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Ocean acidification reduces demersal zooplankton that reside in tropical coral reefs

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Abstract

The *in situ* effects of ocean acidification on zooplankton communities remain largely unexplored. Using natural volcanic CO₂ seep sites around tropical coral communities, we show a three-fold reduction in the biomass of demersal zooplankton in high-CO₂ sites compared to sites with ambient CO₂. Differences were consistent across two reefs and three expeditions. Abundances were reduced in most taxonomic groups. There were no regime shifts in zooplankton community composition and no differences in fatty acid composition between CO₂ levels, suggesting ocean acidification affects the food quantity but not the quality for nocturnal plankton feeders. Emergence trap data show that the observed reduction in demersal plankton may be partly attributable to altered habitat. Ocean acidification changes coral community composition from branching to massive bouldering coral species, and our data suggest that bouldering corals represent inferior daytime shelter for demersal zooplankton. Since zooplankton represent a major source of nutrients for corals, fish, and

other planktivores, this ecological feedback may represent a novel mechanism of how coral reefs will be affected by progressive ocean acidification.

Main text

Increased levels of anthropogenic CO₂ in the atmosphere catalyze processes that can collectively impact zooplankton communities. Concurrent with ocean warming, absorbed carbon dioxide changes the ocean chemistry by reducing seawater pH, carbonate ion concentrations, and saturation states of calcium carbonate in a process called ocean acidification^{1–4}.

Although the ramifications of ocean acidification on zooplankton communities are poorly understood, their impacts are potentially far-reaching due to their pivotal role in marine ecosystems and the carbon cycle. Zooplankton are a major food source for planktivores, and they also support bacterial and phytoplankton production through their excretion of nitrogen and phosphorus compounds⁵. Furthermore, they contribute to the biological pump as consumers of CO₂-fixing phytoplankton⁶. The subsequent sedimentation and burial of fecal pellets and zooplankton carcasses act as a sink for CO₂ that may help mitigate CO₂ emissions. Thus, in order to support predictions of the future effects of ocean acidification on marine benthic and pelagic ecosystems and CO₂ fluxes, it is essential to understand the effects of ocean acidification on zooplankton communities.

Ocean acidification studies of zooplankton have primarily focused on single-species laboratory experiments, with very few of the >7000 described species⁷ having been studied to date. Studies have reported severe direct effects

on some calcifying plankton^{8–10}, attributable to the increased energy requirements needed to acquire carbonate ions as building blocks for calcification. In contrast, existing studies suggest that non-calcifiers like copepods are generally not directly affected by ocean acidification^{11–14}. Although single-species experiments advance our understanding of the underlying mechanisms governing the direct effects of elevated CO₂ on organisms, they have limited capacity to predict the effect of ocean acidification on entire communities¹⁵. This is particularly true for zooplankton considering that calcifying species usually comprise a small proportion of the communities, and many of the non-calcifying species evaluated were generalists that are naturally found under wide ranges of environmental conditions and hence tolerate laboratory conditions^{16–19}. Therefore, to understand how ocean acidification may impact zooplankton in the future, entire communities need to be evaluated *in situ* under ocean acidification conditions.

The long-term effects of elevated carbon dioxide on marine ecosystems and entire communities have been studied at a few unique submarine volcanic CO₂ seeps. We used two such volcanic seeps in Papua New Guinea as natural laboratories, which release nearly pure CO₂ into tropical fringing coral reefs. Coral reefs are highly vulnerable to ocean acidification because of the sensitivity of their foundation species, namely corals and crustose coralline algae, and the dissolution of reef carbonate substrata at reduced pH^{20–22}. If the zooplankton that live in coral reefs are impacted by ocean acidification too, this could further strain the already jeopardized coral reefs.

The bulk of zooplankton in coral reefs are demersal, meaning the organisms live on or above substrata during the day and migrate into the water column at night^{23,24}. We compared zooplankton communities residing near CO₂ seeps with communities living at control sites. Seawater at the high-CO₂ seeps averaged 7.8 pH_T (pH at total scale; for spatial and temporal variability see Supplementary Fig. 1), while at the adjacent control sites (without seep activity) it averaged 8.0 pH_T (refs. ^{20,25}). All study sites had similar seabed topography, depth 2-3 m, tidal range <0.9 m, and longshore currents 2-4 cm s⁻¹, with an average water residence time of ~2.5 hrs over all seeps. We compared demersal zooplankton abundances, biomass, and community compositions along high-CO₂ and control sites at CO₂ seeps on Dobu and Upa-Upasina reefs using horizontal surface net tows and emergence traps on three separate expeditions.

Loss of reef-associated demersal zooplankton due to ocean acidification

At night, when the demersal zooplankton emerged, zooplankton had consistently higher biomass (mg m⁻³) at the control sites compared to the high-CO₂ sites. Across the two reefs (Dobu and Upa-Upasina) and all three expeditions, control sites had on average 2.83 (SE = 0.19) times greater zooplankton biomass than high-CO₂ sites (range: 1.45 - 4.85, N = 24; Fig. 1a). At each reef, zooplankton biomass was low and similar between CO₂ regimes during the day. On average, control sites had 9.33 (SE = 1.25) times more zooplankton biomass at night than during the day, whereas for the high-CO₂ sites that ratio was 3.14 (SE = 0.39). There was no difference in zooplankton biomass

during the day or at night between the offshore control and offshore high-CO₂ sites. Offshore sites were ~200-300 m from the coastline at water depths of 50-70 m. At night, biomass at the offshore sites averaged 3.66 (SE = 1.15) times less than the control sites, confirming that the bulk of the zooplankton were indeed resident to the reefs. The zooplankton composition also differed between offshore waters and the reef. Biomass of bulk zooplankton at control sites remained higher than at high-CO₂ sites throughout the entire night, and the diurnal migration patterns were similar between control and high-CO₂ sites (Fig. 1b).

For individual zooplankton taxa, our analyses revealed significant ($p < 0.05$) reductions in abundances (individuals m⁻³) at the high-CO₂ sites compared to control sites for most taxa, and no taxon preferred the high-CO₂ sites (Fig. 2). For example, for the copepod family Pontellidae, abundance at the high-CO₂ sites was 0.17 of that at the control sites with a 95% confidence interval of (0.09, 0.32). Additional to the CO₂ effects, abundances of some taxa also varied significantly between sites or between expeditions. A few taxa (Centropagidae, Oithonidae, Cumacea) remained unaffected by CO₂ (ratios >1.0, but standard errors including 1.0). For all other taxa, the values and 95% confidence intervals remained below 1.0, with taxonomic groups at the high-CO₂ sites reduced. Abundances for copepod taxa at the high-CO₂ were between 12-71% of those at the control sites, and for non-copepod taxa this decline in abundances at the high-CO₂ sites ranged between 19-48%. A ranking of the sensitivity of taxa showed that 10% of the taxa at the high-CO₂ sites had declined to <20% of the

control abundances, while 84% of the taxa had declined to <50%. The most sensitive copepod taxa were Monstrilloda and Pontellidae (abundance ratios = 0.12 and 0.17, respectively), and amphipods and ostracods were the most sensitive non-copepod taxa (abundance ratios = 0.19 for both).

Most resident copepod taxa were reduced in abundance, including those families predicted by laboratory experiments to be resilient to ocean acidification. One of the dominant copepod families was Acartiidae, a widely distributed group that is also known to reside demersally within coral reefs¹⁶. Acartiidae abundance was 14 times higher at the control than the high-CO₂ sites, despite previous short-term CO₂ exposure laboratory experiments suggesting that the survival, body size, developmental speed, egg production, and hatching rates of Acartiidae are negligibly affected by the magnitude of seawater pH change expected by the end of the century^{11,26,27}. This discrepancy of results highlights the need for field observations to validate laboratory predictions of direct and indirect impacts of rising CO₂ levels.

Abundances of all non-copepod zooplankton taxa were also reduced under ocean acidification, except for cumacean crustaceans. Also, zooplankton taxa that remain planktonic their entire lives (e.g. all copepods, amphipods, isopods, mysids, ostracods), and larval zooplankton that grow into larger organisms (e.g. decapod larvae, echinoderm larvae), were all reduced under ocean acidification, although to varying degrees.

No major shifts in zooplankton communities were caused by ocean acidification

Community analyses showed that there was no species turnover between the control and high-CO₂ sites. There was neither species replacement nor any taxon that proliferated in the high-CO₂ environment. There were, however, slight shifts in the percent composition of the already present taxa within the community since each taxon had a slightly different sensitivity to ocean acidification, but no new groups filled the niche or replaced other taxa in the CO₂-impacted habitat. Zooplankton communities were distinct between Upa-Upasina and Dobu reefs and between expeditions, but all had similar reactions to ocean acidification: all taxonomic groups present in the control sites persisted in the high-CO₂ sites, albeit at much lower abundances (Fig. 3).

There were also no major shifts in the biochemical composition of the zooplankton community. Specifically, the fatty acid content of bulk zooplankton samples was not different between the control and high-CO₂ sites during the second expedition (permanova: $p = 0.440$), although it did vary between the two reefs ($p = 0.001$). Zooplankton predators, including carnivorous plankton, corals and fishes, are thus likely to encounter quantitative but not biochemical changes in zooplankton food between high-CO₂ and control sites.

Reduced habitat complexity causes abundance loss for some zooplankton taxa

The causes of reduced zooplankton abundances at high-CO₂ could be due to physiological, behavioral, or ecological effects, including habitat loss and changes in the food web. At the high-CO₂ sites, coral cover is maintained with 31% and 33% hard coral coverage at the control and high-CO₂ sites, yet the composition of coral communities shifts from branching corals to massive bouldering corals with massive bouldering corals more than doubling (from 10.7% at the control sites to 24.9% cover at the high-CO₂ sites), while the structurally complex corals were reduced three fold (from 12.9% to 4.3% cover)²⁰. Coral rubble remained similar with 3.0% cover at the control sites and 2.6% cover at the high-CO₂ sites. Such loss in complexity has consequences for the organisms that rely on corals as habitat²⁸. To determine substrata preferences of the various zooplankton taxa for their daytime residence, emergence traps were placed over 1.0 m squares dominated by three different reef substrata (branching coral, massive boulder coral, and coral rubble). Emergence traps captured demersal zooplankton at night during their vertical migration when they swam into dimly illuminated (3 lumens) codends. Traps were retrieved 2-3 hours after dark, yielding a mean of 13,677 (SE = 1,948) individual zooplankton per trap at the control sites and 6,504 (SE = 787) at the high-CO₂ sites. The exact composition of the substrata within these squares was determined from photographs, distinguishing seven substrata types (branching coral, massive boulder coral, and coral rubble, sand, macro algae, turf, and other).

Data from the emergence traps showed that 16 of the 19 most common taxa of zooplankton showed reduced abundances under increased CO₂. Additionally, the abundances of 11 of the 19 taxa were positively correlated with the cover of coral rubble or branching coral (Fig. 4), of which branching coral was reduced at the high-CO₂ sites.

Nine zooplankton taxa were negatively correlated with massive boulder coral (which are abundant at high CO₂), sand, macro algae, and/or turf algae. Sand, macro algae, and turf algae were never dominant substrata in the squares at either high-CO₂ or control sites (max. 15% cover), and yet they appeared to provide shelter for some taxa (e.g. Oithonidae and Pontellidae) but were negatively associated with others (e.g. Arietellidae, Paracalanidae, Sapphirinidae). Only four zooplankton taxa showed no substratum preference. This suggests that reduced availability of branching corals at the high-CO₂ sites, and increased presence of massive bouldering corals, contributed to the reduction of several zooplankton taxa at the high-CO₂ sites.

Other causes for abundance loss

Altered habitat quality is one explanation for reduced zooplankton abundance, however other direct and indirect causes also likely contribute. Phytoplankton is food for herbivorous and omnivorous taxa (e.g. Acartiidae, Centropagidae, Harpacticoida, Oithoniidae, Oncaeididae, Paracalanidae, Pontelidae, Gastropoda larvae, Polychaeta, and Pteropoda). However, total organic carbon, total nitrogen, chlorophyll *a*, and phaeophytin concentrations did

not differ between the high-CO₂ and control sites ($p > 0.05$ for all phytoplankton). Supplementary Tables 2 and 3 show mean phytoplankton biomass values, and the significance of CO₂, reef, and time (day versus night) on affecting phytoplankton biomass). This suggests that food limitation did not control the abundances of the herbivorous and omnivorous taxa. Changes in density or nutritional quality of phytoplankton in response to high-CO₂ (ref. ²⁹) are unlikely due to the short residency time, although elevated carbon dioxide can promote phytoplankton production³⁰. The observed reductions in herbivorous and omnivorous zooplankton suggest that per capita phytoplankton availability may even increase. In contrast, zooplanktivorous zooplankton (e.g. the carnivorous Arietellidae, Corycaeidae, Sapphirinidae, Amphipoda, Decapoda larvae, Isopoda, Mysida, Ostracoda, Chaetognatha, and fish larvae) are likely to experience diminished food abundances, with potential flow-on effects on their abundances.

The impact of ocean acidification on zooplankton swimming behavior is unstudied. Zooplankton motility is a requisite for feeding, avoiding predators, and vertical migration. Our finding that migration behavior was unaffected by high-CO₂ levels at the high-CO₂ sites suggests their ability to access resources and evade predation appears to remain intact. Nevertheless, behavioral responses of individual taxa to high-CO₂ cannot be excluded as a contributing mechanism. For example, high-CO₂ disrupts discriminatory and swimming behaviors in response to olfactory cues in some tropical reef fish species^{31,32}, and similarly unexpected results are possible for some zooplankton taxa.

Zooplankton migration against vertical currents can enrich zooplankton

near reefs³³. Although the horizontal tows were not conducted directly over the bubble streams, gas bubbles at the seep sites should have enhanced vertical currents and, hence, zooplankton densities particularly for the fast-swimming larger zooplankton. The consistently lower zooplankton densities near the seep sites, for all taxa, suggest that vertical currents played no major role for explaining the observed differences in zooplankton biomass between high-CO₂ and control sites.

Biological consequences for coral reefs and marine ecosystems

Reduced zooplankton abundances may have far-reaching consequences for marine ecosystems and fisheries. In coral reefs, planktivores are an important trophic guild that includes many reef associated adult and larval fish and the reef building corals. Corals rely on heterotrophy for essential nutrients not acquired through their symbionts for tissue and skeletal growth^{34,35,36}. Increasing heterotrophy is one mechanism for some coral species to compensate for the increased energy demand for calcification under ocean acidification^{37,38}, and yet this option may be diminished if zooplankton abundances are severely reduced. Of note is that we only investigated macrozooplankton abundances, not microzooplankton or microbes in the water column. Thus, corals that feed on smaller organisms or those few coral species that continually feed during the day and not just at night³⁹ may still fare well under reduced abundances of macrozooplankton^{40,41}, which may be the case for the massive bouldering corals that are present at the high-CO₂ sites in high abundances since they appear to

not be negatively affected by the documented reduction in macrozooplankton abundances.

We showed that reduced abundances of demersal zooplankton were in part related to indirect ecological effects of ocean acidification, including changes in their day-time habitat, as branching corals and coral rubble were replaced by massive bouldering corals at high-CO₂. This indirect effect is specific to reef-associated zooplankton and not relevant for oceanic plankton. However, ecological changes (habitat quality and food web structures) due to ocean acidification may also alter demersal zooplankton communities in other coastal marine ecosystems.

In addition to acidification, increased atmospheric CO₂ is warming the oceans⁴², driving some zooplankton species poleward⁴³, enlarging oxygen minimum zones, and restricting vertical migration and distribution of some zooplankton taxa^{44,45}. Stratification is becoming more pronounced, suppressing vertical mixing and prompting up-welled waters to shoal, which through reductions in nutrients and production can also reduce zooplankton by as much as 80%⁴⁶. Our findings outline an additional pathway how zooplankton can be affected by CO₂, suggesting that coral reefs and other coastal ecosystems may be more vulnerable than expected to the rising CO₂ levels if the very basis of their food webs is diminished.

Methods

Zooplankton Sampling and Laboratory analysis

Zooplankton biomass, abundances and community composition were compared between CO₂ regimes (control and high-CO₂ sites), each at two reefs (Dobu and Upa-Upasina; Milne Bay Province, Papua New Guinea), and expeditions (1,2,3). Samples were collected at night (2100-0200 hours local time) and mid-day (1200-1400 hours) for a total of 24 days during three separate expeditions (17 to 27 January 2013, 24 May to 9 June 2013, 29 March to 2 April 2014), using a 100 µm Nansen plankton net (aperture: 70 cm). Horizontal tows were conducted along 30 m transects at both CO₂ sites and reefs, both over the reef (2-3 m depth) and offshore (50-70 m water depth). At the high-CO₂ sites, transects were located along the edge of the seeps but not in the bubble streams to prevent sampling where zooplankton might be disturbed by the bubbles, and to not fill the net with gas bubbles. A hand-held GPS and a HydroBios flowmeter recorded tow distance to determine the volume of water filtered. Three replicate transects were collected at each location. Bulk zooplankton from additional net tows were frozen at -80°C and analyzed for their fatty acid composition using gas chromatography^{47,48}.

To compare diurnal patterns, horizontal tows were additionally conducted over a 24-hour period at the high-CO₂ and control sites of Upa-Upasina during the third expedition once per week for four weeks, with tows every three hours during daylight hours and every 2 hours during the night.

Daytime habitat preference for three dominant substrata (branching coral, coral rubble, and massive bouldering coral) was tested with emergence traps.

The traps consisted of nine custom made pyramid-shaped tents (100 μ m mesh net, $L \times W \times H$: 1 m x 1 m x 0.75 m) with detachable codends that had light (3 lumens) fixed inside to attract zooplankton. Three traps were placed over each of the three types of substrata (>50% branching coral, coral rubble, or bouldering coral) during the third expedition at Upa-Upasina. Over the course of 10 days, the 9 traps were placed in random locations over the different substratum types alternating between the high-CO₂ and the control site. The high-CO₂ site and the control sites were both sampled 5 days each. A photo was first taken of the 1.0 m² quadrat of substratum before the trap was placed over it. Emergence traps were tethered unsealed to the reef substrata with nylon string. Contamination from external zooplankton was expected to be low (a few organisms per trap per night), since demersal zooplankton emerge upward and are unlikely to crawl under a physical barrier, i.e. the trap. Emergence traps were deployed during daylight hours (1300 hrs) before zooplankton emerged into the water column, and the codends were retrieved 3-4 hours after dark (2100-2200 hrs).

From both the horizontal tows and the emergence traps, the contents of the codends were stored in a 4% formaldehyde-seawater solution. Later, replicate subsamples were analyzed in the laboratory. Copepods were identified to family level, and non-copepods were identified to class or order. After identification, samples were split in half with a Folsom splitter and half of the sample was placed onto pre-weighed and pre-combusted GF/F 47 mm filters and Aluminum tins. Samples were dried at 60°C for 24 hours before weighing to obtain biomass data (mg dry weight m⁻³).

Seawater Chemistry

The seawater chemistry at Upa-Upasina and Dobu reefs has been documented previously^{20,25}. The pH at the total scale (pH_T) averaged 8.0 at the control sites and 7.8 at the high-CO₂ sites. The control sites are exposed to a relatively stable pH_T level whereas the high-CO₂ sites experience more variable pH_T levels. Water samples were collected during the expeditions and fixed with mercuric chloride solution and later analyzed for their dissolved inorganic carbon (DIC) and total alkalinity (A_T) using a Versatile Instrument for the Determination of Total Inorganic Carbon and Titration Alkalinity (VINDTA 3C). DIC and A_T were used to calculate other seawater parameters (Supplementary Table 1), including pH at total scale (pH_T), partial pressure of carbon dioxide (pCO₂: μatm), bicarbonate (HCO₃⁻: μmol kg⁻¹), and carbonate (CO₃²⁻: μmol kg⁻¹) using the Excel macro CO2SYS⁴⁹ under the constraints set by Dickson and Millero⁵⁰.

Phytoplankton in the water column

Phytoplankton quantity in the water column were compared between control and high-CO₂ sites at Dobu and Upa-Upasina reefs to determine the amount of food available to herbivorous zooplankton. Water samples were collected at midnight (0000 hr) and midday (1200 hr) using a Niskin bottle. Onboard the M/V Chertan, 3 L of water was immediately filtered through 47 mm GF/F filters and stored in liquid nitrogen. Later in the laboratory, pigments were dark-extracted in 100% acetone and samples were placed in a fluorometer and

measured for the quantity of total oceanic carbon (TOC, $\mu\text{g L}^{-1}$), total nitrogen (TN, $\mu\text{g L}^{-1}$), chlorophyll *a* ($\mu\text{g L}^{-1}$), and phaeophytin ($\mu\text{g L}^{-1}$). Mean TOC, TN, chl *a*, and phaeophytin values are presented in Supplementary Table 2. Generalized linear models (GLMs) were used to determine the statistical significance of environmental factors (CO₂, reef, time, and interaction terms) on the phytoplankton levels (Supplementary Table 3).

Statistical Analysis

Abundance data were averaged across replicate transects (or emergence traps) within CO₂ levels, reefs and nights. Log ratios (high-CO₂/control) of zooplankton abundance (individuals m⁻³) for each zooplankton taxon were estimated with generalized additive mixed models (GAMM) with log link function and quasipoisson distribution using the predictors CO₂ (high-CO₂, control), reef (Upa-Upasina, Dobu), and expedition (1,2, 3; Supplementary Table 4).

Redundancy analysis (RDA) was used to assess the relationship between zooplankton communities and environmental variables (CO₂, reef, and expedition). Zooplankton abundances were 4th-root transformed. Permutation tests were used to determine the statistical significances of the environmental variables between the zooplankton communities.

To determine substratum preference of each zooplankton taxon, the photos were digitally adjusted for tilt and size. The percent coverage was estimated for the targeted substrata (coral rubble, branching coral and bouldering coral), as well as for other co-existing groups including sand, macroalgae, and

turf algae. The influence of the percent coverage of each substratum category, CO₂, reef, and expedition on the abundance of each zooplankton taxon was also evaluated using generalized linear models (GLMs) using a log link function and quasipoisson distribution.

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Author contributions

JNS, KEF, CR, AC designed the experiment. JNS and KEF did the fieldwork. JNS did the laboratory work. GD, JNS, KEF did the statistical analysis. All authors contributed to writing the manuscript.

Competing financial interests

The authors declare no competing financial interests.

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Figure Legends

Figure 1: Differences in zooplankton biomass between control and high-CO₂ sites, derived from horizontal net tows. Zooplankton biomass (a) at the two reefs (Dobu and Upa-Upasina) and three expeditions at night, and (b) a 24-h sampling campaign showing the persistence of nightly vertical migration at both the high-CO₂ and control site of Upa-Upasina reef. Control sites are represented in blue, and high-CO₂ sites are represented in red.

Figure 2. Abundance ratios (high-CO₂/control) for selected zooplankton taxa. The circles and bars represent the means and 95% confidence intervals respectively. The ratios of abundances of zooplankton taxa between the control and the high-CO₂ sites are significantly different at the 5% level if their error bars do not include the value 1.0.

Figure 3. Differences in communities of nocturnal reef-associated zooplankton between control and high-CO₂ conditions at two reefs (Dobu and Upa-Upasina) across three expeditions. The vectors of the redundancy analysis biplots

represent the directions of increased abundance (individuals m⁻³) of the various taxa. Dots represent average values across three net tows per night and CO₂ condition (blue: control, red: high-CO₂).

Figure 4. Influences of CO₂, Reef, Date, and substratum on dominant zooplankton taxa from emergence traps. Substrata are percent cover of: CR = coral rubble, BC = branching coral, MC = massive (bouldering) coral, SA = sand, MA = macro algae, and TA = turf algae. ‘***’ indicates <0.001 significance, ‘**’ indicates <0.05 significance, and empty boxes indicate ‘none significance’. For CO₂ and the substrata, green and purple boxes indicate positive and negative relationships, respectively.