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# Influence of dietary nitrate supplementation on local sweating and cutaneous vascular responses during exercise in a hot environment.

Amano, T

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## Influence of dietary nitrate supplementation on local sweating and cutaneous vascular responses during exercise in a hot environment

--Manuscript Draft--

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<b>Corresponding Author:</b>	Narihiko Kondo, Ph.D. Kobe University Kobe, Hyogo JAPAN	
<b>Corresponding Author Secondary Information:</b>		
<b>Corresponding Author's Institution:</b>	Kobe University	
<b>Corresponding Author's Secondary Institution:</b>		
<b>First Author:</b>	Tatsuro Amano	
<b>First Author Secondary Information:</b>		
<b>Order of Authors:</b>	Tatsuro Amano	
	Dai Okushima	
	Brynmor Breese	
	Stephen Bailey	
	Shunsaku Koga	
	Narihiko Kondo	
<b>Order of Authors Secondary Information:</b>		
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<b>Abstract:</b>	<p><b>Purpose:</b> We investigated the influence of inorganic nitrate (NO<sub>3</sub><sup>-</sup>) supplementation on local sweating and cutaneous vascular responses during exercise in hot conditions.</p> <p><b>Method:</b> Eight healthy, young subjects were assigned in a randomized, double-blind, crossover design to receive NO<sub>3</sub><sup>-</sup>-rich beetroot (BR) juice (140 mL/day, containing ~8 mmol of NO<sub>3</sub><sup>-</sup>) and NO<sub>3</sub><sup>-</sup>-depleted placebo (PL) juice (140 mL/day, containing ~0.003 mmol of NO<sub>3</sub><sup>-</sup>) for 3 days. On day 3 of supplementation, subjects cycled at an intensity corresponding to 55% of <math>\dot{V}O_{2max}</math> for 30 minutes in hot conditions (30C, 50% relative humidity). Chest and forearm sweat rate (SR) and skin blood flow (SkBF), were measured continuously. Cutaneous vascular conductance (CVC) was calculated by SkBF/mean arterial pressure (MAP). Results: Prior to exercise, plasma NO<sub>3</sub><sup>-</sup> (21 ± 6 and 581 ± 161 M) and nitrite (NO<sub>2</sub><sup>-</sup>, 87 ± 28 and 336 ± 156 nM) concentrations were higher after BR compared to PL supplementation (P ≤ 0.011, n=6). Oesophageal, mean skin, and mean body temperatures during exercise were not different between conditions. In addition, BR supplementation did not affect SR, SkBF, and CVC during exercise. A lower MAP was found after 30 minutes of exercise following BR supplementation (112 ± 6 and 103 ± 6 mmHg for PL and BR, respectively, P = 0.021).</p>	

	Conclusion: These results suggest that inorganic NO <sub>3</sub> - supplementation, which increases the potential for O <sub>2</sub> -independent NO production, does not affect local sweating and cutaneous vascular responses, but attenuates blood pressure in young healthy subjects exercising in a hot environment.
<b>Response to Reviewers:</b>	see attachment

## Responses to the Reviewers' Comments

We sincerely appreciate the reviewers' constructive comments that have allowed us to improve our manuscript. We noticed that a similar study which investigated the effect of beetroot supplementation on thermoregulatory and cardiovascular responses was recently published in European Journal of Applied Physiology (Kent et al. Effect of dietary nitrate supplementation on thermoregulatory and cardiovascular responses to submaximal cycling in the heat, Eur J Appl Physiol 118:657-668, 2018). Given that their findings strength our current study, we have decided to add this information to the discussion for blood pressure regulation (P10, L313-320). In addition, based on the comment 4 from reviewer #2, we presented CVC as absolute values (AU/mmHg) but not as % of baseline in the revised manuscript. Thus, the results obtained from core temperature thresholds and slopes for CVC were somewhat changed in the revised manuscript. The absolute CVC analysis revealed that there was no influence of beetroot juice supplementation on the thresholds and slopes for CVC during exercise (P20, Table 3) which is more consistent to general findings in the present study.

### Reviewer #1

#### Comment 1

This is very nice study with strong physiological background. Although no effect of beetroot juice on heat loss responses, this study provides important information to advance our understanding of how oral intake of beetroot juice can modulate heat loss. Paper is well organized and concise. I have some minor comments specifically for discussion and interpretation of authors results. The authors do not necessarily reflect all of my comments in the manuscript, as some of comments are just my thoughts.

We sincerely appreciate the positive comment.

#### Comment 2

Although the authors rational is based on peripheral mechanisms, since taking beetroot can influence both central and peripheral mechanisms, is it possible that central increase in NO modulates for example thermoregulatory center thereby modulating efferent signaling to thermoeffectors? My understanding is that cardiovascular response can be influenced by central NO bioavailability based on animal studies.

This reviewer raised an important point. While the precise influence of beetroot juice supplementation on the central thermoregulatory mechanisms is unknown in the present study, it is traditionally considered that the shift of core temperature threshold for heat loss responses would reflect the activity of thermoregulatory center in the brain (e.g., Nadel et al. JAP 37:515-520, 1974). Given that we did not find any differences in the core temperature thresholds for sweating and CVC responses in the revised analysis based on absolute CVC (Table 3), it could be assumed that the beetroot juice supplementation does not affect the central thermoregulatory mechanisms in the present study.

#### Comment 3

Did authors measure respiratory variables such as VO<sub>2</sub> during exercise? I think beetroot juice can lower VO<sub>2</sub> during exercise (increased muscle efficiency), which may affect heat production? ultimately affect rate of increase in core temperature??

As the reviewer suggested, there might be a possibility that beetroot juice supplementation lowered VO<sub>2</sub> during exercise; however, we did not measure VO<sub>2</sub> in the present study. A recent study reported that beetroot juice supplementation lowers VO<sub>2</sub> but elevates rectal temperature during exercise in hot condition (Kuennen et al. EJAP 2015). Therefore, it is unknown and

difficult to reveal how beetroot juice supplementation affected heat generation in the present study.

**Comment 4**

P4, L119

> "following the ingestion of 8 mmol NO<sub>3</sub><sup>-</sup> following ingestions" fix the text as repeating "following ingestion" twice.

As the reviewer noted, we have revised the manuscript as shown below.

P4, line 118

"...systolic blood pressure following the ingestion of 8 mmol NO<sub>3</sub> ~~following ingestions~~ (Breese et al. ...."

**Comment 5**

P9, "We further assumed that NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> in sweat appearing onto the skin would be reduced to NO, and..." this is interesting idea, perhaps future study evaluating forearm immersion to NO<sub>3</sub><sup>-</sup> or NO<sub>2</sub><sup>-</sup>-rich water would increase sweating and cutaneous vasodilation is warranted.

We agree with the reviewer's comment. Indeed, it has been reported that the topical application of a prodrug that generate NO can penetrate skin and induce cutaneous vasodilation (Vercelino et al. J Mater Sci: Mater Med, 2013). It would be interesting to investigate if this is true for sweating at rest and during exercise.

**Comment 6**

As for the potential mechanisms underpinning no effect of beetroot juice on sweating, it may be that NO<sub>3</sub><sup>-</sup> or NO<sub>2</sub><sup>-</sup> does not pass through vessels, or does not enter into sweat gland such that no change in NO bioavailability in sweat glands. Perhaps measuring NO<sub>3</sub><sup>-</sup> or NO<sub>2</sub><sup>-</sup> in sweat can answer this possibility. Please consider adding this point.

As the reviewer pointed out, we do not know if the NO<sub>3</sub><sup>-</sup> and/or NO<sub>2</sub><sup>-</sup> actually arrived at the sweat glands in the present study. We have addressed this point in the discussion in the revised manuscript as shown below.

P11, Line 338-342

*"Limitations*

There were several limitations in the present study. Firstly, while we observed increases in plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations, it was unclear whether NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> delivery to sweat glands, and by extension the potential for NO synthesis, was increased in the present study. Future research should assess sweat NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations to verify or refute this possibility."

**Comment 7**

P9, L288 regarding individual variation, did the authors see some individuals show some improvement in sweating or cutaneous vasodilation??? I guess the authors did some analysis with VO<sub>2</sub>max, then perhaps better to briefly discuss in term of VO<sub>2</sub>max..

As the reviewer suggested, there were some subjects who showed increased heat loss responses after beetroot juice supplementation compared with placebo trial during exercise. However, we are unable to clarify if these effects were due to supplementation or between-day or between-site variation. We have analyzed individual data set as a function of VO<sub>2</sub>max while we did not see results that could be explained meaningfully. Thus, we decided not to revise this

point in the revised manuscript.

#### **Comment 8**

Can the authors discuss why blood pressure lowering effect occurred at the end of exercise only??

This comment indicates an important point. We observed a trend for the reduction of MAP at 15 min ( $P = 0.093$ ) and 20 min ( $P = 0.060$ ) during exercise. Thus, while the precise reason is unknown, it is assumed that the exercising time is an important factor for the attenuation of MAP during exercise in hot condition. We expect that the lowering blood pressure at the end of exercise might be associated with the fall of blood pH and  $PO_2$  that potentiate the reduction of  $NO_2^-$  to NO. However, given that we could not assess this possibility in the present study, future study is needed to confirm this possibility. We have revised the manuscript to address this point in the revised manuscript as shown below.

P8, line 231-237, Results

“A supplementation  $\times$  time interaction effect was observed for MAP ( $P = 0.035$ ,  $\eta_p^2 = 0.265$ ,  $1-\beta = 0.782$ , Fig. 1). Post hoc analysis revealed that the BR supplementation lowered MAP during exercise, which attained significance after 30 min of exercise ( $112 \pm 6$  and  $103 \pm 6$  mmHg for PL and BR, respectively,  $P = 0.021$ ,  $\eta_p^2 = 0.559$ ,  $1-\beta = 0.724$ ,  $CI_{95} = -15$  to  $-2$  mmHg) but not 15 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.093$ ,  $\eta_p^2 = 0.350$ ,  $1-\beta = 0.389$ ,  $CI_{95} = -11$  to  $1$  mmHg) or 20 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.060$ ,  $\eta_p^2 = 0.418$ ,  $1-\beta = 0.489$ ,  $CI_{95} = -11$  to  $0$  mmHg) of exercise (Fig. 1).”

P11, line 330-334

“It is also interesting that we observed the reduction of MAP at the end of exercise only. It is expected that the lowering blood pressure at the end of exercise might be associated with the fall of blood pH and  $PO_2$  that potentiate the reduction of  $NO_2^-$  to NO (Castello et al. 2006; Modin et al. 2001) while future investigation is needed to confirm this possibility.”

#### **Reviewer #2**

##### **Comment 1**

In the current study, Amano et al. examined the effects of 3-day dietary nitrate supplementation with beetroot juice on local sweating and cutaneous vascular responses during exercise in the heat. The authors identified a modest reduction in mean arterial pressure at end-exercise following beetroot juice supplementation, but no effects on local sweating or cutaneous vascular conductance were evident at either chest or forearm skin sites. The data appear to be carefully collected and I generally agree with the conclusions drawn from the results. However I do have some concerns with the study in its current form that need to be addressed.

We appreciate for the reviewer's suggestion and comments. We have revised the manuscript based on the comments.

##### **Comment 2**

Although females in this study may have technically been tested within a given phase of the menstrual cycle, the 10-day gap between trials suggests that circulating estrogen levels may have been very different between experimental sessions for some females. For example, during the follicular phase estrogen levels are lowest within the first 5-7 days, then a sharp

increase in estrogen occurs between days 7-14 as ovulation approaches. Since estrogen has well-established effects on NO bioavailability and cutaneous vasodilator responses to heating it would have been more appropriate to test only males or to test females 1-month apart to avoid this potential limitation.

Thank you very much for suggesting this important point. As the reviewer suggested, there might be a possibility that the menstrual cycle was not well controlled for in the female participants and that potential sex differences affected the results in the present study. However, given the small number of participants for males (n=5) and female (n=3), it is difficult to compare the results between males and females to interpret the data set meaningfully. Thus, we have decided to address this issue as a limitation in the revised manuscript as shown below.

P11, line 345-350

“Finally, while we tried to conduct female experiments in the same phase of menstrual cycle, there remained a possibility that the circulating sex hormone levels differed between the trials since we did not measure blood sex hormones concentrations in the present study. Given that the sex hormone levels might affect local cutaneous blood flow response through NO dependent mechanism (Charkoudian et al. 1999), this point is worthy of future study.”

### **Comment 3**

How was sample size determined for this study? Previous work demonstrating fairly modest effects of beetroot juice on CVC during local and whole body heating used similar sample sizes, however, in these studies CVC data were expressed as %maximum, which improves measurement precision considerably over absolute or %baseline values. Given the current data are presented as absolute SKBF (AU) and CVC normalized to %baseline, the relatively poor between-site and between-day reliability for these approaches necessitates a larger sample size to make meaningful inferences about the cutaneous vasodilator response, unless the anticipated effect size is large. This issue needs to be clearly addressed by the authors.

We determined the sample size based on the mean and SD of a previous study reported an increase in CVC%max in response to local heating following beetroot juice supplementation (Keen et al. Microvas Res 98:48-53, 2014) which suggested a minimal sample size of n=4 with 80% power and  $\alpha = 0.05$ . Thus we thought that the sample size n=8 would be adequate in the present study, however, we did not consider the method to normalize CVC for the sample size calculation. As the reviewer suggested, this issue may limit the findings in the present study. Thus, we decided to address a limitation about the reliability and the power of the CVC results as shown below.

P11, line 342-345

“Secondly, given that we did not normalize CVC as % of maximum vasodilation as has previously been conducted (Keen et al. 2014; Levitt et al. 2015), the potential inter-day and inter-site variations in cutaneous vascular response might have influenced the reliability of CVC in the present study.”

### **Comment 4**

Beyond the issue of reliability, expressing CVC as %baseline has produced some confusing results here. In figure 2, SKBF is higher on the chest compared to the forearm during exercise. If the authors had presented absolute CVC (AU/mmHg), blood pressure would have been accounted for and the same general trend between forearm and chest sites during exercise would have been evident. However, with the data converted to %baseline CVC, it now appears that conductance on the chest is lower than the forearm for each condition, which does not accurately reflect what is going on at both skin sites. Since CVC is typically very low during

rest under normothermic conditions, modest changes can have a large impact on the results when normalizing to this value, especially when a large vasodilatory response occurs. Even though baseline CVC is very low for both skin sites, it is still higher on average for the chest compared to the forearm. This means that for the same absolute CVC value during exercise, the response will appear much smaller when even a minor increase in baseline CVC occurs. Since measurement reliability is also an issue here, I would highly recommend reporting absolute CVC values over %baseline when normalizing to maximum is not an option.

The reviewer suggested an important point. As suggested, the expression of CVC would have an important impact in the present study and thus we decided to present absolute CVC instead of CVC of %BL in the revised manuscript. As the reviewer suggested, general trend between CVC and SkBF became similar indicating higher CVC on the chest than that of forearm. We have revised the manuscript based on absolute CVC throughout. The changes in discussion was shown as below.

P9-10, line 294-304

“It has recently been reported that NO<sub>3</sub><sup>-</sup> supplementation increased CVC during passive heating (Levitt et al. 2015). These authors also reported that the increased CVC was due to a reduction in MAP during normothermic resting and passive hyperthermic conditions, whilst the SkBF per se was not influenced by the supplementation (Levitt et al. 2015). We did not observe measurable differences in CVC between conditions (Fig. 2) despite a reduction in MAP during exercise (Fig. 1). **Given that the CVC was not measurably impacted by BR supplementation in the present study (Fig. 2), despite a reduction in mean arterial pressure during exercise (Fig. 1), it appears that BR supplementation has a distinct influence on cutaneous vascular response between whole body passive heating and exercise. However, the mechanisms for the disparate effects of BR supplementation on cutaneous blood flow during exercise and rest in hyperthermic conditions are unknown and therefore warrants further research.**”

#### **Comment 5**

1. It is not always clear if the p-values being reported are for main effects or for individual comparisons. Please clarify this in the abstract and results.

Thank you very much for suggesting this point. Based on the reviewer's comment, we have improved the abstract and results to clarify what the specified P values relate to.

#### **Comment 6**

2. Please be consistent with reporting of p-values throughout the manuscript. In some cases raw p-values are reported and in other cases P<0.05 is used.

Based on the reviewer's comment, we have revised the manuscript to specify the actual P values.

#### **Comment 7**

Please include confidence intervals for effect size estimates.

Based on the reviewer's comment, we have included the confidence intervals to explain the results as shown below.

P7-8, line 220-257

“**RESULTS**



### *Plasma nitrate and nitrite concentrations*

Compared with PL, three days BR juice supplementation increased resting plasma  $\text{NO}_3^-$  [ $P = 0.000$ ,  $d = 4.916$ ,  $1-\beta = 1.000$ , 95% confidence interval for mean difference ( $\text{CI}_{95}$ ) = 390 to 729  $\mu\text{M}$ ] and  $\text{NO}_2^-$  ( $P = 0.011$ ,  $d = 2.222$ ,  $1-\beta = 1.000$ ,  $\text{CI}_{95} = 88$  to 410  $\mu\text{M}$ , Table 1).

### *Cardiovascular, thermal, and perceived parameters*

There were no differences in HR ( $P = 0.262$ ,  $d = 0.190$ ,  $1-\beta = 0.110$ ,  $\text{CI}_{95} = -1$  to 5 beats/min) and MAP ( $P = 0.173$ ,  $d = 0.416$ ,  $1-\beta = 0.344$ ,  $\text{CI}_{95} = -9$  to 2 mmHg) at rest between PL and BR supplementations (Table 2). Resting  $T_{\text{es}}$  ( $P = 0.069$ ,  $d = 0.667$ ,  $1-\beta = 0.704$ ,  $\text{CI}_{95} = -0.01$  to 0.23  $^{\circ}\text{C}$ ),  $T_{\text{b}}$  ( $P = 0.051$ ,  $d = 0.118$ ,  $1-\beta = 0.635$ ,  $\text{CI}_{95} = 0$  to 0.20  $^{\circ}\text{C}$ ), and  $T_{\text{sk}}$  ( $P = 0.616$ ,  $d = 0.526$ ,  $1-\beta = 0.504$ ,  $\text{CI}_{95} = -0.23$  to 0.36  $^{\circ}\text{C}$ ) were not different in BR compared with PL (Table 2). A supplementation  $\times$  time interaction effect was observed for MAP ( $P = 0.035$ ,  $\eta_p^2 = 0.265$ ,  $1-\beta = 0.782$ , Fig. 1). Post hoc analysis revealed that the BR supplementation lowered MAP during exercise, which attained significance after 30 min of exercise ( $112 \pm 6$  and  $103 \pm 6$  mmHg for PL and BR, respectively,  $P = 0.021$ ,  $\eta_p^2 = 0.559$ ,  $1-\beta = 0.724$ ,  $\text{CI}_{95} = -15$  to -2 mmHg) but not 15 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.093$ ,  $\eta_p^2 = 0.350$ ,  $1-\beta = 0.389$ ,  $\text{CI}_{95} = -11$  to 1 mmHg) or 20 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.060$ ,  $\eta_p^2 = 0.418$ ,  $1-\beta = 0.489$ ,  $\text{CI}_{95} = -11$  to 0 mmHg) of exercise (Fig. 1). The attenuation of MAP in BR relative to PL at 30 min of exercise was related to the levels of  $\dot{V}_{\text{O}_{2\text{max}}}$  such that individuals with smaller  $\dot{V}_{\text{O}_{2\text{max}}}$  showed a larger attenuation of MAP ( $P = 0.048$ ,  $R^2 = 0.50$ ). Neither a main effect of supplementation (all  $P \geq 0.129$ , all  $\eta_p^2 \leq 0.298$ , all  $1-\beta \leq 0.319$ ) nor an interaction (all  $P \geq 0.069$ , all  $\eta_p^2 \leq 0.312$ , all  $1-\beta \leq 0.529$ ) was observed for HR,  $T_{\text{es}}$ ,  $T_{\text{sk}}$ ,  $T_{\text{b}}$ , and RPE during exercise (Fig. 1).

### *Sweating and cutaneous vascular responses*

Neither a main effect of supplementation ( $P = 0.164$ ,  $\eta_p^2 = 0.256$ ,  $1-\beta = 0.270$ ) nor an interaction effect (all  $P \geq 0.121$ , all  $\eta_p^2 \leq 0.250$ , all  $1-\beta \leq 0.437$ ) was observed in SR during exercise (Fig. 2). Similarly, there were no main effects of supplementation (all  $P \geq 0.114$ , all  $\eta_p^2 \leq 0.318$ , all  $1-\beta \leq 0.346$ ) and skin region (all  $P \geq 0.089$ , all  $\eta_p^2 \leq 0.358$ , all  $1-\beta \leq 0.401$ ) or these interaction effect (all  $P \geq 0.135$ , all  $\eta_p^2 \leq 0.289$ , all  $1-\beta \leq 0.309$ ) for  $T_{\text{es}}$  and  $T_{\text{b}}$  thresholds and slopes for SR (Table 3). A higher SkBF and CVC on the chest compared to the forearm was observed as indicated by a significant main effect of skin region during exercise (SkBF;  $P = 0.008$ ,  $\eta_p^2 = 0.660$ ,  $1-\beta = 0.883$ ,  $\text{CI}_{95} = 0.116$  to 0.530 AU, CVC;  $P = 0.012$ ,  $\eta_p^2 = 0.619$ ,  $1-\beta = 0.823$ ,  $\text{CI}_{95} = 0.001$  to 0.012 AU/mmHg, Fig. 2). The BR supplementation and regional difference did not affect  $T_{\text{es}}$  and  $T_{\text{b}}$  thresholds and slopes for CVC such that there were no main effects of supplementation (all  $P \geq 0.087$ , all  $\eta_p^2 \leq 0.360$ , all  $1-\beta \leq 0.403$ ) and skin region (all  $P \geq 0.079$ , all  $\eta_p^2 \leq 0.377$ , all  $1-\beta \leq 0.427$ ) or these interaction effect (all  $P \geq 0.305$ , all  $\eta_p^2 \leq 0.149$ , all  $1-\beta \leq 0.161$ ) for  $T_{\text{es}}$  and  $T_{\text{b}}$  thresholds and slopes for CVC (Table 3).”

### **Comment 8**

Please address spelling mistakes throughout the manuscript.

We have double-checked the manuscript for the spelling mistakes.

### **Reviewer #3**

#### **Comment 1**

The study is on an interesting and relevant topic, with the potential mechanistic linkage between nitrate supplementation and blood flow/sweating clearly laid out in the Introduction.

The Methods is very sound in terms of research design, the Results are clearly presented, and the Discussion puts the results and also the existing literature clearly into context. Very nicely done.

We appreciate the very positive comment for this manuscript.

[Click here to view linked References](#)

## **Influence of dietary nitrate supplementation on local sweating and cutaneous vascular responses during exercise in a hot environment**

Tatsuro Amano<sup>1,2</sup>, Dai Okushima<sup>3</sup>, Brynmor C. Breese<sup>4</sup>, Stephen J. Bailey<sup>5</sup>, Shunsaku Koga<sup>3</sup>,  
and Narihiko Kondo<sup>1</sup>

<sup>1</sup>Laboratory for Applied Human Physiology, Graduate School of Human Development and Environment, Kobe University, Kobe, Japan

<sup>2</sup>Laboratory for Exercise and Environmental Physiology, Faculty of Education, Niigata University, Niigata, Japan

<sup>3</sup>Applied Physiology Laboratory, Kobe Design University, Kobe, Japan

<sup>4</sup>School of Biomedical & Healthcare Sciences, Plymouth University, Plymouth, United Kingdom

<sup>5</sup>School of Sport, Exercise and Health Sciences, Loughborough University, United Kingdom

*Running head:* Beetroot juice and heat loss responses during exercise

*Address for correspondence:*

Narihiko KONDO, PhD,

Laboratory for Applied Human Physiology,

Graduate School of Human Development and Environment, Kobe University

3-11 Tsurukabuto, Nada-ku, Kobe 657-8501, Japan

Tel: +81-78-803-7816, Fax: +81-78-803-7929

E-mail: kondo@kobe-u.ac.jp

**ABSTRACT**

**Purpose:** We investigated the influence of inorganic nitrate ( $\text{NO}_3^-$ ) supplementation on local sweating and cutaneous vascular responses during exercise in hot conditions. **Method:** Eight healthy, young subjects were assigned in a randomized, double-blind, crossover design to receive  $\text{NO}_3^-$ -rich beetroot (BR) juice (140 mL/day, containing  $\sim 8$  mmol of  $\text{NO}_3^-$ ) and  $\text{NO}_3^-$ -depleted placebo (PL) juice (140 mL/day, containing  $\sim 0.003$  mmol of  $\text{NO}_3^-$ ) for 3 days. On day 3 of supplementation, subjects cycled at an intensity corresponding to 55% of  $\dot{V}\text{O}_{2\text{max}}$  for 30 minutes in hot conditions (30°C, 50% relative humidity). Chest and forearm sweat rate (SR) and skin blood flow (SkBF), were measured continuously. Cutaneous vascular conductance (CVC) was calculated by SkBF/mean arterial pressure (MAP). **Results:** Prior to exercise, plasma  $\text{NO}_3^-$  ( $21 \pm 6$  and  $581 \pm 161$   $\mu\text{M}$ ) and nitrite ( $\text{NO}_2^-$ ,  $87 \pm 28$  and  $336 \pm 156$  nM) concentrations were higher after BR compared to PL supplementation ( $P \leq 0.011$ ,  $n=6$ ). Oesophageal, mean skin, and mean body temperatures during exercise were not different between conditions. **In addition, BR supplementation did not affect SR, SkBF, and CVC during exercise.** A lower MAP was found **after 30 minutes of exercise** following BR supplementation ( $112 \pm 6$  and  $103 \pm 6$  mmHg for PL and BR, respectively,  $P = 0.021$ ). **Conclusion:** These results suggest that inorganic  $\text{NO}_3^-$  supplementation, which increases the potential for  $\text{O}_2$ -independent NO production, does not affect local sweating and cutaneous vascular responses, but attenuates blood pressure in young healthy subjects exercising in a hot environment.

**KEYWORDS:** Nitric oxide synthesis, thermoregulation, heat loss response, sweat glands

**ABBREVIATIONS:** ANOVA, analysis of variance; d, Cohen's d; CVC, cutaneous vascular conductance; HR, heart rate;  $\dot{V}\text{O}_{2\text{max}}$ , maximal oxygen uptake; MAP, mean arterial blood pressure;  $T_b$ , mean body temperature;  $T_{\text{sk}}$ , mean skin temperature;  $\text{NO}_3^-$ , nitrate; NO, nitric oxide; NOS, nitric oxide synthase;  $\text{NO}_2^-$ , nitrite;  $T_{\text{es}}$ , oesophageal temperature;  $\eta_p^2$ , partial eta-squared; RPE, rating of perceived exertion;  $T_{\text{re}}$ , rectal temperature; SkBF, skin blood flow; SD, standard deviation; SR, sweat rate

## 31 INTRODUCTION

32  
33 Sweating and cutaneous vasodilation are vital physiological functions that dissipate heat from  
34 the body during exercise. Previous studies suggest that nitric oxide (NO) is an important  
35 signalling molecule for modulating sweat rate (SR) and cutaneous blood flow in humans  
36 (Stapleton et al. 2014; Welch et al. 2009; Kellogg et al. 1998; McNamara et al. 2014; Wilkins  
37 et al. 2003; Fujii et al. 2016). There are two pathways for NO generation in humans. The most  
38 recognized is the enzymatic NO synthase (NOS) pathway, which catalyses the oxidation of L-  
39 arginine to NO and L-citrulline (Moncada and Higgs 1991). More recently, it has been shown  
40 that NO can be produced O<sub>2</sub>-independently through the stepwise reduction of inorganic nitrate  
41 (NO<sub>3</sub><sup>-</sup>) to nitrite (NO<sub>2</sub><sup>-</sup>) and subsequently NO (i.e. NO<sub>3</sub><sup>-</sup>→NO<sub>2</sub><sup>-</sup>→NO pathway) (Lundberg et  
42 al. 2008). The importance of NOS-derived NO on physiological responses that promote heat  
43 loss is already well defined, as evidenced by a lower SR and cutaneous vasodilation during  
44 exercise or passive heat stress following inhibition of skin NOS activity (Welch et al. 2009;  
45 Kellogg et al. 1998; Wilkins et al. 2003; Stapleton et al. 2014; Fujii et al. 2016; Amano et al.  
46 2017a). On the other hand, the influence of the NO<sub>3</sub><sup>-</sup>→NO<sub>2</sub><sup>-</sup>→NO pathway on heat loss  
47 responses during exercise has not been fully investigated.

48  
49 Following ingestion, NO<sub>3</sub><sup>-</sup> is absorbed and concentrated by the salivary glands for delivery to  
50 the oral cavity for second pass metabolism (Spiegelhalter et al. 1976). Here, oral microflora  
51 catalyses the reduction of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup> (Duncan et al. 1995). Ingested NO<sub>2</sub><sup>-</sup> is subsequently  
52 reduced to NO and other reactive nitrogen species in the acidic pH of the stomach (Benjamin  
53 et al. 1994). It is also clear that a portion of the ingested NO<sub>2</sub><sup>-</sup> passes into the systemic  
54 circulation, as evidenced by a dose-dependent increase in venous plasma [NO<sub>2</sub><sup>-</sup>] after oral NO<sub>3</sub><sup>-</sup>  
55 ingestion (Kapil et al. 2010; Wylie et al. 2013a). As this circulating NO<sub>2</sub><sup>-</sup> arrives at the skin  
56 microvasculature, the ensuing fall in P<sub>O2</sub> (Kerger et al. 1995) would be conducive to the  
57 reduction of NO<sub>2</sub><sup>-</sup> to NO (Castello et al. 2006) and might promote increases in NO-mediated  
58 cutaneous vasodilation (Kellogg et al. 1998; Fujii et al. 2016; Wilkins et al. 2003; Shastry et al.  
59 1998; McNamara et al. 2014). It is also possible for circulating NO<sub>2</sub><sup>-</sup> to pass into the eccrine  
60 sweat glands (Weller et al. 1996). Subsequently, NO<sub>2</sub><sup>-</sup> might be reduced to NO, a reaction that  
61 would be facilitated by the acidic pH present in eccrine sweat (Morimoto and Johnson 1967).  
62 In addition, NO<sub>3</sub><sup>-</sup> secreted in sweat might undergo reduction to NO<sub>2</sub><sup>-</sup> when exposed to dermal  
63 NO<sub>3</sub><sup>-</sup> reductases with this NO<sub>2</sub><sup>-</sup> undergoing subsequent reduction to NO within the acidic

64 conditions of the skin (Burry et al. 2001; Weller et al. 1996). This dermal NO then has the  
65 potential to diffuse through the skin to promote vasodilation (Vercelino et al. 2013). Therefore,  
66 NO<sub>3</sub><sup>-</sup> supplementation has the potential to augment sweating and cutaneous vascular responses  
67 via NO-mediated signalling during exercise.

68  
69 In contrast to the postulate that NO<sub>3</sub><sup>-</sup> supplementation has the potential to augment SR, it has  
70 recently been reported that dietary NO<sub>3</sub><sup>-</sup> supplementation does not affect whole body sweat loss  
71 (indirectly inferred from changes in body mass) during submaximal treadmill walking in hot  
72 conditions (Kuennen et al. 2015). However, it is important to note the large inter-regional  
73 differences in local SR and skin blood flow (SkBF) previously reported across human skin  
74 (Havenith et al. 2008; Smith and Havenith 2011; Taylor and Machado-Moreira 2013; Kuno  
75 1956; Hertzman and Randall 1948). Since higher SkBF would deliver more NO<sub>2</sub><sup>-</sup> to the sweat  
76 gland, NO<sub>3</sub><sup>-</sup> supplementation might be particularly effective at augmenting local SR at skin  
77 sites where blood flow is high (e.g. torso) compared to skin sites where blood flow is low (e.g.  
78 extremes) (Hertzman and Randall 1948). It has been reported that NO<sub>3</sub><sup>-</sup> supplementation can  
79 increase cutaneous vasodilation to local heating (Keen et al. 2014) and whole body passive  
80 heat stress (Levitt et al. 2015). However, since disparate mechanisms underlie cutaneous blood  
81 flow regulation at rest and during exercise (McNamara et al. 2014; Fujii et al. 2016) and since  
82 the influence of NO<sub>3</sub><sup>-</sup> supplementation on regional SkBF has not been investigated, further  
83 research is required to explore whether the greater cutaneous blood flow after NO<sub>3</sub><sup>-</sup>  
84 supplementation is also manifest during exercise, and whether these effects might be site-  
85 specific.

86  
87 The purpose of the present study was to investigate the influence of NO<sub>3</sub><sup>-</sup>-rich beetroot juice  
88 (BR) supplementation on local sweating and cutaneous vascular responses during exercise in  
89 a hot environment. We hypothesized that BR supplementation would augment local sweating  
90 and cutaneous vasodilation on the chest to a greater extent than on the forearm during exercise  
91 in a hot condition.

## 92 93 **MATERIALS AND METHODS**

### 94 95 *Ethical approval*

96 Each participant was informed of the purpose and procedures of the study prior to providing  
97 written informed consent. This study was approved by the Human Subjects Committee of the

98 Graduate School of Human Development and Environment, Kobe University (Kobe, Japan),  
99 and conformed to the standards set forth in the latest revision of the Declaration of Helsinki.

100

#### 101 *Participants*

102 Five males and three females participated in the present study (mean  $\pm$  SD age:  $24 \pm 4$  years,  
103 height:  $1.70 \pm 0.09$  m, and mass:  $62.7 \pm 10.3$  kg, maximum oxygen uptake,  $\dot{V}O_{2\max}$ :  $43 \pm 6$   
104 ml/kg/min). Participants were healthy and active and were excluded if they had history of  
105 hypertension, heart disease, diabetes, autonomic disorders or smoking. All participants were  
106 not currently taking prescription medication. None of the females were using oral  
107 contraceptives and all participated in the experimental testing sessions either during the self-  
108 reported follicular or luteal phases without crossing phases. All experiments were conducted  
109 between the month of June and August.

110

#### 111 *Dietary intervention*

112 Participants were randomly assigned in a crossover, double-blind design to receive 3 days of  
113 dietary supplementation with  $\text{NO}_3^-$ -rich beetroot juice (BR) (140 mL/day;  $\sim 8$  mmol  $\text{NO}_3^-$ ; Beet  
114 It, James White Drinks, Ipswich, UK) or  $\text{NO}_3^-$ -depleted BR as a placebo (PL; 140 mL/day;  
115 0.0034 mmol  $\text{NO}_3^-$ ; Beet It, James White Drinks, Ipswich, UK). The dose of BR administered  
116 was based on a previous dose-response study reporting an increase in plasma  $\text{NO}_3^-$  and  $\text{NO}_2^-$   
117 concentration and peak reduction in systolic blood pressure following the ingestion of 8 mmol  
118  $\text{NO}_3^-$  (Breese et al. 2017; Cermak et al. 2012; Lansley et al. 2011; Kuennen et al. 2015). The  
119  $\text{NO}_3^-$ -depleted BR placebo beverage was identical in color, taste, smell and texture to the  
120 experimental  $\text{NO}_3^-$ -rich BR beverage. The PL beverage was created by passage of the juice,  
121 before pasteurization, through a column containing Purolite A520E ion exchange resin, which  
122 selectively removes  $\text{NO}_3^-$  ions. Four participants began with the BR condition, and the other  
123 four participants began with the PL condition. The subjects were instructed to consume the  
124 beverages (70 mL in the morning and afternoon) on days 1-2 of the supplementation period.  
125 On day 3, the subjects were instructed to consume the beverages over a 10-min period, 2 h  
126 prior to the start of the exercise test (see below), based on recent evidence that plasma [ $\text{NO}_2^-$ ]  
127 peaks at approximately 2-2.5 h post-administration of BR containing 8.4 mmol  $\text{NO}_3^-$  (Wylie et  
128 al. 2013b). A 7-day washout period separated each supplementation period. Throughout the  
129 study, participants were asked to refrain from consumption of green leafy vegetables (e.g.  
130 Spinach), processed meats (e.g. Bacon), and Japanese traditional foods (e.g. Seaweed,

131 Sayaingen beans, Chin gin cai) which are high in  $\text{NO}_3^-$  (Sobko et al. 2010). Since the oral  
132 bacteria are integrated for reducing  $\text{NO}_3^-$  to  $\text{NO}_2^-$  in vivo (Govoni et al. 2008), participants were  
133 also asked to refrain the use of mouthwash.

134

#### 135 *Exercise protocol*

136 After arrival at the laboratory on experimental days, venous blood samples were drawn from  
137 an antecubital vein in a seated position in an air-conditioned room ( $\sim 27^\circ\text{C}$ ) from 6 of 8 subjects  
138 who consented to venipuncture. All exercise trials were performed in an environmental  
139 chamber (SR-3000; Nagano Science, Osaka, Japan) maintained at an ambient temperature of  
140  $30^\circ\text{C}$  and relative humidity of 50% with minimal air movement. Upon entering the chamber,  
141 participants rested in the semi-supine position for a minimum of 60 minutes while instruments  
142 were attached. After recording the baseline data for 5 minutes, participants started cycling at  
143 an exercise intensity of 55% of maximum oxygen uptake ( $\dot{V}\text{O}_{2\text{max}}$ ) for 30 minutes.

144

#### 145 *Measurements*

146 Oesophageal temperature ( $T_{\text{es}}$ ) was measured continuously using a thermocouple temperature  
147 probe (Inui Engineering, Higashi Osaka, Japan). The tip of the probe was covered by silicon  
148 and inserted at a distance of one-fourth of the participant's standing height from the external  
149 nares past the nostril and into the esophagus. Skin temperatures were measured at six skin sites  
150 using the same thermocouples attached with surgical tape. Mean skin temperature ( $T_{\text{sk}}$ ) was  
151 calculated using 6 skin temperatures weighted to the regional proportions determined as  
152 follows: forehead 7%, abdomen 35%, forearm 14%, hand 5%, lower leg 13%, and foot 7%  
153 (Mitchell and Wyndham 1969). The mean body temperature ( $T_{\text{b}}$ ) was calculated as  $0.8 \times T_{\text{es}} +$   
154  $0.2 \times T_{\text{sk}}$  (Stolwijk and Hardy 1966).

155

156 Local SR was measured continuously on left ventral forearm (centre of the forearm) and chest  
157 (under the left clavicle) using a ventilated plastic capsule ( $3.14\text{ cm}^2$ ) that was attached to the  
158 skin using collodion. Anhydrous nitrogen gas was passed through each capsule over the skin  
159 surface at a rate of  $0.7\text{ L}\cdot\text{min}^{-1}$ . Water content from the effluent air was measured using a  
160 capacitance hygrometer (HMP50; Vaisala, Helsinki, Finland). An index of local SkBF on the  
161 forearm and chest were measured continuously using laser-Doppler velocimetry (ALF21;  
162 Advance, Tokyo, Japan) located adjacent to the ventilated capsule. Cutaneous vascular  
163 conductance (CVC) was calculated from the ratio of SkBF to mean arterial blood pressure

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164 (MAP). All temperature, SR and SkBF data were recorded at 1-s intervals using a data logger  
165 (MX100; Yokogawa, Tokyo, Japan) and simultaneously displayed (MX100 standard software;  
166 Yokogawa, Tokyo, Japan) and recorded. Heart rate (HR) and MAP were continuously measured  
167 from left middle finger using the Finometer system (Finometer; Finapres Medical Systems,  
168 Amsterdam, The Netherlands). **Standardized** calibration was conducted before each trial.  
169 **Ratings** of perceived exertion (RPE) was measured **every** 5 minutes based on Borg 6-20 scale  
170 (Borg 1970).

171  
172 Venous blood samples (~4 ml) were drawn into lithium-heparin tubes (7.5 ml Monovette  
173 Lithium Heparin, Sarstedt, Leicester, UK), which have very low levels of  $\text{NO}_2^-$  and  $\text{NO}_3^-$ .  
174 Within 3 min of collection, the samples were centrifuged at 2700 g and 4°C for 10 min. Plasma  
175 was extracted and immediately frozen at -80°C for later analysis of  $\text{NO}_2^-$  and  $\text{NO}_3^-$  using a  
176 modification of the chemiluminescence technique (Bateman et al. 2002). All glassware,  
177 utensils, and surfaces were rinsed with deionized water to remove residual  $\text{NO}_2^-$  and  $\text{NO}_3^-$  prior  
178 to analysis. Following defrosting at room temperature, the  $\text{NO}_2^-$  of the undiluted (non-  
179 deproteinized) plasma was determined by its reduction to NO in the presence of glacial acetic  
180 acid and 4% (w/v) aqueous NaI. The spectral emission of electronically excited nitrogen  
181 dioxide, **produced** from the reaction of NO with ozone, was detected by a thermoelectrically  
182 cooled, red-sensitive photomultiplier tube housed in a Sievers gas-phase chemiluminescence  
183 nitric oxide analyzer (Sievers NOA 280i. Analytix Ltd, Durham, UK). The  $\text{NO}_2^-$  **concentration**  
184 was determined by plotting signal (mV) area against a calibration plot of 100 nM to 1  $\mu\text{M}$   
185 sodium nitrite. Before determination of  $\text{NO}_3^-$ , samples were deproteinized using zinc sulfate  
186 ( $\text{ZnSO}_4$ )/sodium hydroxide (NaOH) precipitation. Aqueous  $\text{ZnSO}_4$  [300  $\mu\text{l}$  5% (w/v)] and 500  
187  $\mu\text{l}$  0.18 M NaOH were added to 100  $\mu\text{l}$  of sample and vortexed for 30 s before being left to  
188 stand at room temperature for 15 min. Thereafter, samples were centrifuged at 4,000 rpm for 5  
189 min, and the supernatant was removed for subsequent analysis. The  $\text{NO}_3^-$  **concentration** of the  
190 deproteinized plasma sample was determined by its reduction to NO in the presence of 0.8%  
191 (w/v) vanadium trichloride in 1 M HCl. The production of NO was detected using the  
192 chemiluminescence nitric oxide analyzer, as described above.

193

#### 194 *Data and statistical analyses*

195 Variables were averaged for 5 minutes at pre-exercise baseline and for every 1 minute during  
196 exercise. SR and CVC were plotted against the changes in  $T_{\text{es}}$  ( $\Delta T_{\text{es}}$ ) and  $T_{\text{b}}$  ( $\Delta T_{\text{b}}$ ) during

197 exercise to assess the core temperature threshold and slope for inducing the responses.  
198 Segmented regression analysis was used to determine the core temperature onset thresholds  
199 and slopes of local SR and cutaneous vasodilation at each skin site (Cheuvront et al. 2009).  
200 The slopes were defined based on the linear portion of the changes in SR and CVC before and  
201 after the appearance of the onset thresholds during the exercise.

202  
203 Baseline data in the BR and PL conditions were compared using a paired Student's t-test.  $T_{es}$   
204 and  $T_b$  thresholds and slopes for SR and CVC between BR and PL were compared using two  
205 way-repeated measures ANOVAs (condition  $\times$  skin region). HR, MAP,  $T_{es}$ ,  $T_{sk}$ , and RPE during  
206 exercise were compared using two way-repeated measures ANOVAs (condition  $\times$  time) with  
207 comparisons of baseline and each 5 minutes of exercise (baseline, 5, 10, 15, 20, 25, and 30  
208 minutes). Three way-repeated measures ANOVAs were performed (condition  $\times$  time  $\times$  skin  
209 region) for SR and CVC during exercise. A Greenhouse-Geisser correction was applied if the  
210 assumption of sphericity was been violated. A Bonferroni correction was applied to control for  
211 the multiple comparisons. When an influence of BR supplementation was observed, a linear  
212 regression analysis was performed to determine the relationship between  $\dot{V}O_{2max}$  and the  
213 variables (see results). The effect size of each ANOVA was calculated and reported as partial  
214 eta-squared values ( $\eta_p^2$ ) and that of each t-test was calculated and reported as Cohen's d (d).  
215 Data are presented as mean  $\pm$  SD, and statistical significance was set at  $P < 0.05$ . All statistical  
216 analyses were performed using a statistical package (SPSS) version 24.0.

## 217 218 **RESULTS**

### 219 220 *Plasma nitrate and nitrite concentrations*

221 Compared with PL, three days BR juice supplementation increased resting plasma  $NO_3^-$  [ $P =$   
222 0.000,  $d = 4.916$ ,  $1-\beta = 1.000$ , 95% confidence interval for mean difference ( $CI_{95}$ ) = 390 to 729  
223  $\mu M$ ] and  $NO_2^-$  ( $P = 0.011$ ,  $d = 2.222$ ,  $1-\beta = 1.000$ ,  $CI_{95} = 88$  to 410  $\mu M$ , Table 1).

### 224 225 *Cardiovascular, thermal, and perceived parameters*

226 There were no differences in HR ( $P = 0.262$ ,  $d = 0.190$ ,  $1-\beta = 0.110$ ,  $CI_{95} = -1$  to 5 beats/min)  
227 and MAP ( $P = 0.173$ ,  $d = 0.416$ ,  $1-\beta = 0.344$ ,  $CI_{95} = -9$  to 2 mmHg) at rest between PL and BR  
228 supplementations (Table 2). Resting  $T_{es}$  ( $P = 0.069$ ,  $d = 0.667$ ,  $1-\beta = 0.704$ ,  $CI_{95} = -0.01$  to  
229 0.23  $^{\circ}C$ ),  $T_b$  ( $P = 0.051$ ,  $d = 0.118$ ,  $1-\beta = 0.635$ ,  $CI_{95} = 0$  to 0.20  $^{\circ}C$ ), and  $T_{sk}$  ( $P = 0.616$ ,  $d =$

0.526,  $1-\beta = 0.504$ ,  $CI_{95} = -0.23$  to  $0.36$  °C) were not different in BR compared with PL (Table 2). A supplementation  $\times$  time interaction effect was observed for MAP ( $P = 0.035$ ,  $\eta_p^2 = 0.265$ ,  $1-\beta = 0.782$ , Fig. 1). Post hoc analysis revealed that the BR supplementation lowered MAP during exercise, which attained significance after 30 min of exercise ( $112 \pm 6$  and  $103 \pm 6$  mmHg for PL and BR, respectively,  $P = 0.021$ ,  $\eta_p^2 = 0.559$ ,  $1-\beta = 0.724$ ,  $CI_{95} = -15$  to  $-2$  mmHg) but not 15 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.093$ ,  $\eta_p^2 = 0.350$ ,  $1-\beta = 0.389$ ,  $CI_{95} = -11$  to  $1$  mmHg) or 20 min ( $110 \pm 2$  and  $105 \pm 4$  mmHg, respectively,  $P = 0.060$ ,  $\eta_p^2 = 0.418$ ,  $1-\beta = 0.489$ ,  $CI_{95} = -11$  to  $0$  mmHg) of exercise (Fig. 1). The attenuation of MAP in BR relative to PL at 30 min of exercise was related to the levels of  $\dot{V}O_{2max}$  such that individuals with smaller  $\dot{V}O_{2max}$  showed a larger attenuation of MAP ( $P = 0.048$ ,  $R^2 = 0.50$ ). Neither a main effect of supplementation (all  $P \geq 0.129$ , all  $\eta_p^2 \leq 0.298$ , all  $1-\beta \leq 0.319$ ) nor an interaction (all  $P \geq 0.069$ , all  $\eta_p^2 \leq 0.312$ , all  $1-\beta \leq 0.529$ ) was observed for HR,  $T_{es}$ ,  $T_{sk}$ ,  $T_b$ , and RPE during exercise (Fig. 1).

#### *Sweating and cutaneous vascular responses*

Neither a main effect of supplementation ( $P = 0.164$ ,  $\eta_p^2 = 0.256$ ,  $1-\beta = 0.270$ ) nor an interaction effect (all  $P \geq 0.121$ , all  $\eta_p^2 \leq 0.250$ , all  $1-\beta \leq 0.437$ ) was observed in SR during exercise (Fig. 2). Similarly, there were no main effects of supplementation (all  $P \geq 0.114$ , all  $\eta_p^2 \leq 0.318$ , all  $1-\beta \leq 0.346$ ) and skin region (all  $P \geq 0.089$ , all  $\eta_p^2 \leq 0.358$ , all  $1-\beta \leq 0.401$ ) or these interaction effect (all  $P \geq 0.135$ , all  $\eta_p^2 \leq 0.289$ , all  $1-\beta \leq 0.309$ ) for  $T_{es}$  and  $T_b$  thresholds and slopes for SR (Table 3). A higher SkBF and CVC on the chest compared to the forearm was observed as indicated by a significant main effect of skin region during exercise (SkBF;  $P = 0.008$ ,  $\eta_p^2 = 0.660$ ,  $1-\beta = 0.883$ ,  $CI_{95} = 0.116$  to  $0.530$  AU, CVC;  $P = 0.012$ ,  $\eta_p^2 = 0.619$ ,  $1-\beta = 0.823$ ,  $CI_{95} = 0.001$  to  $0.012$  AU/mmHg, Fig. 2). The BR supplementation and regional difference did not affect  $T_{es}$  and  $T_b$  thresholds and slopes for CVC such that there were no main effects of supplementation (all  $P \geq 0.087$ , all  $\eta_p^2 \leq 0.360$ , all  $1-\beta \leq 0.403$ ) and skin region (all  $P \geq 0.079$ , all  $\eta_p^2 \leq 0.377$ , all  $1-\beta \leq 0.427$ ) or these interaction effect (all  $P \geq 0.305$ , all  $\eta_p^2 \leq 0.149$ , all  $1-\beta \leq 0.161$ ) for  $T_{es}$  and  $T_b$  thresholds and slopes for CVC (Table 3).

## DISCUSSION

Contrary to our hypothesis, BR supplementation did not affect local SR and cutaneous vascular responses on the chest or forearm during exercise in hot conditions. On the other hand, we observed a lowered end-exercise blood pressure following BR supplementation during exercise

263 in hot conditions. These results suggest that  $\text{NO}_3^-$ -rich BR juice supplementation is not likely  
264 to influence local sweating and cutaneous vascular responses, but can lower systemic blood  
265 pressure during exercise in a hot environment.

266

267 Previous studies have reported a fundamental role for NO in the regulation of sweating during  
268 exercise, as evidenced by a reduction in SR when NOS activity was inhibited at the skin (Welch  
269 et al. 2009; Fujii et al. 2016; Fujii et al. 2015). In the present study, plasma  $\text{NO}_3^-$  and  $\text{NO}_2^-$  were  
270 significantly increased by BR supplementation (Table 1), implying an increased potential for  
271  $\text{O}_2$ -independent NO production (Lundberg et al. 2008). We reasoned that BR supplementation  
272 would increase  $\text{NO}_2^-$  delivery to sweat glands where cutaneous blood flow was higher, thereby  
273 promoting an enhanced sweat response mediated by NO (Welch et al. 2009; Fujii et al. 2016;  
274 Fujii et al. 2015). We further assumed that  $\text{NO}_3^-$  and  $\text{NO}_2^-$  in sweat **secreted** onto the skin would  
275 be reduced to NO, and hence may have diffused through the skin to increase SkBF (Vercelino  
276 et al. 2013). However, the BR-induced increase in plasma  $\text{NO}_3^-$  and  $\text{NO}_2^-$  did not affect local  
277 SR on either the forearm or chest (Fig. 2). In addition, slopes describing the relationship  
278 between sweating response on the chest and forearm against the increase in core temperature  
279 were not affected by BR supplementation (Table 3). Therefore, contrary to the previously  
280 reported influence of NOS-dependent NO production on sweat regulation (Welch et al. 2009;  
281 Fujii et al. 2016; Fujii et al. 2015), it appears that augmenting the  $\text{NO}_3^- \rightarrow \text{NO}_2^- \rightarrow \text{NO}$  pathway  
282 does not modify the sweat response during exercise in the heat, at least following **short-term**  
283 BR administration (**3 days**) employed herein. Given that  $\text{NO}_2^-$ -derived NO production is  
284 potentiated within hypoxic and acidic tissues (Lundberg et al. 2008), there remains the  
285 possibility that exogenous  $\text{NO}_3^-$ -supplementation may modulate sweating in hot environments  
286 at simulated altitude or during high- intensity exercise when NOS-dependent sweating is  
287 abolished (Fujii et al. 2014) as well as in **exercising** older individuals (Stapleton et al. 2014).  
288 In addition, given that NOS-dependent sweating is highly variable between individuals  
289 (Amano et al. 2017a; Amano et al. 2017b), it is conceivable that **enhancing** the  $\text{NO}_3^- \rightarrow \text{NO}_2^-$   
290  $\rightarrow \text{NO}$  pathway may benefit some (but not all) individuals via **an improved** sweating response  
291 when exercising in the heat. Therefore, further studies are required to elucidate the precise  
292 influence of inorganic  $\text{NO}_3^-$  treatment on sweating during exercise.

293

294 It has recently been reported that  $\text{NO}_3^-$  supplementation increased CVC during passive heating  
295 (Levitt et al. 2015). These authors also reported that the increased CVC was due to a reduction

296 in MAP during normothermic resting and passive hyperthermic conditions, whilst the SkBF  
297 per se was not influenced by the supplementation (Levitt et al. 2015). We did not observe  
298 measurable differences in CVC between conditions (Fig. 2) despite a reduction in MAP during  
299 exercise (Fig. 1). Given that the CVC was not measurably impacted by BR supplementation in  
300 the present study (Fig. 2), despite a reduction in mean arterial pressure during exercise (Fig. 1),  
301 it appears that BR supplementation has a distinct influence on cutaneous vascular response  
302 between whole body passive heating and exercise. However, the mechanisms for the disparate  
303 effects of BR supplementation on cutaneous blood flow during exercise and rest in  
304 hyperthermic conditions are unknown and therefore warrants further research.

305  
306 Numerous studies have reported a reduction in blood pressure at rest (Bailey et al. 2009; Keen  
307 et al. 2014; Levitt et al. 2015; Larsen et al. 2006; Sobko et al. 2010; Wylie et al. 2013a; Lee et  
308 al. 2015) and during exercise (Lee et al. 2015; Bond Jr et al. 2013) following  $\text{NO}_3^-$   
309 supplementation. Whilst we did not observe a reduction in blood pressure at rest with  $\text{NO}_3^-$   
310 treatment (Table 2), this lack of effect has also been reported in some previous studies following  
311  $\text{NO}_3^-$  supplementation (Cermak et al. 2012; Larsen et al. 2010; Gilchrist et al. 2013). In contrast  
312 to previous studies that reported an influence of  $\text{NO}_3^-$  supplementation on blood pressure within  
313 thermoneutral ambient conditions, it is noteworthy that we reported a lowering of blood  
314 pressure with BR during exercise in a hot environment. On the other hand, a very recent study  
315 reported that BR supplementation does not alter blood pressure during exercise in trained  
316 cyclists ( $\dot{V}\text{O}_{2\text{max}}$ , 68 ml/kg/min) in hot conditions (Kent et al. 2018). Given that our participants  
317 were comparatively less trained ( $\dot{V}\text{O}_{2\text{max}}$ , 43 ml/kg/min) to those assessed in the study by Kent  
318 et al. (2018), it is possible that aerobic fitness accounted for the inter-study disparity in blood  
319 pressure following BR supplementation during exercise in hot conditions. To support this  
320 observation, we found that individuals with lower aerobic fitness manifest a larger attenuation  
321 of blood pressure during exercise in a hot environment. Notwithstanding this novel observation,  
322 given that unstable and falling blood pressure can signal cardiovascular failure during exercise  
323 in the heat (Rowell 1974), our data suggest that ingesting  $\text{NO}_3^-$ -rich BR prior to exercising in  
324 the heat should be implemented with caution, particularly since its effect on exercise  
325 performance in a hot environment is currently controversial (Kent et al. 2017; McQuillan et al.  
326 2017). While the potential ergogenic effects of BR supplementation appear to be inversely  
327 related to aerobic fitness (Porcelli et al. 2015) and is recommended to enhance endurance  
328 performance in recreationally-active individuals in thermoneutral conditions (Jones 2014), BR  
329 supplement should be used with caution in hot environments to limit the potential for the

330 development of excessive hypotension. It is also interesting that we observed the reduction of  
331 MAP at the end of exercise only. It is expected that the lowering blood pressure at the end of  
332 exercise might be associated with the fall of blood pH and PO<sub>2</sub> that potentiate the reduction of  
333 NO<sub>2</sub><sup>-</sup> to NO (Castello et al. 2006; Modin et al. 2001) while future investigation is needed to  
334 confirm this possibility. Clearly, further studies are required to elucidate the impact and safety  
335 of the blood pressure lowering effects of BR supplementation during exercise in hot conditions.

### 337 *Limitations*

338 There were several limitations in the present study. Firstly, while we observed increases in  
339 plasma NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations, it was unclear whether NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> delivery to sweat  
340 glands, and by extension the potential for NO synthesis, was increased in the present study.  
341 Future research should assess sweat NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup> concentrations to verify or refute this  
342 possibility. Secondly, given that we did not normalize CVC as % of maximum vasodilation as  
343 has previously been conducted (Keen et al. 2014; Levitt et al. 2015), the potential inter-day and  
344 inter-site variations in cutaneous vascular response might have influenced the reliability of  
345 CVC in the present study. Finally, while we tried to conduct female experiments in the same  
346 phase of menstrual cycle, there remained a possibility that the circulating sex hormone levels  
347 differed between the trials since we did not measure blood sex hormones concentrations in the  
348 present study. Given that the sex hormone levels might affect local cutaneous blood flow  
349 response through NO dependent mechanism (Charkoudian et al. 1999), this point is worthy of  
350 future study.

351  
352 In summary, we showed that three days of BR juice supplementation, which increased plasma  
353 NO<sub>3</sub><sup>-</sup> and NO<sub>2</sub><sup>-</sup>, had no influence on sweating and cutaneous vascular responses at multiple  
354 skin sites during exercise in a hot condition among healthy young adults. However, BR juice  
355 supplementation lowered mean arterial blood pressure whilst exercising in the heat. Further  
356 research is required to assess the risk-reward weighting of this hypotensive effect during  
357 exercise in a hot environment.

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5 532 **CONFLICTS OF INTEREST**

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7 533 None.

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540 **FIGURE LEGENDS**

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2 542 **Figure 1.** Heart rate (HR), mean arterial blood pressure (MAP), oesophageal temperature ( $T_{es}$ ),  
3 543 mean skin temperature ( $T_{sk}$ ), mean body temperature ( $T_b$ ), and ratings of perceived exertion  
4 (RPE) during exercise in PL and BR conditions. # indicates a significant difference between  
5 conditions at a given time point ( $P = 0.021$ ).  
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11 547 **Figure 2.** Sweat rate (SR), skin blood flow (SkBF), and cutaneous vascular conductance (CVC)  
12 on forearm and chest during exercise in PL and BR conditions.  
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552 **Table 1.** Plasma **nitrate** and **nitrite** concentrations.

	PL	BR
NO <sub>3</sub> <sup>-</sup> (μM)	21 (6)	581 (161) *
NO <sub>2</sub> <sup>-</sup> (nM)	87 (28)	336 (156) *

553 The values given are the means (SD). NO<sub>3</sub><sup>-</sup>, nitrate; NO<sub>2</sub><sup>-</sup>, nitrite. \*Significantly higher than  
 554 that of PL ( $P \leq 0.011$ ).

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557 **Table 2.** Physiological variables at rest.

	PL	BR
HR (beats/min)	63 (11)	65 (10)
MAP(mmHg)	89 (8)	85 (11)
T <sub>es</sub> (°C)	36.87 (0.12)	36.98 (0.20)
T <sub>sk</sub> (°C)	34.42 (0.58)	34.48 (0.42)
T <sub>b</sub> (°C)	36.38 (0.18)	36.48 (0.20)

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559 The values given are the means (SD). HR, heart rate; MAP, mean arterial blood pressure; T<sub>es</sub>,  
 560 oesophageal temperature; T<sub>sk</sub>, mean skin temperature; T<sub>b</sub>, mean body temperature.

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563 **Table 3.** Oesophageal and mean body temperatures thresholds and slopes for sweating and  
 564 cutaneous vasodilation during exercise.

		SR		CVC	
		PL	BR	PL	BR
Forearm					
T <sub>es</sub>	Threshold (°C)	36.98 (0.21)	37.07 (0.24)	37.06 (0.16)	37.16 (0.25)
	ΔThreshold (°C)	0.11 (0.16)	0.09 (0.12)	0.19 (0.18)	0.18 (0.16)
	slopes (mg/cm <sup>2</sup> /min/°C)	1.27 (0.46)	1.43 (0.45)	-	-
	slopes (AU/mmHg/°C)	-	-	0.0150 (0.0052)	0.0215 (0.0117)
T <sub>b</sub>	Threshold (°C)	36.41 (0.21)	36.50 (0.22)	36.49 (0.14)	36.56 (0.21)
	ΔThreshold (°C)	0.03 (0.09)	0.01 (0.04)	0.11 (0.16)	0.08 (0.08)
	slopes (mg/cm <sup>2</sup> /min/°C)	1.73 (0.70)	1.92 (0.76)	-	-
	slopes (AU/mmHg/°C)	-	-	0.0222 (0.0082)	0.0260 (0.0142)
Chest					
T <sub>es</sub>	Threshold (°C)	37.01 (0.21)	37.10 (0.27)	37.04 (0.15)	37.15 (0.24)
	ΔThreshold (°C)	0.14 (0.15)	0.12 (0.14)	0.17 (0.13)	0.17 (0.10)
	slopes (mg/cm <sup>2</sup> /min/°C)	1.58 (0.61)	2.09 (1.33)	-	-
	slopes (AU/mmHg/°C)	-	-	0.0180 (0.0062)	0.0247 (0.0146)
T <sub>b</sub>	Threshold (°C)	36.42 (0.21)	36.52 (0.23)	36.44 (0.20)	36.56 (0.21)
	ΔThreshold (°C)	0.04 (0.09)	0.04 (0.06)	0.06 (0.08)	0.09 (0.07)
	slopes (mg/cm <sup>2</sup> /min/°C)	2.15 (1.13)	2.42 (1.04)	-	-
	slopes (AU/mmHg/°C)	-	-	0.0273 (0.0082)	0.0288 (0.0144)

565 The values given are the means (SD). T<sub>es</sub>, oesophageal temperature; T<sub>b</sub>, mean body temperature; SR,  
 566 sweat rate; CVC, cutaneous vascular conductance.

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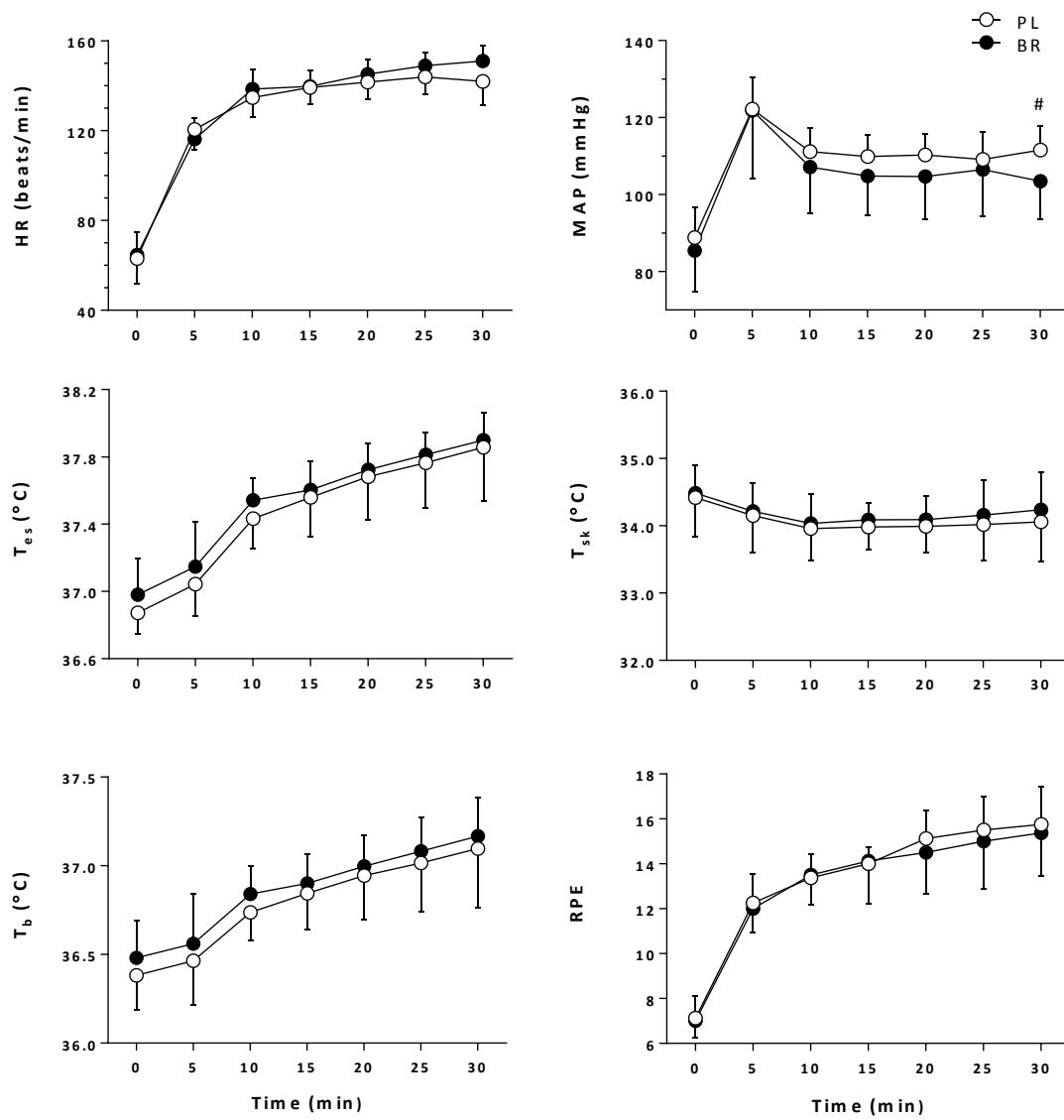


Fig. 1

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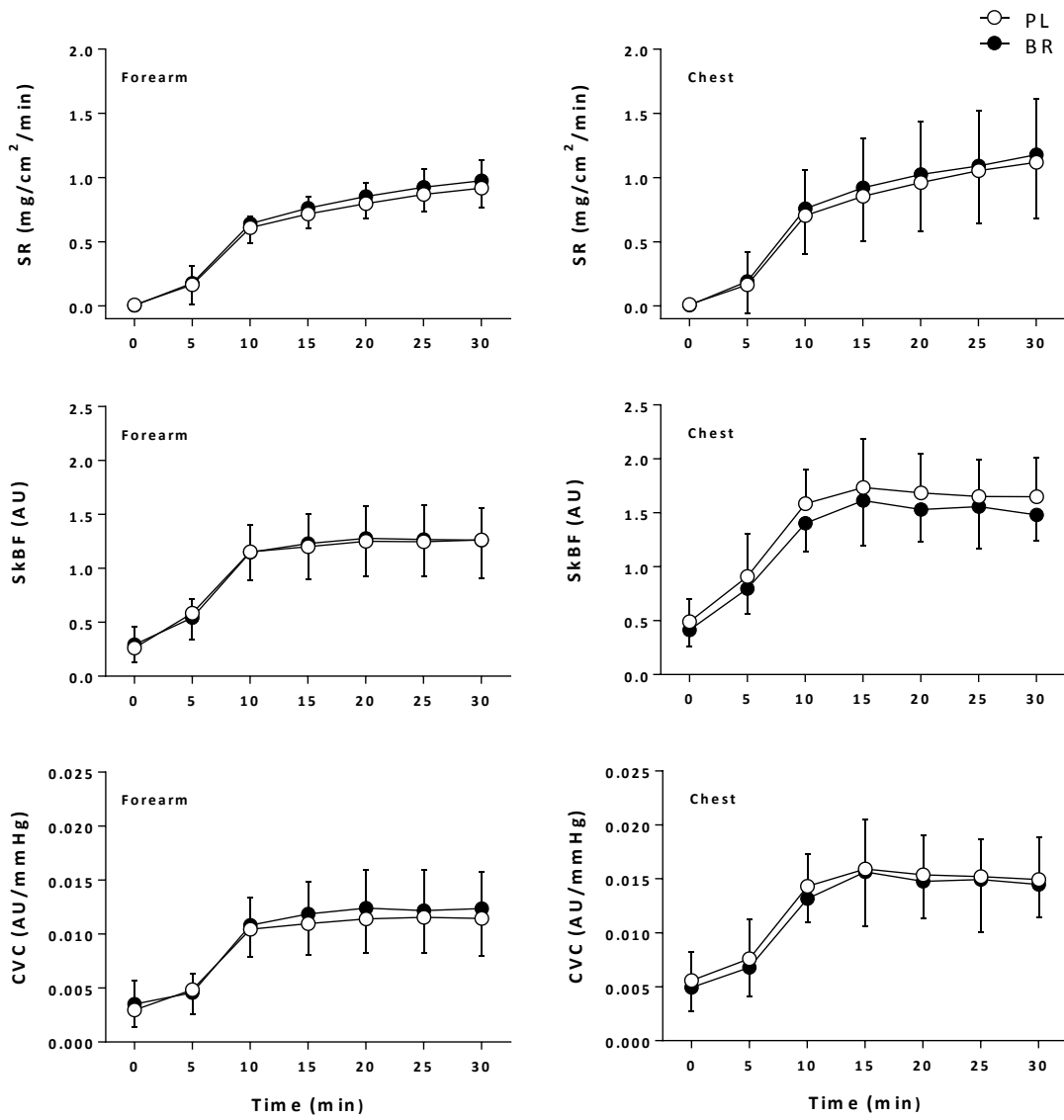


Fig. 2

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## **AUTHOR CONTRIBUTIONS**

Conception and design of research was undertaken by TA, DO, BB, and NK, data collection and analyses was undertaken by TA, DO, and BB, the manuscript was drafted by TA, DO, BB, and SB and all authors (TA, DO, BB, SB, SK, and NK) contributed to data interpretation, editing and revision of manuscript, and approved the final version.