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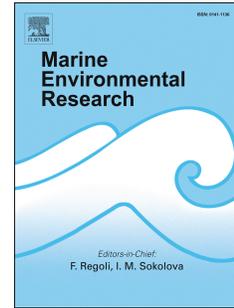
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# Accepted Manuscript

The effects of elevated CO<sub>2</sub> on shell properties and susceptibility to predation in mussels *Mytilus edulis*

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1 **Title:** The effects of elevated CO<sub>2</sub> on shell properties and susceptibility to predation in mussels

2 *Mytilus edulis*

3

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6

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9

10 **Running Head:** Altered shell properties and predation risk in mussels under OA

11

12 **Abstract:**

13 For many species, ocean acidification (OA) is having negative physiological consequences on  
14 their fitness and resilience to environmental change, but less is known about the ecosystem  
15 effects of these changes. Here, we assess how OA conditions predicted for 2100 affects the  
16 biological functioning of an important habitat-forming species *Mytilus edulis* and its  
17 susceptibility to predation by a key predator, the gastropod *Nucella lapillus*. Change in three  
18 physiological parameters in *Mytilus* were assessed: (1) shell thickness and cross-sectional  
19 surface area, (2) body volume and (3) feeding rate, as well as susceptibility to predation by  
20 *N. lapillus*. Shell thickness and cross-section area, body volume and feeding rate of *Mytilus* all  
21 reduced under OA conditions indicating compromised fitness. Predation risk increased by ~26%  
22 under OA, suggesting increased susceptibility of mussels to predation and/or altered predator  
23 foraging behaviour. Notably, predation of large *Mytilus* – that were largely free from predation  
24 under control conditions – increased by more than 8x under OA, suggesting that body size was  
25 no longer a refuge. Our results suggest OA will impact upon ecosystem structure and  
26 functioning and the continued provision of ecosystem services associated with *Mytilus* reefs and  
27 the communities associated with them.

28

29 **Keywords:** climate change, ecosystem engineer, predation, trophic cascade, environmental change;  
30 interaction

31

32 **Highlights:**

- 33
- Ocean acidification is a major threat to marine ecosystem structure and functioning
  - 2100 pCO<sub>2</sub> scenario reduced *Mytilus* shell thickness & area, body volume and feeding rate
- 34

- 35 • Predation risk susceptibility changes under future climate scenarios
- 36 • Large prey not susceptible to predation today 8x more at risk under OA scenarios
- 37 • Significant changes to ecosystem structure and functioning predicted for future

38

## 39 1. INTRODUCTION

40 Climate change is one of the greatest threats to biodiversity globally (Thomas et al., 2004) altering  
41 population and community dynamics (Parmesan and Yohe, 2003) and increasing risks of species  
42 extinction (Thomas et al., 2004). There is overwhelming evidence that human activities are driving  
43 rates of climate change (Henson et al., 2017); the continued emission of greenhouse gases is a  
44 primary driver of increasing global temperatures and ocean acidification (Caldeira and Wickett,  
45 2003). Predictions of global environmental conditions for the end of the century (e.g. RCP8.5  
46 scenario; Stocker et al., 2013) coupled with ever-increasing experimental evidence suggest wide-  
47 ranging impacts of future ocean acidification and warming (OAW) scenarios on marine life  
48 (Poloczanska et al., 2016).

49

50 Climate change may benefit some organisms. A wide-range of taxa including jellyfish, macroalgae,  
51 invertebrates and some fish (e.g. Aprahamian et al., 2010; Hall-Spencer and Allen, 2015), especially  
52 those with Lusitanian evolutionary origins (Lavergne et al., 2010), are demonstrating increased  
53 fitness over wider geographic ranges (e.g. Calosi et al., 2017). But wide-ranging negative effects of  
54 OAW have also been shown or are predicted to alter ecology, behaviour and physiology (Gazeau et  
55 al., 2013; Hughes, 2000; Lemasson et al., 2017a; Lemasson et al., 2017b). For instance, OA has been  
56 shown to alter predator-prey dynamics (Dixson et al., 2010; Harvey and Moore, 2017), intracellular  
57 pH, biological functioning (Pörtner et al., 2004), metabolism (Thomsen and Melzner, 2010), and  
58 individual energetic needs (Gray et al., 2017; Leung et al., 2017). These changes could change  
59 ecosystem structure by amplifying range shifts (Calosi et al., 2017) or cause trophic cascades through  
60 lower abundances of key species and reduced trophic transfer (Rossoll et al., 2012). In addition, a  
61 decrease in critical ecosystem services (ESs) may also occur (Lemasson et al., 2017a, b).

62

63 Molluscs and other calcifying organisms are particularly prone to environmental change and  
64 especially OA (Gazeau et al., 2013; Parker et al., 2013). Increased  $p\text{CO}_2$  has been shown to reduce  
65 calcification (but see Ries et al., 2009), alter crystalline ultrastructure (Duquette et al., 2017; Fitzer et  
66 al., 2016; Leung et al., 2017) and increase dissolution rates in oysters and mussels (Berge et al., 2006;  
67 Gazeau et al., 2007; Ries et al., 2009). These changes are predicted to alter the capacity of  
68 individuals to maintain their exoskeleton via biomineralisation of calcium carbonate mechanisms; an

69 effect illustrated by a reduction in shell thickness (e.g. Chen et al., 2015) and strength (Speights et  
70 al., 2017; Welladsen et al., 2010) in some bivalves. The effect of these changes may extend beyond  
71 the fitness of the individual, affecting the wider ecosystem by changing survivorship and/or  
72 increasing susceptibility of prey to predation (Dixson et al., 2010; Freeman and Byers, 2006) with  
73 consequences that cascade up the food chain.

74  
75 Many calcifying organisms are of ecological and economic importance, and provide numerous  
76 ecosystem services (MEA, 2005). Often ecosystem engineers (*sensu* Jones et al., 1994) or habitat-  
77 forming species, they create habitat for other species and support disproportionately high  
78 biodiversity in comparison to other habitats (Gutierrez et al., 2003). Bivalve molluscs, which include  
79 the mussel *Mytilus* spp., are especially important. Abundant worldwide, mussels account for 30% of  
80 global mollusc aquaculture, and in 2015, global production was ~16.5 million tonnes with a market  
81 value of ~\$18 billion (FAO, 2015). They also provide a number of other important supporting  
82 ecosystem services including nutrient cycling and improving water quality (Asmus and Asmus, 1991;  
83 Dame and Dankers, 1988; Pejchar and Mooney, 2009).

84  
85 Here, we test the effect of future OA scenarios of the functioning of *Mytilus* spp. Firstly, we consider  
86 how OA impacts the fitness of individuals, specifically their shell thickness, body volume, and feeding  
87 rate. We then test if changes in individual fitness alters trophic interactions strength between  
88 *Mytilus* and one of its key predators.

89

## 90 **2. MATERIALS AND METHODS**

91 Adult individuals of *M. edulis* were collected from Queen Anne's Battery Marina, Plymouth  
92 (50°21'50.8"N, 4°07'53.4"W) in October 2016. Mussels were cleaned of all epibiota and placed in  
93 tanks of seawater (temperature  $\approx$  15°C, Salinity  $\approx$  34, pH  $\approx$  8) for 2-wk to acclimatise. All mussels  
94 were measured and grouped into one of two arbitrary size classes: small (40  $\pm$  10mm) and large (60  
95  $\pm$  10mm). Mussels were fed three times a week ~3 mL (concentration = 50,000 cells/mL) of mixed  
96 shellfish diet (Shell diet 1800, Reed Mariculture, USA).

97

### 98 **2.1. Experimental design**

99 After 2-wk acclimation, 30 mussels were randomly selected from each size class and placed in tanks  
100 simulating one of two  $p\text{CO}_2$  emission scenarios (Stocker et al., 2013) representing current (~400  
101 ppm) and 2100 scenarios (1000 ppm) for 8-wk. Three replicate tanks of each treatment were  
102 established for each mussel size class (a total of 12 tanks). Five mussels were placed in each tank,

103 with each mussel marked with non-toxic nail varnish to identify individuals. Mussels were grouped  
104 by size (small and large) as it is predicted that size may: (i) influence metabolism and vulnerability to  
105 climate change (Carey et al., 2016), and (ii) predation risk is greater in smaller mussels (Navarrete  
106 and Castilla, 2003). Mussels were fed as per the acclimation period.

107

## 108 **2.2. OA system**

109 A mesocosm system was used to reach relevant CO<sub>2</sub> concentrations. Full details are described in  
110 Lemasson et al. (2017b) to allow for brevity here. For 400 ppm, each tank contained an air stone,  
111 and atmospheric air was bubbled gently into each replicate tank. For 1000 ppm, pure CO<sub>2</sub> was slowly  
112 released into a Buchner flask mixed with dry air ( $\approx 400$  ppm  $p\text{CO}_2$ ) using multistage CO<sub>2</sub> regulators  
113 (EN ISO 7291; GCE, Worksop, UK). CO<sub>2</sub> levels were monitored using a CO<sub>2</sub> analyser (LI-820; LI-COR,  
114 Lincoln, NE, USA). pH was measured three times a week using a microelectrode (InLab® Expert Pro-  
115 ISM; Mettler-Toledo Ltd, Beaumont Leys, UK) attached to a pH meter (S400 SevenExcellence;  
116 Mettler-Toledo Ltd, Beaumont Leys, UK), calibrated with NIST traceable buffers.

117

## 118 **2.3. Carbonate chemistry**

119 Total alkalinity (TA) was measured weekly using a calibrated potentiometric titrator (TitraLab  
120 AT1000© series HACH Company, USA). Three 50 mL samples were taken from each experimental  
121 tank and tested to calculate TA. Temperature was taken *in situ* using a temperature probe  
122 (HH806AU, Omega, U.K.) and salinity was recorded using a handheld refractometer (S/Mill, Atago,  
123 Tokyo, Japan). These data were used to calculate calcite and aragonite saturation, and CO<sub>2</sub>  
124 concentration in water for acidified and control conditions on a weekly basis using CO<sub>2</sub>SYS software  
125 (Lewis and Wallace, 1998) using Mehrbach solubility constants (Mehrbach et al., 1973), refitted by  
126 Dickson & Millero (1987) (see Supplementary Table 1).

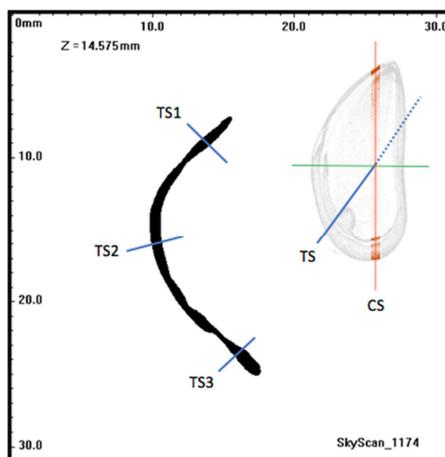
127

## 128 **2.4. Morphological and physiological parameters**

### 129 **2.4.1. Shell thickness and surface area**

130 After 8-wk, two morphological parameters were measured: shell thickness (mm) and the surface  
131 area (mm<sup>2</sup>) of each cross-section. Both metrics were measured using images collected using micro  
132 computerised tomography (microCT) (Skyscan 1174, Bruker, Germany), which produces a  
133 reconstructed 3D image of the individual made up of images taken from 3 planes (x, y, z). Scaled  
134 images of cross-sections (black shell profile; Fig. 1) of the right valve from 10 individuals per  
135 treatment were imported into ImageJ (Schneider et al., 2012). Shell thickness at three haphazardly-  
136 located transverse section points within the lip (TS1), middle (TS2) and umbo (TS3) regions of each

137 cross-section was then measured, as well as the surface area ( $\text{mm}^2$ ) of each cross-section based on  
 138 the scaled image mask (surface area of the black shell in Fig. 1). A two-sample t-test using Welch's  
 139 correction for unequal variance was used to compare shell thickness and cross-sectional surface area  
 140 between mussels from control and elevated  $p\text{CO}_2$  treatments.  
 141



142  
 143 **Figure 1.** Indicative measurement of shell thickness at three transverse section locations (TS1-3)  
 144 from a haphazardly chosen cross-section (CS) of the right valve of *Mytilus* spp. Images taken using  
 145 microCT.

#### 146 147 **2.4.2. Mussel body volume**

148 The body volume of all mussels ( $N = 60$ ) was calculated using water displacement (mL). Individual  
 149 mussels were placed in a 250 mL volumetric cylinder containing 100 mL of sea water (salinity = 35)  
 150 and the liquid displacement estimated (nearest 1 mL). Individuals were measured at the start of the  
 151 experiment ( $t_0$ ) and after 8-wk ( $t_8$ ) of exposure to control and OA conditions. A 3-factor linear mixed-  
 152 effects model (*lme*) was used to test for a change in body volume between  $p\text{CO}_2$  treatments and  
 153 mussel size class after 8-wk, incorporating 'mussel' within 'tank' as a nested random factor to take  
 154 account of the repeated measurement of the same individual. Posthoc pairwise contrasts of  
 155 significant interactions were made using the *multcomp* package in R.

#### 156 157 **2.4.3. Feeding rate**

158 Weekly over the 8-wk experiment, 30 mussels were randomly selected and starved for 24 hours.  
 159 Each individual was then placed in its own beaker with 400 mL of  $2\mu\text{m}$ -filtered seawater (salinity =  
 160 35) and a magnetic stirrer (400 rpm) to ensure the water was well mixed. Once all mussels had fully  
 161 opened, 3 mL of *Tetraselmis* at a density of 10,000 cells/ml was added to each beaker. Three  
 162 replicate 5mL water samples were taken from haphazard locations throughout the chamber (1) prior

163 to the addition of food ( $t_i$ ); (2) immediately after addition of food ( $t_0$ ) to check the initial algal  
 164 concentration; and (3) at 10 minute intervals following food addition for a duration of 30 minutes,  
 165 providing 6 sampling times (i.e.  $t_i$ ,  $t_0$ ,  $t_{10}$ ,  $t_{20}$  and  $t_{30}$ ). If the mussel shut its valves, the chronometer  
 166 was stopped and restarted once the valves re-opened. Counts of algae in all water samples were  
 167 performed in triplicate using a Coulter Counter (Beckman Coulter Z2). Clearance rates (CR) were  
 168 calculated using the following equation after Coughlan (1969):  
 169

$$CR = \frac{V \times \ln\left(\frac{C_{n-1}}{C_n}\right)}{t_n - t_{n-1}}$$

170  
 171 where CR is the clearance rate measured during the 10 minute interval between sampling times  $t_{n-1}$   
 172 and  $t_n$  ( $L \cdot h^{-1}$ ),  $V$  is the volume of the chamber in litres,  $C_{n-1}$  is the concentration ( $cell \cdot L^{-1}$ ) in the sample  
 173 taken at time  $t_{n-1}$  (h), and  $C_n$  is the concentration ( $cell \cdot L^{-1}$ ) in the sample taken at time  $t_n$  (h). Results  
 174 are presented as  $CR_{max}$  - the maximum clearance rate observed over the 30-minute feeding period.  
 175

176 A three-factor linear-mixed effects model (lme) was used to compare maximum clearance rate  
 177 between  $pCO_2$  treatments (400 ppm; 1000 ppm) and mussel size over time (fixed factor; 2, 4, 6 and 8  
 178 wk) with 'tank' included as a random factor to account for potential differences in mussel  
 179 functioning as a result of the experimental tank. Significant differences were compared using post-  
 180 hoc pairwise comparisons (Tukey HSD).  
 181

## 182 **2.5. Predation risk to mussels under control and OA conditions**

183 Change in predation risk to mussels (measured as mortality) under control and OA conditions was  
 184 assessed using a common predator of mussels, the dog whelk, *Nucella lapillus*. *Nucella* were  
 185 collected from the same location as the mussels, returned to the lab and acclimated in either the  
 186 control or OA conditions (as above) for 2-wk. During acclimation, individuals were provided with  
 187 mussels for food, but were starved for 48-h prior to being placed in the experimental tanks.  
 188

189 Predation tanks were arranged in the OA mesocosm system as described above. In total, 24 tanks (3  
 190 tanks per body size  $\times$  predator  $\times$  OA scenario) were established. In each tank, five small or large  
 191 acclimated mussels were placed with or without a single *Nucella lapillus* for 24 h, after which  
 192 mortality was measured as percentage of dead mussels. Mussels were considered dead when the  
 193 mussel shell gaped and would not close despite direct physical disturbance. In tanks without  
 194 predators, there was no mussel mortality in either the 400ppm or 1000ppm tanks. As such, there

195 was no need to include 'predator' (levels: present; absent) as a factor in the analysis, which  
 196 therefore compares mussel mortality in treatments with *Nucella* present only.

197

198 A two-factor linear mixed effects model was used to test for differences in proportional mussel  
 199 mortality between  $p\text{CO}_2$  treatments (400 ppm; 1000 ppm) and mussel size class (small; large), with  
 200 'tank' included as a random factor to account for potential differences in predation risk as a result of  
 201 the experimental tank. All statistical analyses were conducted using R (R Development Core Team,  
 202 2017)

203

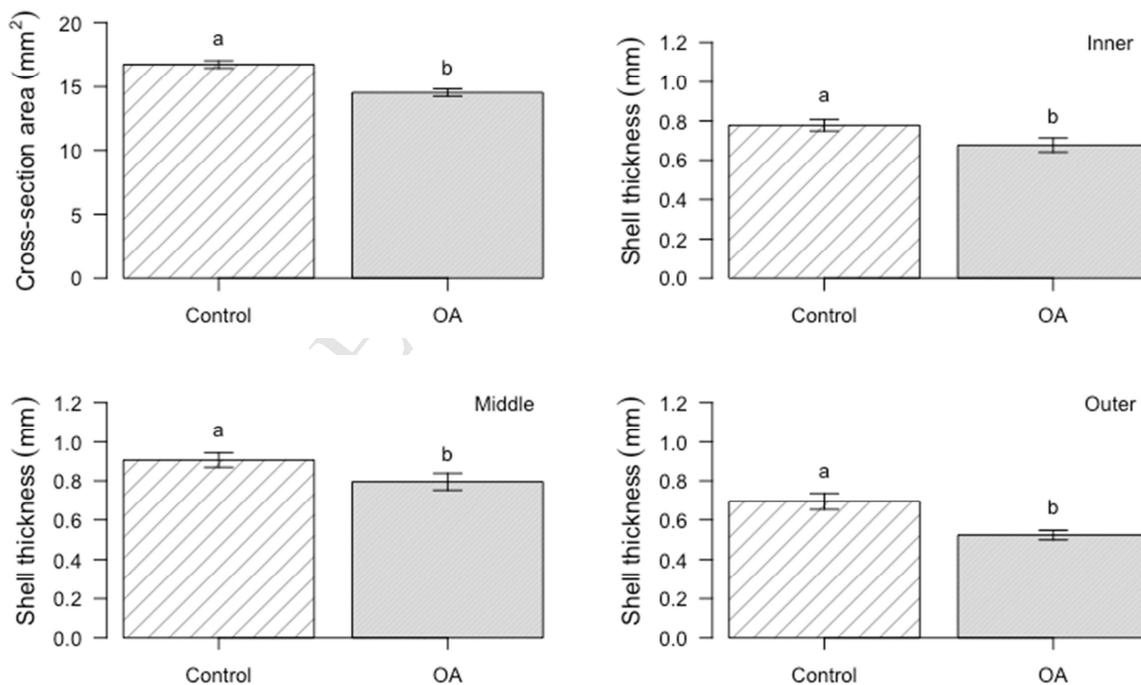
### 204 3. RESULTS

#### 205 3.1. Morphological parameters

##### 206 3.1.1. Shell thickness

207 Shells held under OA conditions were on average, 13-25% thinner and their cross-sectional surface  
 208 area  $\sim$ 13% less than mussels held under control  $p\text{CO}_2$  (400 ppm) after 8-wk. Shells became thinner at  
 209 all locations of the shell, reducing by  $\sim$ 0.10 mm (TS1), 0.11 mm (TS2) and 0.17 mm (TS3) respectively  
 210 (Fig. 2). Body size had no effect on size or surface area reductions.

211



212

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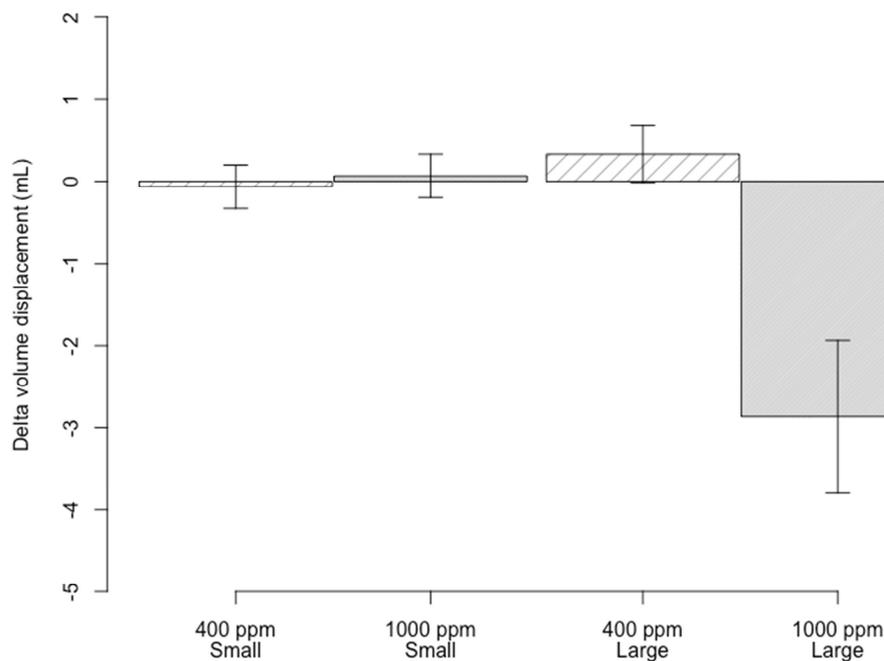
214 **Figure 2.** Mean ( $\pm$  S.E) cross-sectional surface area (mm<sup>2</sup>), and shell thickness (mm) at the inner  
 215 (TS1), middle (TS2) and outer edges (TS3) of the right valve of *M. edulis* after 8-wk exposure to

216 Control (400ppm  $p\text{CO}_2$ ) and OA (1000ppm  $p\text{CO}_2$ ) conditions. Different letters indicate significant  
 217 differences between groups ( $p < 0.05$ ).

218

### 219 3.1.2. Mussel body volume changes

220 There were significant changes in mussel body volume depending on the size of the mussel and  $p\text{CO}_2$   
 221 conditions ( $F_{1,56} = 9.85$ ,  $p < 0.01$ ; Fig. 3). There was no change in body volume in small mussels  
 222 irrespective of  $p\text{CO}_2$  level, nor in large mussels under control (400 ppm) conditions. In large mussels,  
 223 however, there was a 17% reduction in volume under 1000 ppm after 8-wk (Fig. 3).



224

225 **Figure 3.** Change in mean body volume of small and large *M. edulis* measured by water displacement  
 226 (mL  $\pm$  S.E.) after 8-wk exposure to control (400ppm  $p\text{CO}_2$ ) and OA (1000ppm  $p\text{CO}_2$ ) treatments.  
 227 Negative values indicate loss of volume.

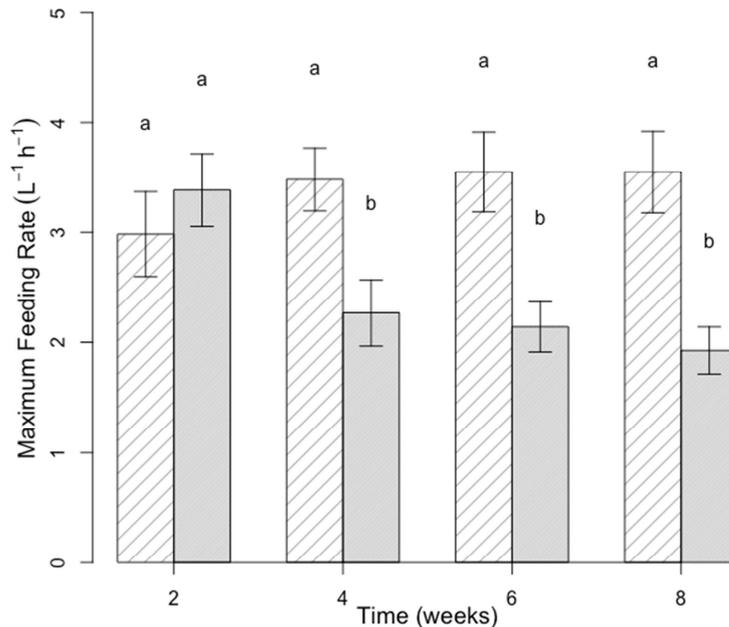
228

### 229 3.1.3. Feeding rate

230 Maximum clearance rates ( $\text{L}\cdot\text{h}^{-1}$ ) were significantly different between mussels from control and OA  
 231 treatments over time ( $F_{3,96} = 6.53$ ,  $p < 0.001$ ) but mussel body size had no effect on these changes.  
 232 After 2-wk, there was no difference in CR irrespective of mussel size or treatment, but after 4-wk,  
 233 there was a significant reduction of between ~ 35 - 47% in feeding rate in OA treatments in  
 234 comparison to the control, where no change in CR was observed (Fig. 4). After wk-4, there was no  
 235 further change in CR within the OA treatment.

236

237



238

239 **Figure 4.** Change in mean ( $\pm$  S.D.) maximum feeding rate ( $L \cdot h^{-1}$ ) of *M. edulis* (small and large) over 8-240 wk in control (400 ppm  $pCO_2$ ; hatched bars) and OA (1000 ppm  $pCO_2$ ; grey bars) treatments.241 Different letters indicate significant differences in mean values between groups ( $p < 0.05$ ).

242

243 **3.2. Predation**244 There was a significant effect of mussel size ( $F_{1,8} = 10.7$ ,  $p < 0.01$ ) and  $pCO_2$  (treatment:  $F_{1,8} = 16.7$ ,  $p <$ 245 0.01) on the mortality of *M. edulis* by *N. lapillus* (Fig. 5). Mussel mortality increased by differing246 amounts depending on mussel size. In small mussels, mortality increased by  $\sim 1.7x$  (proportional247 mortality of between 0.4-0.68), but in large mussels, mortality increased more than  $\sim 8.3x$ 

248 (proportional mortality of between 0.06 – 0.5). Comparing the ratio of mortality of small and large

249 mussels in each OA treatment, mortality risk of small mussels was 80% higher than for large mussels

250 in control treatments. In the OA treatment, however, small mussel mortality was only 26% higher

251 than that of large mussels as a result of marked increases in large mussel susceptibility to predation

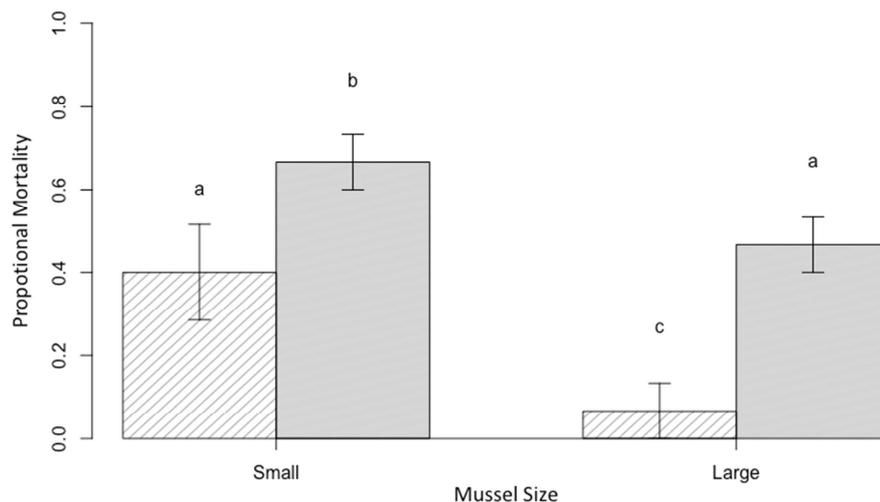
252 (Fig. 5). There was no mussel mortality in either the control or OA treatment when predators were

253 absent.

254

255

256



257  
 258 **Figure 5.** Proportional mortality (mean ± S.E.) of small and large *M. edulis* from *Nucella lapillus* in  
 259 control (400 ppm pCO<sub>2</sub>; hatched bars) and OA (1000 ppm pCO<sub>2</sub>; grey bars) treatments. Different  
 260 letters indicate significant differences between treatment means (p<0.05).

261

#### 262 4. DISCUSSION

263 Worldwide, negative consequences of OA on species performance are continuing to be shown (e.g.  
 264 Gazeau et al., 2013). Here, experiments manipulating atmospheric pCO<sub>2</sub> concentrations to match  
 265 those predicted for 2100 revealed negative impacts on a number of physiological traits in *Mytilus*  
 266 *edulis* including feeding rate, body volume and shell morphometry (shell thickness and surface area),  
 267 as well as increased predation risk from a key intertidal predator, the dogwhelk *Nucella lapillus*.

268

269 Reduced growth under OA has been shown in a number of bivalve species and other taxa (Gazeau et  
 270 al., 2013; Kroeker et al., 2013; Milano et al., 2016; Waldbusser et al., 2013), especially in those with  
 271 the capacity to divert energy from growth to other physiological processes (Michaelidis et al., 2005;  
 272 Wood et al., 2008). Energy diversion is an adaptive strategy, with mechanisms such as metabolic  
 273 upregulation used to ameliorate the negative effects of OA on processes such as shell dissolution  
 274 (Leung et al., 2017) and biomineralisation (Fitzer et al., 2016). The energetic costs of diversion  
 275 mechanisms have, in some taxa, been compensable by increased feeding (Lardies et al., 2017;  
 276 Ramajo et al., 2016; Thomsen et al., 2013), although no evidence of a response (Sanders et al., 2013)  
 277 or reduced feeding has also been shown in a number of studies (Navarro et al. 2016). Here, after 2-  
 278 wk exposure to future OA conditions, *Mytilus edulis* demonstrated a reduction in feeding (clearance  
 279 rate) of up to 47% that continued for the duration of the study. It is well documented that reduced  
 280 feeding rate can reduce scope for growth (Widdows and Johnson, 1988), and this was manifested as  
 281 reductions in shell thickness and shell surface area over the 8-wk period. Volume also reduced, but  
 282 only in large individuals. This suggests that over time, OA conditions sufficiently hampered key

283 physiological pathways (Hüning et al., 2013; Weiner and Addadi, 2011) important for shell and  
284 somatic tissue maintenance, and that adult *Mytilus edulis* - unlike juvenile conspecifics (Thomsen et  
285 al., 2013) - have limited capacity to mitigate the effects of OA by increasing feeding. Reduced feeding  
286 appears especially problematic for larger individuals who are likely to have greater basal energetic  
287 requirements than smaller individuals (Lefort et al., 2015), resulting in significant reductions in  
288 mussel volume (~17%) over relatively short time scales, with potential consequences for  
289 reproductive output and population stability for the future (Knights, 2012).

290

291 Shell maintenance is a key function for many taxa, especially calcifying species including molluscs,  
292 echinoderms and corals (Bibby et al., 2007; Gazeau et al., 2013; Maier et al., 2012; Milano et al.,  
293 2016; Riebesell et al., 2000) who use their shells for buoyancy and protection (e.g. Sherker et al.,  
294 2017). Numerous studies have shown the potential of OA to reduce an organism's capacity to obtain  
295 sufficient calcium carbonate to maintain their shell (e.g. Kroeker et al., 2014; Waldbusser et al.,  
296 2013), which over time, can lead to increased susceptibility to predation (Fitzer et al., 2015). Here,  
297 after just 8-wk exposure to 1000 ppm  $p\text{CO}_2$ , reductions in shell thickness and surface area were  
298 observed across the whole shell; the outer edge (where the shell is typically thinnest) exhibited  
299 greatest reductions (up to 25%). Molluscan shells are usually composed of two species of calcium  
300 carbonate; aragonite and calcite. In the case of *M. edulis*, while it uses a combination of aragonite  
301 (nacreous layer) and calcite (prismatic layer), it is predominantly (~83%) made up of aragonite  
302 (Hubbard et al., 1981), which is more susceptible to dissolution by carbonic acid than calcite (Feely  
303 et al., 2004). From this experiment, it is not clear which of the surfaces have eroded due to OA; we  
304 can only state that there were reductions in shell thickness and surface area. Irrespective of this,  
305 thinner, more brittle shells can be expected over relatively short-term exposures to OA. Over time,  
306 the biomineralisation process may change to compensate for this. Individuals have been shown to  
307 switch between the carbonate polymorphs used during biomineralisation, which may lead to more  
308 OA resilient shells in the future (e.g. Leung et al., 2017), but at this stage, negative consequences for  
309 shell maintenance processes are predicted.

310

311 OA conditions are likely to have wider ecosystem effects beyond changes in individual responses (i.e.  
312 changes in biological functioning). For example, a number of studies have shown that predator-prey  
313 interactions can be influenced by increased  $p\text{CO}_2$  (Bibby et al., 2007; Harvey and Moore, 2017; Lord  
314 et al., 2017; Xu et al., 2017). Differentiating between impacts on the functioning of the prey,  
315 predator or both can be difficult. In instances where the predator and prey are both likely to be  
316 under stress due to the environment (i.e. both the predator(s) and prey are calcifying species), then

317 non-linear or interactive responses are possible, with both trophic levels potentially implementing  
318 mitigation measures to counteract the stress associated with the environmental conditions. This  
319 may explain some of the inconsistencies in response to OA reported in the literature. For example,  
320 under OA, Sanford et al. (2014) found predation of *Crassostrea virginica* by predatory snails  
321 increased by 20-48%, while Harvey and Moore (2017) found *Nucella lapillus* reduced feeding to the  
322 point of starvation, releasing prey from predation. Schalkhauser et al. (2013) observed impaired  
323 clapping ability in *Pecten maximus* under OA, resulting in reduced escape performance due to  
324 diverted energetics.

325  
326 In this study, OA led to marked increases in the predation risk for mussels. Mussel mortality in OA  
327 treatments increased over control conditions to differing degrees depending on mussel size. In small  
328 mussels, proportional mortality nearly doubled (1.7x), but in large mussels, proportional mortality  
329 increased >8x from being relatively 'predation risk free' under control conditions. Size-selectivity is  
330 important in many predator-prey interactions (Thompson, 1975; Ward, 1991) and previously, *N.*  
331 *lapillus* has been shown to demonstrate strong size selectivity towards smaller mussels, in order to  
332 reduce handling time (Thompson, 1975). Increased mortality due to reduced shell thickness has  
333 been shown in other taxa (Sanford et al., 2014) and thickening of the shell is a recognised defence  
334 mechanism to limit mechanical predation (Leonard et al., 1999). Thinned shells may therefore  
335 reduce predator handling time increasing the risk of predation in larger individuals (Coleman et al.,  
336 2014; Fitzer et al., 2015). Alternatively, a change in predator foraging behaviour (Burrows and  
337 Hughes, 1989; Etter, 1996) by *Nucella* in an attempt to counter the negative effects of OA by  
338 increasing its own feeding and/or adapting its foraging strategy selecting prey of higher nutritional  
339 quality (Kohl et al., 2015) could account for the increased mortality observed. Such a mechanism  
340 may account for the differential response of *Nucella* to prey under OA seen here in comparison to  
341 the response seen in Harvey and Moore (2017) who used barnacles as a prey item, which may be  
342 less nutritious on a per individual basis. While it is not possible to differentiate between the  
343 mechanisms in this instance, the switch in foraging and significant increase in predation risk to  
344 *Mytilus* (especially to larger individuals) suggests considerable change in ecosystem structure and  
345 functioning under OA may occur (Schmitz and Trussell, 2016).

346  
347 Our results demonstrate the potential for OA to impact upon the biological functioning of individual  
348 organisms and alter predator-prey dynamics. These findings point toward considerable impacts of  
349 OA on the continued provision of ecosystem services by *Mytilus*. Beyond the biological responses  
350 shown here, our results suggest a range of ecosystem services including provisioning and regulating

351 services (MEA, 2005) may be impacted. For example, reduced clearance rates become quickly  
352 apparent, and may lead to rapid changes in nutrient load and water quality (Asmus and Asmus,  
353 1991; Broszeit et al., 2016; Dame and Dankers, 1988). Changes in shell thickness, body volume and  
354 predation risk may affect the long-term sustainability of populations across multiple trophic levels  
355 and opportunities for their exploitation for seafood. It is clear that the performance of individuals is  
356 being impacted by the ocean conditions predicted for the end of this century, and increasingly, the  
357 effect of these changes on community structure is being realised. As recently advocated by several  
358 authors (Knights et al., 2017; Runting et al., 2017), the next challenge is to combine our  
359 understanding of how individual species respond to and potentially ameliorate the effects of  
360 environmental change into a holistic ecosystem assessment in order to better understand wider  
361 ecosystem change.

362

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367

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575 **Supplemental Material**

576

577 **CO<sub>2</sub> system parameters**

578 Characteristics of the seawater in which *M. edulis* was kept is summarised in Table 1. Salinity and  
 579 temperature remained similar in both OA and control conditions. Conversely as  $p\text{CO}_2$  increased from  
 580 400 ppm (control) to 1000 ppm (OA), there was a decrease in total alkalinity, pH, dissolved  
 581 carbonate and aragonite concentrations.

582

583 **Table 1.** Seawater chemistry in the experimental tanks (mean  $\pm$  S.D.).

CO <sub>2</sub> system parameters	Control (400 ppm)	OA (1000 ppm)
pH	8.065 $\pm$ 0.02	7.619 $\pm$ 0.02
Salinity (psu)	36.16 $\pm$ 0.40	36.01 $\pm$ 0.63
Temperature (°C)	15.50 $\pm$ 0.27	15.58 $\pm$ 0.18
TA ( $\mu\text{mol.kg}^{-1}$ )	2078.83 $\pm$ 105.60	2059.83 $\pm$ 123.40
$p\text{CO}_2$ ( $\mu\text{atm}$ )	411.23 $\pm$ 25.96	1074.17 $\pm$ 68.02
CO <sub>3</sub> <sup>2-</sup> ( $\mu\text{mol.kg}^{-1}$ )	154.03 $\pm$ 4.36	63.61 $\pm$ 7.60
$\Omega_{\text{ca}}$	3.50 $\pm$ 0.21	1.39 $\pm$ 0.19
$\Omega_{\text{ar}}$	2.20 $\pm$ 0.19	0.87 $\pm$ 0.15

584 TA, total alkalinity; CO<sub>3</sub><sup>2-</sup>, carbonate ion concentration;  $\Omega_{\text{ca}}$ , saturation state of calcite;  $\Omega_{\text{ar}}$ , saturation  
 585 state of aragonite

586

587