2018-05

Microplastics in sub-surface waters of the Arctic Central Basin

Kanhai, LDK

<http://hdl.handle.net/10026.1/11255>

10.1016/j.marpolbul.2018.03.011

Marine Pollution Bulletin

Elsevier BV

All content in PEARL is protected by copyright law. Author manuscripts are made available in accordance with publisher policies. Please cite only the published version using the details provided on the item record or document. In the absence of an open licence (e.g. Creative Commons), permissions for further reuse of content should be sought from the publisher or author.

Microplastics in sub-surface waters of the Arctic Central Basin

La Daana K. Kanhai^{a, b*}, Katarina Gardfeldt^c, Olga Lyashevskaya^a, Martin Hassellöv^d, Richard C. Thompson^b, Ian O'Connor^a

^aMarine and Freshwater Research Centre, Galway Mayo Institute of Technology, Dublin Road, Galway, Ireland

^bMarine Biology and Ecology Research Centre, School of Biological and Marine Sciences, Plymouth University, Drake Circus, Plymouth, Devon PL4 8AA, United Kingdom

^cDepartment of Chemistry and Chemical Engineering, Chalmers University of Technology, Göteborg, SE-412 96, Sweden

^dDepartment of Chemistry and Molecular Biology, University of Gothenburg, Göteborg, Sweden

Abstract

Polar oceans, though remote in location, are not immune to the accumulation of plastic debris. The present study, investigated for the first time, the abundance, distribution and composition of microplastics in sub-surface waters of the Arctic Central Basin. Microplastic sampling was carried out using the bow water system of icebreaker Oden (single depth: 8.5 m) and CTD rosette sampler (multiple depths: 8 – 4369 m). Potential microplastics were isolated and analyzed using Fourier Transform Infrared Spectroscopy (FT-IR). Bow water sampling revealed that the median microplastic abundance in near surface waters of the Polar Mixed Layer (PML) was 0.7 particles m⁻³. Regarding the vertical distribution of microplastics in the ACB, microplastic abundance (particles m⁻³) in the different water masses was as follows: Polar Mixed Layer (0 - 375) > Deep and bottom waters (0 – 104) > Atlantic water (0 – 95) > Halocline i.e. Atlantic or Pacific (0 – 83).

Keywords

Microplastic, Marine debris, Arctic Ocean, Sub-surface waters, Pollution, Water column

*Corresponding Author

Email Address: ladaana.kanhai@research.gmit.ie

Introduction

The Arctic Ocean, though the smallest in the world, is unique due to its distinct abiotic features and the highly specialised ecosystem it supports. Key anthropogenic drivers which may put pressure on this ecosystem include (i) climate change, (ii) harvest and fisheries, (iii) persistent, bio-accumulative and toxic contaminants, (iv) industrial development, (v) shipping, and (vi) invasive alien species (CAFF 2017). Plastic contaminants in the world's oceans have emerged as an issue of global importance due to their ubiquitous distribution, long-range transport potential, persistence and perhaps most importantly the potential threat they pose to marine organisms (UNEP 2011). Remote polar oceans such as the Arctic Ocean have not been immune to the entry of plastics as a combination of long-range transport processes and local anthropogenic activities have contributed to the plastic debris in these areas.

Characteristic abiotic features which set the Arctic Ocean apart from other oceanic basins include (i) a central area of perennial pack ice, (ii) seasonal extremes in solar irradiance, ice and snow cover, temperature and riverine inflow, and (iii) an upper layer of lower salinity water due to freshwater input from rivers and seasonal sea-ice melt (CAFF 2013). This unique ecosystem is a habitat for a vast array of marine organisms, some of which are (i) endemic to the region, (ii)

commercially important, (iii) apex predators, (iv) central to the functioning of the ecosystem, and (v) threatened as evidenced by their inclusion in the IUCN Red List of Threatened Species (CAFF 2013, CAFF 2017).

Despite its remote location away from major population centres and the low coastal population in its surrounding shelf areas, both macro and microplastics were detected in the various environmental compartments of the Arctic Ocean. Between 2002 and 2014, macroplastics were detected on the seafloor (2500 m depth) of the eastern Fram Strait at the HAUSGARTEN observatory (Bergmann and Klages 2012; Tekman et al. 2017). Sightings of buoyant macroplastics were also made during ship and helicopter observation surveys in the Barents Sea and Fram Strait (Bergmann et al. 2016). A citizen-science study also recently reported the presence of macroplastics on six beaches of the Svalbard Archipelago (Bergmann et al. 2017a). Arctic sea ice was reported by Obbard et al. (2014) as having microplastic concentrations (38 – 234 particles m³ of ice) several orders of magnitude greater than highly contaminated oceanic waters. Lusher et al. (2015) first reported on microplastic abundances in surface and sub-surface waters south and southwest of Svalbard. Amélineau et al. (2016) later reported on microplastic abundance in surface waters east of Greenland. Regarding Arctic species, microplastics have been detected in the gular pouches of Little Aulks (*Alle Alle*), (Amélineau et al. 2016), as well as in the stomachs of juvenile polar cod (*Boreogadus saida*), (Kuhn et al. 2018). Microplastics were also detected in sediments (collection depths 2340 – 5570 m) from the Fram Strait (Bergmann et al. 2017b). Recently, results from a circumpolar expedition of the Arctic indicated that concentrations of floating plastic ranged between 0 – 320 000 items km⁻² in the Greenland and Barents Sea and 0 – 27 000 items km⁻² in the rest of the Arctic Ocean (Cózar et al. 2017).

70

71 Plastic contaminants are introduced to the Arctic Ocean due to a combination of (i) long-range
72 transport processes, e.g. via oceanic currents, biotransport and riverine input, and (ii) local
73 anthropogenic activities, e.g. shipping. The three oceanic currents which supply the greatest water
74 volumes to the Arctic Ocean are the (i) West Spitsbergen Current i.e. the polar limb of the North
75 Atlantic circulation which carries warm water from the North Atlantic Current (9.5 Sverdrup, Sv
76 $= 10^6 \text{m}^3 \text{s}^{-1}$), (ii) a cold ocean current that enters from the Pacific Ocean via the Bering Strait (1.5
77 Sv) and, (iii) a branch of the North Atlantic Current, which flows along the Siberian coastline (1.0
78 Sv), (Zarfl and Matthies 2010). These oceanic currents may also transport plastics to the Arctic
79 Ocean with the estimated plastic flux to this region ranging between 62 000 to 105 000 tons per
80 year (Zarfl and Matthies 2010). Models based on a particle-trajectory approach for studying the
81 fate of marine debris in the open ocean highlighted the northward transport of marine debris to
82 polar regions and the formation of a sixth so-called garbage patch in the Barents Sea (van Sebille
83 et al. 2012). Bio-transport is another long-range transport process via which plastics may enter
84 polar regions. Plastic ingestion was reported in Northern Fulmars (*Fulmaris glacialis*) and Thick-
85 billed Murres (*Uria lomvia*) in the Arctic (Mallory 2008; Provencher et al. 2012; Trevail et al.
86 2015). Some studies suggested that the seabirds had ingested plastics during their wintering in the
87 North Atlantic Ocean and had then transported the contaminants to the Arctic upon migration
88 (Mallory 2008; Provencher et al. 2012). Riverine discharge from Siberian (Ob, Yenisei and Lena)
89 and Canadian (Mackenzie) rivers are other potential sources of plastics to the Arctic. Obbard et al.
90 (2014), however, point out that the contribution of riverine discharge to plastic input in the Arctic
91 is projected to be low due to the fact that these rivers flow through sparsely populated watersheds.
92 Local anthropogenic activities are another source of plastics to the Arctic. Increased ship traffic

93 due to shipping and tourism was found to be positively correlated with increased litter densities in
94 the Fram Strait (Bergmann and Klages 2012; Tekman et al. 2017).

95
96 The intense focus by scientists on the near-surface layer of the ocean for microplastics has been
97 due in part to the presumption that the majority of particles would be found in this region of the
98 water column given the inherent densities of individual synthetic polymers. Such a theorization
99 led to traditional techniques that involved nets, manta trawls as well as the seawater intake of
100 vessels that sampled only the upper few metres of the water column for microplastics. Yet, several
101 studies indicated that a mismatch existed between observed and expected plastic concentrations in
102 surface oceanic waters when estimated plastic production and projected inputs to the oceans were
103 considered (Cózar et al. 2014; Eriksen et al. 2014). It was therefore proposed that several
104 mechanisms potentially influenced the vertical distribution of microplastics within the water
105 column and led to their transport out of surface waters. Some of these mechanisms included (i)
106 incorporation into marine aggregates (Long et al. 2015), (ii) biofouling (Fazey and Ryan 2016),
107 (iii) incorporation into faecal matter (Cole et al. 2016) and, (iv) hydrodynamic factors such as wind
108 (Kukulka et al. 2012). Despite the theorization that surface waters are not the ultimate repository
109 for plastic debris in the marine environment (Cózar et al. 2014), few studies ventured beyond
110 traditional near-surface microplastic monitoring to investigate their vertical distribution in the
111 water column.

112
113 Microplastic pollution in the Arctic Ocean is an issue that warrants attention due to the potential
114 threats that these contaminants may pose to the inhabitants of this unique ecosystem. A practical
115 step towards addressing this issue and evaluating the extent of the problem involves assessing the

abundance, distribution and composition of microplastics in Arctic waters. Whilst microplastic monitoring in the marine environment has traditionally focused on surface waters, the reality is that the vast majority of marine organisms inhabit sub-surface waters. Monitoring microplastics in sub-surface waters is particularly relevant as it can also provide some insight into the whereabouts of the ‘missing plastic’ from surface waters. To our knowledge, the present study sought for the first time (i) to provide a spatial overview of microplastic abundance, distribution and composition in the Polar Mixed Layer (PML) of the Arctic Central Basin (ACB) and, (ii) to determine whether microplastics in the ACB were being transported out of surface waters by assessing their vertical distribution in the water column.

2.0 Materials and Method

2.1 Study Area

The Arctic Ocean is comprised of a deep central basin surrounded by extensive continental shelves (CAFF 2013). The bathymetry of the Arctic Ocean is such that the Lomonosov Ridge separates the central basin into the Canadian (Amerasian) and Eurasian basins with the basins being further sub-divided by the (i) Gakkel Ridge, into the Amudsen and Nansen basins and, (ii) Alpha Ridge, into the Makarov and Canada basins (Jakobssen et al. 2004; Rudels 2015, Figure 1). A major structuring element of the Arctic marine ecosystem is sea ice which floats on the surface layer impeding surface mixing and influencing freshwater and heat fluxes (CAFF 2013). In the Arctic Ocean, there is a distinct vertical stratification of the water column giving rise to three major water layers (i) Polar Surface Water (PSW) which includes the Polar Mixed Layer (PML) and the halocline, (ii) Atlantic Water and, (iii) deep and bottom waters (Rudels 2015, Figure 1). The PML (approximately 50 m deep) is the uppermost surface layer of low salinity water formed as a result

of sea ice melt and the influx of freshwater from riverine sources (Rudels et al. 1991; CAFF 2013; Jakobsson et al. 2014). Beneath the PML is a halocline (50 – 250 m), characterised by a strong salinity increase with depth and comprised of either Pacific waters or Atlantic waters with the Pacific halocline being deeper than the Atlantic halocline (Rudels et al. 1991; Jakobsson et al. 2004). Below the halocline lies an intermediate water layer comprising of dense saline Atlantic water. The deep and bottom waters also referred to as Arctic deep water ranges from a depth of approximately 900 m and extend to the seafloor (Rudels et al. 1991; CAFF 2013).

2.2 Sample Collection

Underway samples

This study was conducted onboard the Swedish icebreaker Oden during the Arctic Ocean 2016 expedition. The vessel departed Longyearbyen, Svalbard on August 8th 2016 and traversed approximately 4943 nautical miles in the Arctic Ocean until its return on September 19th 2016 (Figure 2). Sub-surface oceanic water pumped onboard the vessel via the bow water system was sampled for microplastics according to Lusher et al. (2014). Sampling was conducted for a period of approximately 6 weeks (9 August to 16 September 2016). Since each sample constituted the filtration of approximately 2000 L of water, the total survey effort for this study was approximately 116 000 L of water (58 samples).

Seawater from a continuous intake located at the keel of the ship (depth 8.5 m) was pumped onboard the vessel using a rotary positive displacement pump (Universal II Series Pump, Waukesha Cherry-Burrell) at a flow rate of 85 L/min (at optimal capacity) and transported to the laboratory via stainless steel pipes. Prior to reaching the laboratory, the seawater passed through a

stainless steel primary filter (pore size 2.5 mm) which was beyond the control of the investigator. The discharge of grey water in relation to the seawater intake was not an issue since the seawater intake was located towards the front of the vessel whilst grey water was discharged mid-vessel. In the laboratory, seawater from the vessel's bow water system was allowed to flow through a covered stainless steel sieve (250 μ m) by means of a connection hose fitted into the wooden sieve cover. For the duration of the sampling, the stainless steel sieve was supported in a wooden stand. Based on Lusher et al. (2014), approximately 2000 L of water was filtered for each sample. The length of time taken for the filtration of the specified volume of water was determined by calculation of the flow rate of the seawater. A flow meter, attached at a point prior to the entry of the water into the sieve, was also used to verify the volume of water filtered. Once the specified volume of water was filtered, the sieve was removed and Milli-Q water was used to wash retained material from the sieve into a clean container. The collected material was then filtered under vacuum onto glass microfiber paper (GF/C); Whatman: 47 mm, pore size: 1.2 μ m, using a Buchner funnel and a vacuum flask (Lusher et al. 2014). Each filter paper was then placed into a clean plastic petri dish, covered and stored in a freezer (-20 °C) until returned to the laboratory. At the start and at the end of each sample, positioning data were collected. Data for various environmental variables were obtained from the vessel's (i) thermosalinometer (water temperature, salinity) and (ii) weather station (wind speed and direction).

CTD samples

A rosette water sampler containing 24 Niskin bottles coupled to a Sea-Bird SBE 911 conductivity-temperature-depth (CTD) sensor suite (hereafter referred to as CTD) was used to collect subsurface water samples and hydrographic data at 9 sampling locations in the Arctic Ocean. Upon

deployment from the vessel, the CTD entered the water and was allowed to descend to the bottom layer. During the descent, Niskin bottles were open with water flowing through them. It was during the up-cast that multiple Niskin bottles were closed at specific depths in order to facilitate the collection of a specified volume of water. A total of 48 water samples were retrieved during the 9 CTD casts to sample for microplastics. At 7 of the CTD casts, 6 water depths were sampled with 48 L of water collected at each depth i.e. 4 Niskin bottles (12 L) per depth. At 2 CTD casts, 3 water depths were sampled with 21 L of water collected at each depth i.e. 3 Niskin bottles (7 L) per depth. At a particular sampling location, the overall goal was to collect samples in the near-surface, mid-water and bottom layers within the water column thereby reflecting the main water masses. As such, exact sampling depths were determined by the information provided by the salinity and temperature sensors on each downcast. The deepest samples at each CTD cast were collected at least 10 m above the seafloor at a given location. Following each CTD cast, Niskin bottle taps were rinsed with Milli-Q water and a clean hose was attached. Water from bottles closed at the same depth was passed through the same stainless steel sieve (250 μm) held in a covered wooden stand. Once water from all bottles at a specific depth had been filtered, the sieve was removed and Milli-Q water was used to wash retained material from the sieve into a clean container. The collected material was filtered under vacuum onto glass microfiber paper (GF/C); Whatman: 47 mm, pore size: 1.2 μm , using a Buchner funnel and a vacuum flask. Each filter paper was then folded and placed into an aluminium foil packet and stored in a freezer (-20 °C) until returned to the laboratory.

2.3 Method Validation and Contamination Prevention

For the underway samples, potential contamination during sample processing was evaluated by (i) leaving clean plastic petri dishes with filter paper exposed to the air during vacuum filtration, and (ii) passing an aliquot (250 mL) of Milli-Q water through clean GF/C filter paper under vacuum. For the CTD samples, potential contamination was assessed by filling a clean Niskin bottle with Milli-Q water and subjecting it to the exact process a sample underwent. Measures taken to prevent contamination in the laboratory included (i) wearing lab coats (cotton/polyester blend), cotton clothing and gloves (nitrile) during sample processing, (ii) placing a wooden cover over the stainless steel sieve during filtration to prevent airborne contamination, and (iii) washing all containers used during sample processing with Milli-Q water before reuse.

2.4 Laboratory analyses

Filter papers were removed from the freezer, left to dry and then visually examined under a dissecting microscope (Olympus SZX10) equipped with a polariser and camera (Q Imaging Retiga 2000R). Potential microplastics were isolated and processed (photographed and length measurements taken) prior to transferring to a clean filter paper in a labelled petri dish (Kanhai et al. 2017). All potential microplastics were analysed by Fourier transform infrared (FT-IR) spectroscopy on a Thermo Scientific Nicolet iN10 FT-IR spectrometer. The instrument was equipped with a potassium bromide (KBr) beamsplitter and an internal mercury cadmium telluride (MCT) detector which was cooled with liquid nitrogen. Microscope-reflectance sampling was performed and spectra were recorded as the average of 256 scans in the spectral wave number range of 4000 - 675 cm^{-1} at a resolution of 4 cm^{-1} . Thermo Scientific's OMNIC Picta Version 9 spectroscopy software was used for processing and evaluating all spectra. Prior to analysing each sample, background scans were performed and sample spectra were automatically corrected. Each

sample spectrum was compared with those of known standard polymers in the (i) Hummel Polymer Sample library, (ii) Polymer Laminate Films library, and (iii) Wizard library. Values of between 0 and 100 % were produced for each match between sample and reference spectra with the highest percentage representing the closest match. Particles for which there was uncertainty regarding the identity of the polymer (specifically fragments and some fibres) were subjected to further FT-IR spectroscopy on a Bruker Vertex 70 Infrared Spectrometer coupled to a Hyperion 1000 microscope (Kanhai et al. 2017). Samples which produced spectra with a match < 60 % were automatically rejected while those with a match of > 70 % were automatically accepted. All spectra with matches > 60 % were individually examined to ensure that there was clear evidence of peaks from the sample corresponding to known peaks of standard polymers and that instances of the misidentification of natural and semi-synthetic polymers was reduced (Comneau-Stancu et al. 2017).

2.5. Statistical analyses

All statistical analyses were performed using R version 3.2.3 (R Core Team 2015). Descriptive statistics, histograms and box plots were generated and tests of normality (Supplementary Tables 1 and 2) were conducted on all data to determine whether parametric or non-parametric statistical analyses were appropriate. Correlation analyses were performed between individual environmental variables and microplastic abundance for both underway and CTD samples. A generalized additive model (GAM) was developed using the underway data and a generalized linear mixed model (GLMM) was developed using the CTD data to determine which environmental variables had an effect on microplastic abundance.

3. Results

3.1. Quality Control

In conjunction with the collection of samples via the underway system of the vessel, a total of 24 blanks (air contamination-12, method-12) were run (Supplementary Table 3). No synthetic polymers were found in the method blanks. However, a single synthetic fibre (blue, polyethylene terephthalate, 0.438 mm) was found in the last air contamination blank. For 6 of the 9 CTD casts, at least one method blank was run (Supplementary Table 4). Between 0 and 3 synthetic fibres were found in each of the method blanks. The synthetic fibres that were found included polyethylene terephthalate (n = 8), polyacrylonitrile (n = 1) and polyvinyl chloride (n = 1).

3.2. Overview of findings

Underway samples

A total of 303 particles were isolated from the underway samples and analysed by FT-IR spectroscopy. Of these, 46 particles were excluded because of uncertainty regarding their identity (< 60 % match to reference spectra) and in the minority of cases (n = 6) due to their length (< 250 µm). Of the remaining particles (n = 257), 14 were macro-particles (i.e. > 5 mm in length) and 243 were micro-particles (< 5 mm in length). Of the macro-particles, 11 were confirmed as macroplastics having the following polymer types: polyethylene terephthalate (4), polyamide blend (4) and polyacrylonitrile (3). Of the 243 micro-particles, 110 were natural (cellulosic), 16 were semi-synthetic (cellulose-based e.g. rayon) and 117 were synthetic. All further analyses and discussions focus on the 117 confirmed microplastics.

The majority (94 %) of microplastics were fibres and 6 % were fragments. In terms of colour, the most prevalent were blue (49 %) and transparent (25 %) (Figure 3a). Approximately 62 % of the microplastics occurred in the larger size classes of 1.0 – 2.0 mm and 2.0 – 5.0 mm (Figure 4a). Microplastic polymer types included polyester (n = 88), blends (n = 11), polyacrylonitrile (n = 8), polyamide (n = 5) and polyvinyl chloride (n = 5), (Figure 5a). The overall category of ‘polyester’ included both polyethylene terephthalate (PET) and other polyesters while blends included either polyamide blends or polyester blends.

CTD samples

A total of 157 particles were isolated from the CTD samples and analysed by FT-IR spectroscopy. Of these, 14 were excluded for the reasons mentioned above for the underway samples. Of the remaining particles, 2 were categorised as macroplastics (> 5 mm) and included PET and polyacrylonitrile. Of the 141 micro-particles (< 5 mm), 39 were natural (cellulosic), 8 were semi-synthetic (cellulose-based e.g. rayon) and 94 were synthetic. All further analyses and discussions focus on the 94 confirmed microplastics.

Overall, the characteristics of the microplastics isolated from CTD samples were similar to those from the underway samples in that (i) the majority (96 %) of microplastics were fibres and 4 % were fragments, (ii) the most prevalent colours were blue (46 %) and transparent (22 %) (Figure 3b), (iii) the majority (64 %) of microplastics were in the larger size classes of 1.0 – 2.0 mm and 2.0 – 5.0 mm (Figure 4b), and (iv) microplastic polymer types included polyester (n = 74), blends (n = 12), polyacrylonitrile (n = 6), polyamide (n = 1) and polyvinyl chloride (n=1), (Figure 5b).

The overall category of polyester included both polyethylene terephthalate (PET) and polyester while blends included only polyamide blends.

3.2 Microplastic abundance and distribution in the Arctic Ocean

Based on the underway samples (collection depth 8.5 m), microplastic abundance in sub-surface waters in the Arctic Ocean ranged between 0 – 7.5 particles m^{-3} with a median of 0.7 particles m^{-3} (interquartile range 0.4 – 1.0), (Figure 2, Supplementary Table 5). For the majority of the sampling sites, microplastic abundance ranged between 0 – 1.0 particles m^{-3} . However, at a few sites, microplastic abundances were between 2 – 2.5 particles m^{-3} and at two sites it was at 5 and 7.5 particles m^{-3} respectively. Based on the CTD samples (collection depths between 8 – 4369 m), microplastic abundance in sub-surface waters in the Arctic Ocean ranged between 0 – 375 particles m^{-3} with a median of 20.8 particles m^{-3} (interquartile range 20.8 – 62.5) (Figures 2 and 5, Supplementary Table 6). With the exception of CTD cast 4, the CTD casts (1 – 3) nearer the periphery of the Arctic Central Basin (ACB), i.e. in the Nansen Basin (Yermak Plateau), reflected a comparatively higher abundance of microplastics in the water column than other CTD casts within the ACB (Figures 2, 6). It must be noted however that CTD casts 1 – 3 sampled the upper 850 m of the water column and as such would have sampled particles from the Polar Mixed Layer (PML), Atlantic halocline and Atlantic water (Figures 1, 6). This is in contrast to the other CTD casts which sampled a much more extensive vertical range throughout the water column by including deep bottom water in excess of 1000 m depth. Overall, there was no statistically significant correlation between microplastic abundance and depth (Spearman's rank correlation, $\rho = 0.06$, $p\text{-value} = 0.7$). However, upon examination of individual CTD casts, it is apparent that microplastic abundance was not uniform at various depths in the water column and that there were

certain depths that reflected higher microplastic abundances (Figure 6). Additionally, microplastic abundance (particles m^{-3}) in the different water masses of the ACB was as follows: Polar Mixed Layer (0 - 375) > Deep and bottom waters (0 – 104) > Atlantic water (0 – 95) > Halocline i.e. Atlantic or Pacific (0 – 83), (Table 1).

3.3. Association between environmental variables and microplastic abundance in samples

Correlation analyses were conducted to determine whether there was any association between environmental variables and microplastic abundance in the samples. For both the underway and CTD samples, there was no statistically significant correlation between microplastic abundance and any of the ancillary environmental variables of temperature, salinity, wind direction, wind speed, depth and density (Supplementary Tables 1 and 2). Specifically, there was no statistically significant correlation between microplastic abundance at depth (Supplementary Table 2). However, for the underway samples, there was a statistically significant weak negative correlation between microplastic abundance and latitude (Spearman's rank correlation, $\rho = -0.286$, $p\text{-value} = 0.03$).

Using the underway data, a Generalized Additive Model (GAM) was developed to further determine whether environmental variables influenced microplastic count in the underway samples. In this model, the response variable was microplastic count (number of microplastics per sample) and initial explanatory variables included location (latitude, longitude), physico-chemical properties associated with sub-surface waters (temperature, salinity) and weather data (wind direction, wind speed). In the model, the Poisson family distribution of error terms was specified with a log link function since microplastic abundance data were count data. The output of the

initial model was examined and based on this non-parametric smoothers were accordingly applied to the explanatory variables. A scale invariant tensor product smooth (te) was applied to latitude and longitude while a cubic regression spline (bs = cr) was applied to all other variables except wind direction to which no smoother was applied (based on initial model plots). Non-significant explanatory variables (as evidenced by their p-values) were eliminated in a stepwise manner until a GAM with the lowest Akaike Information Criterion (AIC) score and the fewest explanatory variables was obtained. The final GAM (R-sq = 0.396) was as shown below:

Microplastic count ~ te(**latitude, longitude**) + s(**temperature**, br = "cr") + s(**wind speed**, bs = "cr")

All of the explanatory variables that were present in the final model (shown in bold) were found to have a significant influence on microplastic count in water samples from the Arctic Ocean (wind speed, p-value = 0.0006, latitude, longitude, p-value = 0.0007, temperature, p-value = 0.0483).

A generalized linear mixed model (GLMM) was developed using the CTD data to determine the influence of environmental variables on microplastic count in the CTD samples. In this model, the response variable was microplastic count (number of microplastics per sample) and initial explanatory variables included location (latitude, longitude), physicochemical properties associated with sub-surface waters (temperature, salinity), depth at which water was sampled and ctd number. All the explanatory variables were included in the model as fixed effects, with the exception of ctd number which was included as a random effect. In the model, the Poisson family distribution of error terms was specified with a log link function since microplastic abundance data

were count data. Based on the preliminary finding that there was a statistically significant correlation between depth and salinity (Spearman's rank correlation, $\rho = 0.852$, $p\text{-value} = 8.156 \times 10^{-13}$), temperature and salinity (Spearman's rank correlation, $\rho = 0.506$, $p\text{-value} = 0.00074$) and depth and density (Spearman's rank correlation, $\rho = 0.973$, $p\text{-value} = < 2.2 \times 10^{-16}$), interactions between these variables (denoted by ':') were included in the initial model. Non-significant explanatory variables (as evidenced by their p -values) were eliminated in a stepwise manner until a model with the lowest Akaike Information Criterion (AIC) score and the fewest explanatory variables was obtained. The significance of the random effect (ctd number) in the final model was verified by using analysis of variance (ANOVA) to compare the full final model to a reduced model (random effect deleted). The model with the random effect was shown to be significantly different (ANOVA, $p\text{-value} = 0.0008$, $\Delta\text{AIC} = 9.19$) from the model without the random effect. The model with the lower AIC score ($\text{AIC} = 190.59$) was retained as the final mixed effects model as shown below:

Microplastic count ~ **latitude** + **temperature:salinity** + (1|ctd)

Latitude ($p\text{-value} = 0.0198$) and the physicochemical parameters of temperature and salinity ($p\text{-value} = 7.46 \times 10^{-5}$), as shown in bold, were the explanatory variables that were found to have a significant influence on microplastic count in the CTD samples.

4.0 Discussion

The discovery of microplastics in virtually every environmental phase (sea ice, water, sediments, biota) of the Arctic and Southern Oceans has revealed that polar oceans, though remote, are not

immune to the entry of plastic contaminants to their ecosystems (Bergmann and Klages 2012; Obbard et al. 2014; Lusher et al. 2015; Amélineau et al. 2016; Bergmann et al. 2016; Bergmann et al. 2017a; Bergmann et al. 2017b; Cincinelli et al. 2017; Cózar et al. 2017; Isobe et al. 2017; Tekman et al. 2017; Waller et al. 2017). The present study expands the knowledge base about plastics in the Arctic by providing evidence for the existence of microplastics in the Polar Mixed Layer (PML) as well as some insight into the vertical distribution of microplastics in the Arctic Central Basin (ACB). This region of the Arctic, though of low productivity, has been recognised as an Ecologically/Biologically Significant Marine Area (EBSA) due to its uniqueness/rarity, provision of a critical habitat and ability to support specialised biota (CAFF 2017). There is cause for concern about microplastics in Arctic waters since laboratory studies have shown that these contaminants may (i) hinder algal photosynthesis/growth (Bhattacharya et al. 2010; Besseling et al. 2014), (ii) reduce feeding and energy reserves of lugworms (Besseling et al. 2013; Wright et al. 2013), (iii) reduce filtering activity and decrease lysosomal membrane stability in mussels (Von Moos et al. 2012; Wegner et al. 2012), (iv) reduce feeding and reproductive output in copepods (Cole et al. 2015) and, (v) cause liver stress, negatively impact upon cholinergic neurotransmission and lead to endocrine disruption in fish (Oliveira et al. 2013; Rochman et al. 2013; Rochman et al. 2014). It must be pointed out, however, that some laboratory experiments which reported negative effects of microplastics on marine organisms used microplastic concentrations of 42 to 10 000 particles/mL or 42 million to 10 billion particles m^{-3} (Phuong et al. 2016). In context, microplastic abundance in the ACB as reported by the present study ranged from 0 – 7.5 particles m^{-3} (based on underway sampling) and 0 – 375 particles m^{-3} (based on CTD sampling). Although the ecological impact of microplastics upon the Arctic ecosystem presently remains unknown, it is plausible that these contaminants could pose a threat to its inhabitants.

412

413 The present study showed for the first time the pervasiveness of microplastics throughout the water
414 column of the Arctic Central Basin. Between depths of 8 – 4400 m, microplastic abundance in the
415 ACB ranged between 0 – 375 particles m⁻³ (based on CTD sampling). Such findings provide
416 evidence that in natural conditions microplastics are being vertically transported out of surface
417 waters. These findings also give some indication as to the whereabouts of the ‘missing plastic’
418 from oceanic surface waters (Cózar et al. 2014; Eriksen et al. 2014). Recently, Courtene-Jones et
419 al. (2017) also reported on microplastic abundance (70.8 particles m⁻³) in deep oceanic waters
420 (2227 m at the Rockall Trough, North East Atlantic Ocean) and similarly suggested the possibility
421 of vertical re-distribution of microplastics within the water column. Although it remains unclear
422 as to which mechanisms are specifically operating in the ACB to influence the vertical transport
423 of particles, previous studies have provided several possibilities. Specifically, some laboratory
424 experiments showed that aggregates of algae species (*Chaetoceros neogracile*, *Rhodomonas*
425 *salina*) were capable of incorporating and concentrating microplastics and that the microplastics
426 impacted the sinking rates of the aggregates (Long et al. 2015). In the Arctic Ocean, it is certainly
427 plausible that marine aggregates may be playing a role in the vertical transport of microplastics
428 due to the existence of phytoplankton in the ACB (CAFF 2017) and the fact that transparent
429 exopolymer particles (TEPs); which are excreted by algae and are important components of marine
430 aggregates, have been reported in sub-surface waters as far north as the Fram Strait (Engel et al.
431 2017). Biofouling is another possibility as field studies have shown that plastic particles exposed
432 to natural conditions became sufficiently fouled, had their average material density affected
433 leading them to sink (Fazey and Ryan 2016). The incorporation of microplastics into faecal matter
434 is another means by which microplastics may be vertically transported out of surface waters given

that laboratory experiments have shown that zooplankton may egest microplastics within densely packed faecal pellets which in natural conditions would sink or in some cases be eaten by other biota (Cole et al. 2016).

Sea ice is an integral component of the Arctic Ocean's ecosystem and as such possibly exerts an influence on microplastic abundance in sub-surface waters. Sea ice floating on the surface of the water column in the Arctic Ocean can potentially act as (i) a source of microplastics upon melting, (ii) a physical barrier to wind and as such reduce vertical mixing of surface waters and, (iii) a physical barrier to influx of polluted surface waters. Based on the analysis of sub-sections of four ice cores, sea ice in the Arctic Ocean was reported to contain orders of magnitude more microplastic than contaminated oceanic waters suggesting that sea ice potentially acts as both a sink and a source of microplastics (Obbard et al. 2014). Apart from Obbard et al. (2014) no data exists in the published literature regarding either the spatial or vertical distribution of microplastics in sea ice from the Arctic Ocean. In the upper water column, the absence of sea ice cover means that wind stress can generate turbulence and lead to vertical mixing of buoyant plastic debris (Kukulka et al. 2012). More recently, Cózar et al. (2017) suggested that sea ice can also act as a physical barrier preventing the surface advance of polluted Atlantic water into the Arctic Ocean. The present study highlighted that the Polar Mixed Layer (PML) of the ACB reflected the highest overall microplastic abundance (particles m^{-3}): Polar Mixed Layer (0 - 375) > Deep and bottom waters (0 – 104) > Atlantic water (0 – 95) > Halocline i.e. Atlantic or Pacific (0 – 83). As previously mentioned, the PML is the uppermost surface layer of low salinity water (approximately 50 m deep) formed as a result of sea ice melt and the influx of freshwater from riverine sources (Rudels et al. 1991; CAFF 2013). It is possible that one of the reasons that the highest microplastic

abundances were recorded in this layer is due to its proximity to microplastic sources such as melting sea ice as well as sea-going vessels (especially if they are releasing wastewater to the environment). Furthermore, in the present study, the highest microplastic abundances were reported nearer to the periphery of the Arctic Central Basin (ACB), i.e. in waters north of Svalbard. It is possible that the lack of permanent sea ice cover in this region of the Arctic allows incoming Atlantic water to have a greater influence on near surface waters thereby resulting in higher microplastic abundances. A recent circumpolar expedition of the Arctic Ocean similarly reported that the north eastern Atlantic sector of the Arctic was a hotspot of plastic debris due to the influence of incoming Atlantic water (Cózar et al. 2017).

The present study showed that there was a predominance of fibrous microplastics (> 90%) in sub-surface waters of the ACB. This dominance of fibres in sub-surface waters was similarly reported in (i) the north east Pacific Ocean (75 %), (ii) the north east Atlantic Ocean (96 %), (iii) south/southwest of Svalbard (95 %), and (iv) the Atlantic Ocean (96 %) (Desforges et al. 2014; Lusher et al. 2014; Lusher et al. 2015; Kanhai et al. 2017). Fibrous microplastics in the marine environment most likely originate from textile materials and fishing gear (Andrady 2017). Studies have indicated that washing clothes may lead to the release of fibrous materials in the order of > 1900 fibres per wash or as much as 700 000 fibres per 6 kg load of acrylic fabric (Browne et al. 2011; Napper and Thompson 2016). A recent study in the Ross Sea revealed that the highest concentration of fibrous microplastics (54 %) was found close to the effluent of a sewage treatment plant at the scientific Mario Zucchelli Station, Antarctica (Cincinelli et al. 2017). Fibrous microplastics may enter the Arctic Ocean through a combination of long range transport processes (e.g. via oceanic currents, riverine input) or more in-situ activities such as the release of wastewater

from vessels operating in the region. Another mechanism which was recently suggested as being responsible for plastic fibres in Arctic sea ice was atmospheric transport (Cózar et al. 2017). This is certainly plausible as there have been reports of the atmospheric fallout of synthetic polymers in both urban and sub-urban environments in France (Dris et al. 2016). It must be highlighted that while fibrous microplastics seem to be dominant in certain sub-surface waters, other studies conducted in surface Arctic waters reported a predominance of filaments (97 %) and fragments (73 %), (Amélineau et al. 2016; Cózar et al. 2017).

Analytical techniques such as FT-IR and Raman spectroscopy are a central component of microplastic studies which aim to provide unambiguous identification of synthetic polymers in environmental samples. Omission of this critical step is likely to be a key contributor to an overestimation of microplastic abundances due to the inclusion of non-synthetic polymers in microplastic counts. In the present study, for example, only a percentage (underway - 48 %, CTD - 67%) of the particles were confirmed as synthetic polymers with the remainder being a combination of natural and semi-synthetic polymers. Of the synthetic polymers in the present study, the most abundant (underway – 74 %, CTD – 78 %) was polyethylene terephthalate (PET). A member of the polyester family, PET is one of the five major types of commodity plastics commonly found in the marine environment (Andrady 2011; Andrady 2017). As a thermoplastic, PET is often used in manufacturing beverage containers and packaging materials and its fibres are used in clothing. Overall, this finding of a high prevalence of polyesters in sub-surface waters of the Arctic Central Basin was also corroborated by those of other Arctic studies which reported that polyester was the most prevalent synthetic polymer in waters south/southwest of Svalbard (15 %), in waters of the east Greenland Sea (53 %), and in sea ice (21 %), (Obbard et al. 2014; Lusher et

al. 2015; Amélineau et al. 2016). Other polymers in sub-surface Arctic waters in this study included polyacrylonitrile, polyamide and polyvinyl chloride. Of note is the fact that the majority of synthetic polymers in the present study had densities greater than that of seawater (Andrady 2017). Even though investigators are able to generate information regarding the identity of polymers in environmental samples, definitive statements cannot be made about the origin of the plastics. Based on the identity and type of synthetic polymers found in sub-surface Arctic waters, it is likely that they originated from textiles, fishing gear, beverage containers and packaging materials (Andrady 2011; Andrady 2017).

Within the water column, the distribution of microplastics is in a state of flux due to the influence of multiple factors. The development of models based on simultaneously acquired environmental and microplastic data is immensely useful in this respect as they can provide some discernment regarding the variables influencing measures of microplastic abundances in the marine environment. In the present study, the utilisation of a generalized additive model (GAM) and a generalized linear mixed effects model (GLMM) was particularly relevant due to the ability of both models to handle non-normal data and in the case of the GLMM to differentiate between fixed and random effects. Visual inspection of microplastic abundances in sub-surface waters (Figure 2) revealed that the highest microplastic abundances were located to some extent towards the periphery of the Arctic Central Basin (ACB). Bearing this in mind, it was presumed that the ‘location’ of sampling could have influenced the number of microplastics that were found in the samples. Cózar et al. (2017) also noted this spatial heterogeneity in the distribution of plastics in the Arctic. For these reasons, latitude and longitude were included as a proxy of location in the models. In the GAM, both variables were included using a smoother and therefore it was not

assumed that latitude and longitude had a linear effect on microplastic count. Furthermore, during the period of sampling (August/September 2016), the Arctic Central Basin (ACB) was not completely covered by sea ice. Since the vessel traversed areas of open water, wind was included in the model as it could have influenced microplastic abundance at specific sampling sites. Both models suggested that location, oceanographic (temperature, salinity) and atmospheric variables (wind) had a significant influence on microplastic counts in samples of water from the Arctic Ocean. The findings of the present study must be taken in the context of the number of samples used in the generation of the GAM ($n = 58$). Models that are based on a low number of samples are weak e.g. standard errors are inflated, etc. However, such models allow us to delve a bit deeper into the factors influencing measurements of microplastic abundance in the Arctic Ocean. Findings of the present study were also corroborated by previous studies which indicated that water temperature, salinity and wind also had a significant effect on microplastic abundance (Lusher et al. 2014; Lusher et al. 2015; Kanhai et al. 2017).

Comparative assessments between oceanic basins are critical in providing an indication of the extent of microplastic pollution in the marine environment. A major challenge, which demands caution when drawing conclusions from such comparisons, is the lack of standardization of microplastic sampling methods (depth of collection, mesh size of net/sieve, etc). Microplastic abundances in the present study were not normally distributed and therefore the median was reported as it is the most relevant measure of central tendency for such data. However, in order to enable comparability with other published studies, which generally did not report median microplastic abundances nor made statements about the normality of their data, the mean was also reported in Supplementary Table 7. In the present study, sub-surface waters (depth 8.5 m) in the

Arctic Central Basin (ACB), sampled via the bow water system, had a mean microplastic abundance of 0.97 ± 1.20 particles m^{-3} . In comparison to other studies that employed similar methods (i.e. the underway system of vessels) to sample sub-surface waters, microplastic abundance in the ACB was lower than values reported (i) in the north eastern Pacific Ocean (279 ± 178 particles m^{-3}), (ii) in the North Atlantic Ocean ($13 - 501$ particles m^{-3}), (iii) off Svalbard (2.68 ± 2.95 particles m^{-3}), (iv) in the north east Atlantic Ocean (2.46 ± 2.43 particles m^{-3}), and (v) in the Atlantic Ocean (1.15 ± 1.45 particles m^{-3}), (Desforges et al. 2014; Lusher et al. 2014; Enders et al. 2015; Lusher et al. 2015; Kanhai et al. 2017, Supplementary Table 7). The only oceanic basin for which there were reports of lower microplastic abundances in sub-surface waters was the Ross Sea (0.17 ± 0.34 particles m^{-3}), (Cincinelli et al. 2017; Supplementary Table 7). Although the methods used to sample sub-surface waters for microplastics in the above mentioned studies were similar, the fact remains that the variation of several factors e.g. mesh size of sieve ($1 - 300 \mu\text{m}$), sampling depth ($3 - 11$ m), etc., amongst the studies could have impacted the reported microplastic abundances. In terms of assessing the vertical distribution of microplastics in the marine environment, Bagaev et al. (2017) was the only other published study which utilised a similar sampling method (Niskin bottles) in the Baltic Sea. Being cognisant of the fact that no confirmatory analytical techniques or blanks were used by Bagaev et al. (2017), microplastic abundance at multiple depths in sub-surface waters of the Arctic Ocean (mean: 46 ± 62 particles m^{-3} ; range: $0 - 375$ particles m^{-3} ; depths sampled: $8 - 4400$ m) was lower than reported for the Baltic Sea (mean: 310 ± 520 particles m^{-3} ; $70 - 2600$ particles m^{-3} ; depths sampled: $1 - 218$ m). Similar to the findings of the present study whereby the highest microplastic abundances were found in the uppermost water layer i.e. the PML, Bagaev et al. (2017) reported that near- surface and near-bottom water layers in the Baltic Sea had higher fibre concentrations than intermediate

layers and that this was possibly due to greater turbulence and density stratification in those layers. Of note is the fact that microplastic abundance in deep waters of the ACB (0 – 104 particles m⁻³, depths 1000 – 4400 m, sieve 250 µm) was similar to those reported for deep waters at the Rockall Trough, North East Atlantic Ocean (70.8 particles m⁻³, depth 2227 m, sieve 80 µm), (Courtenes-Jones et al. 2017). Overall, it must be acknowledged that an underestimation of microplastic abundance in the Arctic Central Basin (ACB) could have occurred in the present study as the mesh size of the sieve was only 250 µm leading to an exclusion of smaller sized particles. Nevertheless, the Arctic's remote geographic location away from major population centres, its low population in its surrounding continental shelves and relatively low in-situ anthropogenic activities (e.g. shipping) are all factors which may explain the lower microplastic abundances in sub-surface waters within the Arctic Central Basin (ACB). From an oceanographic perspective, the reduced contribution of Atlantic water in its upper water layers due to the dominance of the polar mixed layer is another possible explanation for lower microplastic abundances in the Arctic in comparison to other oceanic basins. Presumably more polluted, Atlantic water which originates from the more densely populated southern latitudes has its surface advance into the Arctic Ocean hindered due to freshwater released from melting ice and other physical barriers such as the sea ice itself and the Novaya Zemlya islands (Cózar et al. 2017). However, a plausible future scenario for the Arctic in the context of a changing climate is that microplastic abundance in near-surface layers of the Arctic Ocean may increase upon melting of contaminated sea ice and opening up of shipping lanes due to a decrease in sea ice extent (Obbard et al. 2014; Cózar et al. 2017).

Of interest is the fact that the present study managed to sample microplastics in sub-surface waters at approximately 8.5 m depth by two independent methods i.e. by the bow water system of the

vessel (underway sampling) and the rosette water sampler (CTD sampling). CTD samples (n = 9) retrieved from an average depth of 8.5 m indicated that microplastic abundance in the ACB ranged between 0 – 148 particles m⁻³, with a median of 20.8 particles m⁻³. By comparison, samples collected via the underway system (n = 58) at 8.5 m indicated that microplastic abundance in the ACB ranged between 0 – 7.5 particles m⁻³, with a median of 0.7 particles m⁻³. Although both methods sampled water at an average depth of 8.5 m, calculated microplastic abundances from both methods are not directly comparable due to the differences associated with the methods. Whereas underway sampling involved filtration of a greater volume of water (approximately 2000 L) over a longer distance and a longer sampling time (> 2 hours), the CTD sampling involved the collection and subsequent filtration of a smaller volume of water (21 L or 48 L) at a single location in a shorter period of time (minutes). The advantage of using the underway system is that microplastic abundances over a larger spatial area can be quantified whilst the vessel is in transit. By comparison, CTD sampling facilitates the quantification of microplastic abundance at specific locations making it less likely to mask contamination hotspots. However, some of the major limitations associated with CTD microplastic sampling are (i) the vessel must stop at sampling stations to collect samples, (ii) deployment and retrieval of the rosette water sampler is time consuming and, (iii) only small volumes of water can be collected in comparison to the underway sampling. The limitation of filtering smaller volumes of water is twofold in that there can be (i) false negatives whereby microplastics are not sampled despite being present in the environment or, (ii) microplastics are found in the samples but scaling up to relevant units (particles m⁻³) has a greater effect on microplastic abundances.

One of the major challenges that investigators face when quantifying marine microplastic abundance is sample contamination. In addition to employing strict measures to control contamination during sampling and processing, it is important that checks are carried out to quantify potential contamination of samples. For underway samples in the present study, although method blanks were free of contamination by synthetic particles, a single synthetic fibre was found in one air contamination check. In context, there were between 0 – 15 synthetic particles in each underway sample, with an average of 2 synthetic particles per sample. For CTD samples in the present study, between 0 – 3 synthetic fibres were found in the method blanks. In context, between 0 – 18 synthetic particles were found per CTD sample (21 L or 48 L), with an average of 2 particles per sample. In both cases, if contamination were an issue, its contribution to the reported microplastic abundances in the present study would be substantial. However, the possibility of airborne contamination in the actual underway samples is projected to be low since (i) 92 % of the air contamination checks (11 of the 12 petri dishes) were free of synthetic particles and, (ii) air contamination checks had maximum exposure to the atmosphere while actual samples had minimal exposure. With respect to the CTD samples, it is proposed that synthetic fibres in the method blanks may have been introduced into the Niskin bottle during the transfer of Milli-Q water or could have been present from the previous CTD cast and remained in the bottle due to insufficient rinsing with Milli-Q water prior to the blank. This should not have been an issue for the actual samples since Niskin bottles were rinsed during the downcast and were closed within the water column thus preventing the possibility of airborne contamination.

5.0 Conclusion

The present study demonstrated the pervasiveness of microplastics in sub-surface waters of the Arctic Central Basin (ACB). Two independent sampling techniques led to the discovery of microplastics in near surface waters of the Polar Mixed Layer (PML) i.e. at a single depth of 8.5 m as well as throughout the water column i.e. at multiple depths (8 – 4369 m) of the ACB. Such findings confirm that microplastics are entering the central Arctic Ocean, that they are being vertically transported out of surface waters and that the water column is one of the reservoirs of microplastics in this region. Presently, however, uncertainty exists regarding the actual mechanisms responsible for the vertical transport of microplastics in the Arctic Ocean. Although there was a predominance of fibrous microplastics, the majority of which were polyester, the exact sources of microplastics to the Arctic Ocean remain unknown as they could have been introduced to the ecosystem via long range transport processes or originated from more local sources. The fact that the highest microplastic abundances were recorded in the PML nearer to the periphery of the ACB suggests the influence of location-specific factors e.g. absence of sea ice, proximity to microplastic sources, wind, etc. Knowledge about microplastic abundance, distribution and composition in the Arctic Ocean is vital as it provides (i) quantitative data on the concentrations and types of microplastics that polar organisms are exposed to, (ii) a sound starting point for investigating the potential threat that microplastics pose to the Arctic ecosystem and, (iii) insight into the whereabouts of the ‘missing plastic’ from oceanic surface waters.

Acknowledgements

The authors acknowledge the invaluable support of the staff of the Swedish Polar Research Secretariat (especially Jeanette Axelsson, Robert Holden, Lars Lehnert, Asa Lindgren, Axel Meiton and Per Salo) and the crew of icebreaker Oden with the Arctic Ocean 2016 expedition. The

expert guidance of Mr. Andrew Tonkin (Plymouth University) during FT-IR analyses is also acknowledged. The first author also acknowledges the support of fellow early career scientists (especially Hans-Martin Heyn, Luz Maria Ramirez, Grace Shephard), other researchers (especially Asa Johannisson, Sofi Jonsson, Michelle Nerentorp, Carina Johansson) and staff (especially Bengt Berglund, Ted Juliusen) during the expedition.

Funding

Sampling in the Arctic Ocean was funded by the Swedish Polar Research Secretariat (SPRS) under the Early Career Scientist (ECS) Programme in which the first author was a participant. This work was also co-funded through a MARES Grant. MARES is a Joint Doctorate programme selected under Erasmus Mundus and coordinated by Ghent University (FPA 2011-0016). The funders had no role in study design, data collection, analysis and interpretation, decision to publish, or preparation of the manuscript.

References

- Amélineau, F., Bonnet, D., Heitz, O., Mortreux, V., Harding, A.M.A., Karnovsky, N., Walkusz, W., Fort, J., Gremillet, D., 2016. Microplastic pollution in the Greenland Sea: Background levels and selective contamination of planktivorous diving seabirds. *Environmental Pollution* 219, 1131-1139.
- Andrady, A.L., 2011. Microplastics in the marine environment. *Marine Pollution Bulletin* 62, 1596-1605.
- Andrady, A.L., 2017. The plastic in microplastics: A review. *Marine Pollution Bulletin* 119, 12-22.

686 Bagaev, A., Mizyuk, A., Khatmullina, L., Isachenko, I., Chubarenko, I., 2017. Anthropogenic
687 fibres in the Baltic Sea water column: Field data, laboratory and numerical testing of their motion.
688 Science of The Total Environment 599, 560-571.

689 Bergmann, M., Klages, M., 2012. Increase of litter at the Arctic deep-sea observatory
690 HAUSGARTEN. Marine Pollution Bulletin 64, 2734-2741.

691 Bergmann, M., Sandhop, N., Schewe, I., D'Hert, D., 2016. Observations of floating anthropogenic
692 litter in the Barents Sea and Fram Strait, Arctic. Polar Biology 39, 553-560.

693 Bergmann, M., Lutz, B., Tekman, M.B., Gutow, L., 2017a. Citizen scientists reveal: Marine litter
694 pollutes Arctic beaches and affects wild life. Marine Pollution Bulletin 125, 535-540.

695 Bergmann, M., Wirzberger, V., Krumpen, T., Lorenz, C., Primpke, S., Tekman, M.B., Gerdts, G.,
696 2017b. High Quantities of Microplastic in Arctic Deep-Sea Sediments from the HAUSGARTEN
697 Observatory. Environmental Science & Technology 51, 11000-11010.

698 Besseling, E., Wegner, A., Foekema, E.M., van den Heuvel-Greve, M.J., Koelmans, A.A., 2013.
699 Effects of Microplastic on Fitness and PCB Bioaccumulation by the Lugworm *Arenicola marina*
700 (L.). Environmental Science & Technology 47, 593-600.

701 Besseling, E., Wang, B., Lurling, M., Koelmans, A.A., 2014. Nanoplastic Affects Growth of *S.*
702 *obliquus* and Reproduction of *D. magna*. Environmental Science & Technology 48, 12336-12343.

703 Bhattacharya, P., Lin, S., Turner, J.P., Ke, P.C., 2010. Physical Adsorption of Charged Plastic
704 Nanoparticles Affects Algal Photosynthesis. The Journal of Physical Chemistry C 114, 16556-
705 16561.

706 Browne, M.A., Crump, P., Niven, S.J., Teuten, E., Tonkin, A., Galloway, T., Thompson, R., 2011.
707 Accumulation of Microplastic on Shorelines Worldwide: Sources and Sinks. Environmental
708 Science & Technology 45, 9175-9179.

709 Cincinelli, A., Scopetani, C., Chelazzi, D., Lombardini, E., Martellini, T., Katsoyiannis, A., Fossi,
710 M.C., Corsolini, S., 2017. Microplastic in the surface waters of the Ross Sea (Antarctica):
711 Occurrence, distribution and characterization by FTIR. *Chemosphere* 175, 391-400.

712 Cole, M., Lindeque, P., Fileman, E., Halsband, C., Galloway, T.S., 2015. The Impact of
713 Polystyrene Microplastics on Feeding, Function and Fecundity in the Marine Copepod *Calanus*
714 *helgolandicus*. *Environmental Science & Technology* 49, 1130-1137.

715 Cole, M., Lindeque, P.K., Fileman, E., Clark, J., Lewis, C., Halsband, C., Galloway, T.S., 2016.
716 Microplastics Alter the Properties and Sinking Rates of Zooplankton Faecal Pellets.
717 *Environmental Science & Technology* 50, 3239-3246.

718 Comnea-Stancu, I.R., Wieland, K., Ramer, G., Schwaighofer, A., Lendl, B., 2017. On the
719 Identification of Rayon/Viscose as a Major Fraction of Microplastics in the Marine Environment:
720 Discrimination between Natural and Manmade Cellulosic Fibers Using Fourier Transform Infrared
721 Spectroscopy. *Applied Spectroscopy* 71, 939-950.

722 Conservation of Arctic Flora and Fauna (CAFF), 2013. Arctic Biodiversity Assessment: Status
723 and trends in Arctic biodiversity, in: Meltote, H. (Ed.). *Conservation of Arctic Flora and Fauna*
724 (CAFF), Akureyri, Iceland.

725 Conservation of Arctic Flora and Fauna (CAFF), 2017. State of the Arctic Marine Biodiversity
726 Report. Conservation of Arctic Flora and Fauna International Secretariat, Akureyri, Iceland.

727 Courteney-Jones, W., Quinn, B., Gary, S.F., Mogg, A.O.M., Narayanaswamy, B.E., 2017.
728 Microplastic pollution identified in deep-sea water and ingested by benthic invertebrates in the
729 Rockall Trough, North Atlantic Ocean. *Environmental Pollution* 231, 271-280.

730 Cózar, A., Echevarria, F., Gonzalez-Gordillo, J.I., Irigoien, X., Ubeda, B., Hernandez-Leon, S.,
731 Palma, A.T., Navarro, S., Garcia-de-Lomas, J., Ruiz, A., Fernandez-de-Puelles, M.L., Duarte,

732 C.M., 2014. Plastic debris in the open ocean. *Proceedings of the National Academy of Sciences*
733 111, 10239-10244.

734 Cózar, A., Marti, E., Duarte, C.M., Garcia-de-Lomas, J., van Sebille, E., Ballatore, T.J., Eguiluz,
735 V.M., Gonzalez-Gordillo, J.I., Pedrotti, M.L., Echevarria, F., Trouble, R., Irigoien, X., 2017. The
736 Arctic Ocean as a dead end for floating plastics in the North Atlantic branch of the Thermohaline
737 Circulation. *Science Advances* 3:e1600582.

738 Desforges, J.-P.W., Galbraith, M., Dangerfield, N., Ross, P.S., 2014. Widespread distribution of
739 microplastics in subsurface seawater in the NE Pacific Ocean. *Marine Pollution Bulletin* 79, 94-
740 99.

741 Dris, R., Gasperi, J., Saad, M., Mirande, C.c., Tassin, B., 2016. Synthetic fibers in atmospheric
742 fallout: A source of microplastics in the environment? *Marine Pollution Bulletin* 104, 290-293.

743 Enders, K., Lenz, R., Stedmon, C.A., Nielsen, T.G., 2015. Abundance, size and polymer
744 composition of marine microplastics < 10 µm in the Atlantic Ocean and their modelled vertical
745 distribution. *Marine Pollution Bulletin* 100, 70-81.

746 Engel, A., Piontek, J., Metfies, K., Endres, S., Sprong, P., Peeken, I., Gabler-Schwarz, S., Nothig,
747 E.-M., 2017. Inter-annual variability of transparent exopolymer particles in the Arctic Ocean
748 reveals high sensitivity to ecosystem changes. *Scientific Reports* 7, 4129.

749 Eriksen, M., Lebreton, L.C.M., Carson, H.S., Thiel, M., Moore, C.J., Borerro, J.C., Galgani, F.,
750 Ryan, P.G., Reisser, J., 2014. Plastic Pollution in the World's Oceans: More than 5 Trillion Plastic
751 Pieces Weighing over 250,000 Tons Afloat at Sea. *PLoS ONE* 9, e111913.

752 Fazey, F.M.C., Ryan, P.G., 2016. Biofouling on buoyant marine plastics: An experimental study
753 into the effect of size on surface longevity. *Environmental Pollution* 210, 354-360.

754 Isobe, A., Uchiyama-Matsumoto, K., Uchida, K., Tokai, T., 2017. Microplastics in the Southern
755 Ocean. *Marine Pollution Bulletin* 114, 623-626.

756 Jakobsson., M., Grantz, A., Kristoffersen, Y., Macnab, R., MacDonald, R.W., Sakshaug, E., Stein,
757 R., Jokat, W., 2004. The Arctic Ocean: Boundary Conditions and Background Information, in:
758 Stein, R., MacDonald, R.W. (Eds.), *The Organic Carbon Cycle in the Arctic Ocean*. Springer,
759 Berlin.

760 Kanhai, L.D.K., Officer, R., Lyashevskaya, O., Thompson, R.C., O'Connor, I., 2017. Microplastic
761 abundance, distribution and composition along a latitudinal gradient in the Atlantic Ocean. *Marine*
762 *Pollution Bulletin* 115, 307-314.

763 Kuhn, S., Schaafsma, F.L., van Werven, B., Flores, H., Bergmann, M., Egelkraut-Holtus, M.,
764 Tekman, M.B., van Franeker, J.A., 2018. Plastic ingestion by juvenile polar cod (*Boreogadus*
765 *saida*) in the Arctic Ocean. *Polar Biology*, 1-10.

766 Kukulka, T., Proskurowski, G., Morét-Ferguson, S., Meyer, D.W., Law, K.L.C.L., 2012. The
767 effect of wind mixing on the vertical distribution of buoyant plastic debris. *Geophysical Research*
768 *Letters* 39.

769 Long, M., Moriceau, B.I., Gallinari, M., Lambert, C., Huvet, A., Raffray, J., Soudant, P., 2015.
770 Interactions between microplastics and phytoplankton aggregates: Impact on their respective fates.
771 *Marine Chemistry* 175, 39-46.

772 Lusher, A.L., Burke, A., O'Connor, I., Officer, R., 2014. Microplastic pollution in the Northeast
773 Atlantic Ocean: Validated and opportunistic sampling. *Marine Pollution Bulletin* 88, 325-333.

774 Lusher, A.L., Tirelli, V., O'Connor, I., Officer, R., 2015. Microplastics in Arctic polar waters: the
775 first reported values of particles in surface and sub-surface samples. *Scientific Reports* 5, 1-9.

776 Mallory, M.L., 2008. Marine plastic debris in northern fulmars from the Canadian high Arctic.
 777 Marine Pollution Bulletin 56, 1501-1504.

778 Napper, I.E., Thompson, R.C., 2016. Release of synthetic microplastic plastic fibres from domestic
 779 washing machines: Effects of fabric type and washing conditions. Marine Pollution Bulletin 112,
 780 39-45.

781 Obbard, R.W., Sadri, S., Wong, Y.Q., Khitun, A.A., Baker, I., Thompson, R.C.C.E.F., 2014.
 782 Global warming releases microplastic legacy frozen in Arctic Sea ice. Earth's Future 2, 315-320.

783 Oliveira, M., Ribeiro, A., Hylland, K., Guilhermino, L., 2013. Single and combined effects of
 784 microplastics and pyrene on juveniles (0+ group) of the common goby *Pomatoschistus microps*
 785 (Teleostei, Gobiidae). Ecological Indicators 34, 641-647.

786 Phuong, N.N., Zalouk-Vergnoux, A., Poirier, L., Kamari, A., Chatel, A., Mouneyrac, C., Lagarde,
 787 F., 2016. Is there any consistency between the microplastics found in the field and those used in
 788 laboratory experiments? Environmental Pollution 211, 111-123.

789 Provencher, J.F., Gaston, A.J., Mallory, M.L., O'hara, P.D., Gilchrist, H.G., 2012. Ingested plastic
 790 in a diving seabird, the thick-billed murre (*Uria lomvia*), in the eastern Canadian Arctic. Marine
 791 Pollution Bulletin 60, 1406-1411.

792 R Core Team, 2015. R: A Language and Environment for Statistical Computing. R Foundation for
 793 Statistical Computing Vienna, Austria.

794 Rochman, C.M., Hoh, E., Kurobe, T., Teh, S.J., 2013. Ingested plastic transfers hazardous
 795 chemicals to fish and induces hepatic stress. Scientific Reports 3, 3263.

796 Rochman, C.M., Kurobe, T., Flores, I., Teh, S.J., 2014. Early warning signs of endocrine
 797 disruption in adult fish from the ingestion of polyethylene with and without sorbed chemical
 798 pollutants from the marine environment. Science of The Total Environment 493, 656-661.

799 Rudels, B., Larsson, A.-M., Sehlstedt, P.-I., 1991. Stratification and water mass formation in the
800 Arctic Ocean: some implications for the nutrient distribution. *Polar Research* 10, 19-32.

801 Rudels, B., 2015. Arctic Ocean circulation, processes and water masses: A description of
802 observations and ideas with focus on the period prior to the International Polar Year 2007 - 2009.
803 *Progress in Oceanography* 132, 22-67.

804 Tekman, M.B., Krumpen, T., Bergmann, M., 2017. Marine litter on deep Arctic seafloor continues
805 to increase and spreads to the North at the HAUSGARTEN observatory. *Deep Sea Research Part*
806 *I: Oceanographic Research Papers* 120, 88-99.

807 Trevail, A.M., Gabrielsen, G.W., Kuhn, S., Van Franeker, J.A., 2015. Elevated levels of ingested
808 plastic in a high Arctic seabird, the northern fulmar (*Fulmarus glacialis*). *Polar Biology* 38, 975-
809 981.

810 United Nations Environment Programme (UNEP), 2011. *UNEP Year Book: Emerging issues in*
811 *our global environment*. United Nations Environment Programme (UNEP), Nairobi, Kenya.

812 van Sebille, E., England, M.H., Froyland, G., 2012. Origin, dynamics and evolution of ocean
813 garbage patches from observed surface drifters *Environmental Research Letters* 7, 1-6.

814 von Moos, N., Burkhardt-Holm, P., Kohler, A., 2012. Uptake and Effects of Microplastics on Cells
815 and Tissue of the Blue Mussel *Mytilus edulis* L. after an Experimental Exposure. *Environmental*
816 *Science & Technology* 46, 11327-11335.

817 Waller, C.L., Griffiths, H.J., Waluda, C.M., Thorpe, S.E., Loaiza, I., Moreno, B., Pacherrres, C.O.,
818 Hughes, K.A., 2017. Microplastics in the Antarctic marine system: An emerging area of research.
819 *Science of The Total Environment* 598, 220-227.

820 Wegner, A., Besseling, E., Foekema, E.M., Kamermans, P., Koelmans, A.A., 2012. Effects of
821 Nanopolystyrene on the Feeding Behaviour of the Blue mussel (*Mytilus Edulis* L.). Environmental
822 Toxicology and Chemistry 31, 2490-2497.

823 Wright, S.L., Rowe, D., Thompson, R.C., Galloway, T.S., 2013. Microplastic ingestion decreases
824 energy reserves in marine worms. Current Biology 23, R1031-R1033.

825 Zarfl, C., Matthies, M., 2010. Are marine plastic particles transport vectors for organic pollutants
826 to the Arctic? Marine Pollution Bulletin 60, 1810-1814.

827

828 **List of Tables**

829 Table 1: Microplastic abundance in the various water layers of the Arctic Central Basin (ACB)

830

831 **List of Figures**

832 Figure 1: General overview of the bathymetry and water masses of the Arctic Central Basin
833 [reprinted here with permission from CAFF], (CAFF 2013)

834 Figure 2: Microplastic abundance in the Arctic Central Basin (a) based on bow water sampling at
835 a single depth of 8.5 m, (b) based on CTD sampling at multiple depths (8 – 4369 m)

836 Figure 3: Colours of microplastics found in (a) underway and (b) CTD samples at depths 8 – 4369
837 m

838 Figure 4: Size classes of microplastics found in (a) underway and (b) CTD samples at depths 8 –
839 4369 m

840 Figure 5: Synthetic polymers found in (a) underway and (b) CTD samples at depths 8 – 4369 m

841 Figure 6: Microplastic abundance from the various CTD casts in the Arctic Ocean

842

843 **List of Supplementary Tables**

844 Supplementary Table 1: Descriptive statistics, tests of normality and correlation findings for
845 environmental variables (underway data)

846 Supplementary Table 2: Descriptive statistics, tests of normality and correlation findings for
847 environmental variables (CTD data)

848 Supplementary Table 3: Quality control checks associated with the underway sampling

849 Supplementary Table 4: Quality control checks associated with CTD sampling

850 Supplementary Table 5: Number of microplastics and calculated microplastic abundances for the
851 underway samples

852 Supplementary Table 6: Number of microplastics and calculated microplastic abundances for
853 each CTD cast

854 Supplementary Table 7: Microplastic abundances reported for oceanic basins across the world

855