 372(6507): p. 686-691.
96. Drysdale, A.J., *et al.*, Cannabidiol-induced intracellular Ca<sup>2+</sup> elevations in hippocampal cells. *Neuropharmacology*, 2006. 50(5): p. 621-631.
97. Ryan, D., *et al.*, Interactions of cannabidiol with endocannabinoid signalling in hippocampal tissue. *European Journal of Neuroscience*, 2007. 25(7): p. 2093-2102.
98. Watanabe, K., *et al.*, Inhibition of anandamide amidase activity in mouse brain microsomes by cannabinoids. *Biological & Pharmaceutical Bulletin*, 1996. 19(8): p. 1109-1111.
99. Ryan, D., *et al.*, Cannabidiol Targets Mitochondria to Regulate Intracellular Ca<sup>2+</sup> Levels. *Journal of Neuroscience*, 2009. 29(7): p. 2053-2063.

100. Sagredo O., *et al.*, Cannabidiol reduced the striatal atrophy caused 3-nitropropionic acid *in vivo* by mechanisms independent of the activation of cannabinoid, vanilloid TRPV1 and adenosine A2A receptors. *European Journal of Neuroscience*, 2007. 26: p. 843-851.
101. Tunez, I., *et al.*, 3-Nitropropionic Acid as a Tool to Study the Mechanisms Involved in Huntington's Disease: Past, Present and Future. *Molecules*. 15(2): p. 878-916.
102. Ryan D., *et al.*, Cannabidiol targets mitochondria to regulate intracellular Ca<sup>2+</sup> Levels. *The Journal of Neuroscience*, 2009. 29(7): p. 2053-2063.
103. Filipeanu, C.M., D. deZeeuw, and S.A. Nelemans, Delta(9)-tetrahydrocannabinol activates [Ca<sup>2+</sup>], increases partly sensitive to capacitative store refilling. *European Journal of Pharmacology*, 1997. 336(1): p. R1-R3.
104. Okada, M., *et al.*, The facilitating and suppressing effects of delta-9-tetrahydrocannabinol on the rise in intrasynaptosomal Ca<sup>2+</sup> concentration in rats. *Neuroscience Letters*, 1992. 140(1): p. 55-58.
105. Galiegue, S., *et al.*, Expression of central and peripheral cannabinoid receptors in human immune tissues and leukocyte subpopulations. *European Journal of Biochemistry*, 1995. 232(1): p. 54-61.
106. Leong, S.K. and E.A. Ling, Ameboid and ramified microglia-their interrelationship and response to brain injury. *Glia*, 1992. 6(1): p. 39-47.
107. Heppner, F.L., *et al.*, Activated microglial cells migrate towards sites of excitotoxic neuronal injury inside organotypic hippocampal slice cultures. *European Journal of Neuroscience*, 1998. 10(10): p. 3284-3290.
108. Carlisle, S.J., *et al.*, Differential expression of the CB2 cannabinoid receptor by rodent macrophages and macrophage-like cells in relation to cell activation. *International Immunopharmacology*, 2002. 2(1): p. 69-82.
109. Ramirez, B.G., *et al.*, Prevention of Alzheimer's disease pathology by cannabinoids: Neuroprotection mediated by blockade of microglial activation. *Journal of Neuroscience*, 2005. 25(8): p. 1904-1913.
110. Meda, L., *et al.*, Activation of microglial cells by beta-amyloid protein and interferon-gamma. *Nature*, 1995. 374 (6523): p. 647-650.
111. Xie, Z., *et al.*, Peroxynitrite mediates neurotoxicity of amyloid beta-peptide(1-42)- and lipopolysaccharide-activated microglia. *Journal of Neuroscience*, 2002. 22(9): p. 3484-3492.
112. Puffenbarger, R.A., A.C. Boothe, and G.A. Cabral, Cannabinoids inhibit LPS-inducible cytokine mRNA expression in rat microglial cells. *Glia*, 2000. 29(1): p. 58-69.
113. Klegeris, A., C.J. Bissonnette, and P.L. McGeer, Reduction of human monocytic cell neurotoxicity and cytokine secretion by ligands of the cannabinoid-type CB2 receptor. *British Journal of Pharmacology*, 2003. 139(4): p. 775-786.
114. Cabral, G.A., *et al.*, CB2 receptors in the brain: role in central immune function. *British Journal of Pharmacology*, 2008. 153(2): p. 240-251.
115. Sacerdote, P., *et al.*, The nonpsychoactive component of marijuana cannabidiol modulates chemotaxis and IL-10 and IL-12 production of murine macrophages both *in vivo* and *in vitro*. *Journal of Neuroimmunology*, 2005. 159 (1-2): p. 97-105.

116. Walter, L., *et al.*, Nonpsychotropic cannabinoid receptors regulate microglial cell migration. *Journal of Neuroscience*, 2003. 23(4): p. 1398-1405.
117. Franklin, A. and N. Stella, Arachidonylcyclopropylamide increases microglial cell migration through cannabinoid CB2 and abnormal-cannabidiol-sensitive receptors. *European Journal of Pharmacology*, 2003. 474(2-3): p. 195-198.
118. Hegde, V.L., *et al.*, Attenuation of experimental autoimmune hepatitis by exogenous and endogenous cannabinoids: Involvement of regulatory T cells. *Molecular Pharmacology*, 2008. 74(1): p. 20-33.
119. Waksman, Y., *et al.*, The central cannabinoid receptor (CB1) mediates inhibition of nitric oxide production by rat microglial cells. *Journal of Pharmacology and Experimental Therapeutics*, 1999. 288(3): p. 1357-1366.
120. Stefano, G.B., *et al.*, Macrophage behavior associated with acute and chronic exposure to HIV GP120, morphine and anandamide: endothelial implications. *International Journal of Cardiology*, 1998. 64: p. S3-S13.
121. Sacerdote, P., *et al.*, In vivo and *in vitro* treatment with the synthetic cannabinoid CP55,940 decreases the *in vitro* migration of macrophages in the rat: involvement of both CB1 and CB2 receptors. *Journal of Neuroimmunology*, 2000. 109(2): p. 155-163.
122. Braissant, O., *et al.*, Differential expression of peroxisome proliferator-activated receptors (PPARs): Tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology*, 1996. 137(1): p. 354-366.
123. Breidert, T., *et al.*, The PPAR gamma agonist pioglitazone inhibits neuronal cell loss and glial activation in the substantia nigra of MPTP treated mice. *Movement Disorders*, 2002. 17: p. P130.
124. Rockwell, C.E. and N.E. Kaminski, A cyclooxygenase metabolite of anandamide causes inhibition of interleukin-2 secretion in murine splenocytes. *Journal of Pharmacology and Experimental Therapeutics*, 2004. 311(2): p. 683-690.
125. Dehmer, T., *et al.*, Protection by pioglitazone in the MPTP model of Parkinson's disease correlates with I kappa B alpha induction and block of NF kappa B and iNOS activation. *Journal of Neurochemistry*, 2004. 88(2): p. 494-501.
126. Jung, T.W., *et al.*, Rosiglitazone protects human neuroblastoma SH-SY5Y cells against MPP+ induced cytotoxicity via inhibition of mitochondrial dysfunction and ROS production. *Journal of the Neurological Sciences*, 2007. 253(1-2): p. 53-60.
127. Breidert, T., *et al.*, Protective action of the peroxisome proliferator-activated receptor-gamma agonist pioglitazone in a mouse model of Parkinson's disease. *Journal of Neurochemistry*, 2002. 82(3): p. 615-624.
128. O'Sullivan, S.E., *et al.*, Novel time-dependent vascular actions of Delta(9)-tetrahydrocannabinol mediated by peroxisome proliferator-activated receptor gamma. *Biochemical and Biophysical Research Communications*, 2005. 337(3): p. 824-831.
129. Bouaboula, M., *et al.*, Anandamide induced PPAR gamma transcriptional activation and 3T3-L1 preadipocyte differentiation. *European Journal of Pharmacology*, 2005. 517(3): p. 174-181.
130. Rockwell, C.E., *et al.*, Interleukin-2 suppression by 2-arachidonyl glycerol is mediated through peroxisome proliferator-activated receptor gamma independently of cannabinoid receptors 1 and 2. *Molecular Pharmacology*,

2006. 70(1): p. 101-111.

131. Carrier, E.J., J.A. Auchampach, and C.J. Hillard, Inhibition of an equilibrative nucleoside transporter by cannabidiol: A mechanism of cannabinoid immunosuppression. *Proceedings of the National Academy of Sciences of the United States of America*, 2006. 103(20): p. 7895-7900.

132. Liou, G.I., *et al.*, Mediation of Cannabidiol Anti-inflammation in the Retina by Equilibrative Nucleoside Transporter and A(2A) Adenosine Receptor. *Investigative Ophthalmology & Visual Science*, 2008. 49(12): p. 5526-5531.

133. Esposito, G., *et al.*, Cannabidiol inhibits inducible nitric oxide synthase protein expression and nitric oxide production in beta-amyloid stimulated PC12 neurons through p38 MAP kinase and NF-kappa B involvement. *Neuroscience Letters*, 2006. 399(1-2): p. 91-95.

134. Huestis, M.A., Human cannabinoid pharmacokinetics. *Chemistry & Biodiversity*, 2007. 4(8): p. 1770-1804.

135. Agurell, S., *et al.*, Pharmacokinetics and metabolism of delta-1 tetrahydrocannabinol and other cannabinoids with emphasis on man. *Pharmacological Reviews*, 1986. 38(1): p. 21-43.

136. Ohlsson, A., *et al.*, Plasma delta-9-tetrahydrocannabinol concentrations and clinical effects after oral and intravenous administration and smoking. *Clinical Pharmacology & Therapeutics*, 1980. 28(3): p. 409-416.

137. Lemberge, L., *et al.*, Delta-9-tetrahydrocannabinol - Temporal correlation of psychologic effects and blood levels after various routes of administration. *New England Journal of Medicine*, 1972. 286(13): p. 685-&.

138. Kreuz, D.S. and J. Axelrod, Delta-9-tetrahydrocannabinol - Localization in body fat. *Science*, 1973. 179(4071): p. 391-392.

139. Bornheim, L.M. and M.P. Grillo, Characterization of cytochrome P450 3A inactivation by cannabidiol: Possible involvement of cannabidiol-hydroxyquinone as a P450 inactivator. *Chemical Research in Toxicology*, 1998. 11(10): p. 1209-1216.

140. McArdle, K., *et al.*, Selective inhibition of Delta(9)-tetrahydrocannabinol metabolite formation by cannabidiol *in vitro*. *Toxicology*, 2001. 168(1): p. 133-134.

141. Guy, G.W. and P.J. Robson, A phase I, double blind, three-way crossover study to assess the pharmacokinetic profile of cannabis based medicine extract (CBME) administered sublingually in variant cannabinoid ratios in normal healthy male volunteers (GWPK0215). *Journal of Cannabis Therapeutics*, 2004. 3(4): p. 121-152.

142. Guy, G.W. and P.J. Robson, A Phase I, open label, four-way crossover study to compare the pharmacokinetic profiles of a single dose of 20 mg of a cannabis based medicine extract (CBME) administered on 3 different areas of the buccal mucosa and to investigate the pharmacokinetics of CBME per oral in healthy male and female volunteers. *J. Cannabis Therap*, 2003. 3(4): p. 79-120.